International Plant and Animal Genome XXV. january 2017

A Reference Sequence of the Monoploid Genome of Sugarcane

Olivier GARSMEUR CIRAD, Montpellier, France









Domestication S. officingrum

Highly polyploid

Aneuploid Interspecific

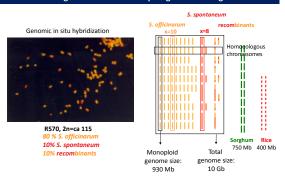
Modern sugarcane cultivars

2n ~ 110-130

S. robustum

2n = 60,80-->200

Global organization of the complex genome of sugarcane

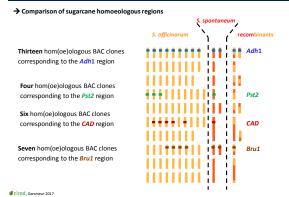


(D'Hont et al, 1996; D'Hont et al, 1998; D'Hont 2005, Piperidis et al 2010)

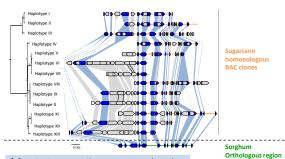
Fine structure and evolution of the sugarcane genome

//*XXXXXX*//

modern cultivars



Analysis of sugarcane homeologs and comparison with sorghum



→ Gene structure conservation among sugarcane homeologs

- → All hom(oe)o-alleles predicted functional
- → High colinearity with sorghum

₫ cirad, Garsmeur 2017

→Transposable elements variations

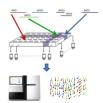
Garsmeur et al. New Phytologist 2010 Charron et al. submitted

DHONT PAG 2015

Sugarcane sequencing strategy - Genes are conserved among sugarcane hom(oe)ologous chromosomes → if we could sequence a set of BACs representing one monoploid genome, it would represents a very useful reference sequence - High colinearity between sugarcane and sorghum →We should be able to use sorghum to identify a core set of sugarcane BACs representing one monoploid genome →Mainly the genes are conserved → we focus on the gene-rich part of the genome

BAC selection through Whole Genome Profiling (WGP) Technology

WGP technology generates short sequence tags from the terminal ends of restriction fragments from pooled BACs



ALBERT HELD

(A) 20,736 BAC clones from R570 Sugarcane BAC library (~ 2x coverage of the monoploid genome)

(B) BAC pooling, DNA extraction and restriction

(C) Sequencing of terminal ends of restricted fragments

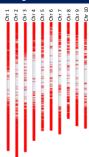
(D) Decovolution using barcodes

→ Around 30 to 50 sequence tags per BAC are produced

→ Anchoring of the produced WGP-tags onto the sorghum genome

∉ cirαd, Garsmeur 2017

Distribution of the sugarcane BAC in the sorghum genome



→11,732 R570 sugarcane BACs anchored onto the sorghum

₡ cirad, Garsmeur 2017

Sugarcane BACs anchor in sorghum gene-rich regions

Sorghum Chromosome 1

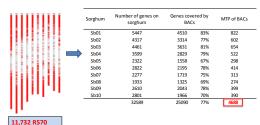


ightarrow BAC mostly distributed in gene-rich regions

DHONT PAG 2015

Minimum tiling path (MTP) of sugarcane BACs

MTP = minimum set of BACs to be sequenced to obtain the best coverage of chromosomes.



sugarcane BACs anchored onto the sorghum → MTP ~4,700 BACs to cover the basic sugarcane genome

∉ Cirod, Garsmeur 2017

Sequencing the MTP of R570 BAC

BAC sequenced through international collaboration

BAC sequenced using PacBio RSII technology and 100X depth coverage



97.5% of BAC assembled in less than 3 contigs

ICSB









The monoploid sugarcane reference sequence

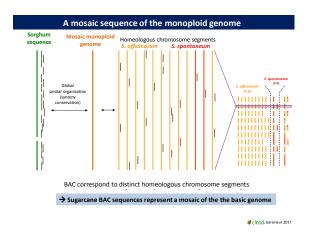
 ${\bf \mbox{$\mbox{}\mbox{$\mbox{\mbo

high quality sequence

 $\hfill \square$ Cover ~80 % of the sorghum genes

₫ cirad, Garsmeur 2017

Sugarcane BAC (syntenic path) Sugarcane BAC (syntenic path) Sorghum genome © cirid. Garaneeur 2017



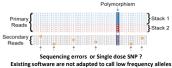
Anchoring BAC sequences onto sugarcane chromosomes

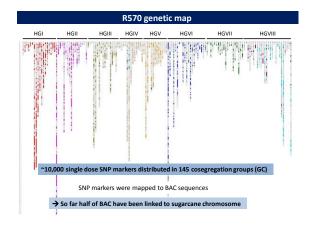
Sugarcane BAC were selected based on synteny conservation with sorghum. It's now essential to anchor them on sugarcane chromosomes

→ Development of a R570 high density genetic map

- Genotyping By Sequencing (Reduced-complexity method)
 - -> Avoid having to genotype the entire genome
 - -> Target and enrich specific loci to ensure sufficient sequence coverage
 - -> Produce thousands of SNP markers
- GBS of a mapping population (94 individuals)
- Development of bioinformatics tools for identifying single dose SNP that are useful for genetic mapping $\,$







BAC sequences annotation

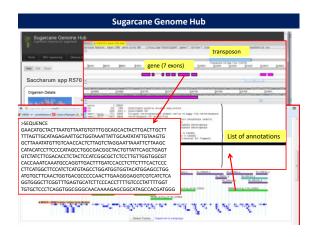
 $\label{lem:continuous} \textbf{Annotation of the BAC is in progress using a pipeline dedicated to sugarcane BAC annotation}$

☑ Predict location, structure and putative function of genes ☑ Has been successfully used to annotate sugarcane BAC sequence



Exploiting R570 transcriptomic resources to improve annotations

Sample	No of raw reads	No of filtered reads	% of reads after filtration
Leaf	64 146 407	62 951 164	98.14
Root	55 065 789	53 976 765	98.02
Stem	57 822 126	56 758 545	98.16
TOTAL	177 034 322	173 686 474	98.11



JGI ibei

This sugarcane sequence will represent an essencial reference

- → To serve as template to align Genotyping By Sequencing data from any variety
 → for association and QTL studies, genomic selection using GBS markers
- → To serve as template to align RNAseq data from any variety → to study gene expression in specific conditions
- → To finely map or clone genes of interest → for marker-assisted selection
- ightarrow To serve as high quality framework to help assemble the whole genome sequence

cirad Jeremy Schmutz Angélique D'Hont Jane Grimwood Guillaume martin Blake Simmons Karen Aitken Carine Charron Paul Berkman Catherine Hervouet **√**KeyGene Gaetan Droc **QAAFI** Stéphanie Bocs Edwin van der Vossen Rudie Antonise Robert Henry SOUTH AFRICA SNA CHPGU LZJ Bernard Potier Derek Watt Marie-Anne Van Sluys Hélène Bergès **International Consortium for Sugarcane Biotechnology** CHARCA, EEAOC, SRA,CTC, CENICANA,CINCAE,CEGICANA, VSI, MSIRI, SASRI, Mitr Phol, FSCL, HARC, ASCL, RGVSG

AUSTRALIA

FRANCE

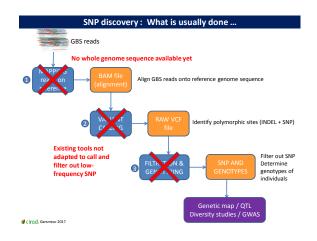
SUPP

Genetic mapping in polyploids is challenging ... The first difficulty resides in the identification of single dose SNP markers that are the one used for genetic mapping. Homoeologous chromosomes AAAT AAAAAAAA Reads Secondary Sequencing errors or Single dose SNP? Sequencing errors or Single dose SNP?

The 2nd difficulty is that existing softwares/methods such as TASSEL/GATK+SAMtools or denovo SNP pipelines such as /UNEAK/STACKS do not well manage with genotyping in polyploids, particularly with highly polyploid and heterozygous genomes

Need to develop and test analytic method adapted to complex genomes to identify high quality single dose SNP markers

€ Cirad, Garsmeur 2017



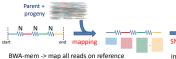
Development of bioinformatics tools for GBS analyses in polyploids

STEP1 = Build a reference pseudo contig from parental GBS reads



CD-HIT-DUP -> remove redondancy within the parental reads
CD-HIT-EST -> CRISTENTIAL OF CHIQUE PERCENTIAL STATES AND THE STATES

STEP2 = Mapping of all reads onto the reference and SNP calling Process_reseq.py



in-house program : count alleles observed at each site

₡ cirad, Garsmeur 2016

→ Identification of Single dose SNP

Developement of bioinformatic tools to identify simplex markers

Sequencing depth cutoff:

30X min
1000X max

Coverage of minor allele observed:

minor allele 2 X

=> 10% missing data max in progeny

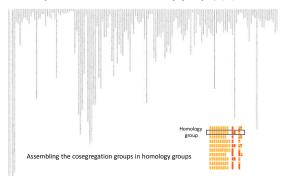
Select markers based on ChiSquare test
-pValue: 0.05
=> analyze the segregation of marker

=> analyze the segregation of mark

entification of Single dose SNP

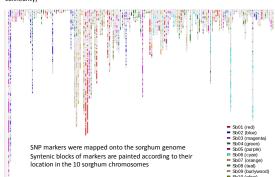
R570 Sugarcane Genetic Map

~10,000 single dose SNP markers distributed in 145 cosegregation groups (GC)

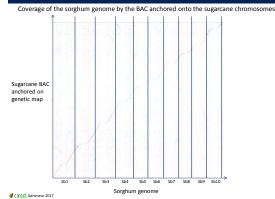


Syntenic blocks between R570 genetic map and Sorghum

Homoelogous sugarcane chromosomes would correspond to same sorghum regions (global colinearity)



Anchoring BAC sequences onto the sugarcane chromosomes



Conclusions

- ☐ We developed a sequencing strategy to produce a reference sequence corresponding to the gene-rich regions of the basic (monoploid) sugarcane genome
- ☐ We have identified and sequenced a set of 5000 BAC covering the gene-rich part of the 10 basic sugarcane chromosomes
- We tested GBS and developed bioinformatics tools able to discover single dose SNP markers in complex polyploid genomes
- ☐ We built a genetic map comprising ~10,000 SNP markers and use it to anchor around half of the BAC
- → Need to increase the number of marker: GBS on selfed progeny from R570 ?
- → Identifing SNP on BAC : Targeted sequence capture on BAC sequence ?

This reference sequence will be a very useful resource for genetics (GWAS, GS) and genomics studies in sugarcane (WGD, expression, ..)

It will also represents an essential high quality frame to help building a whole genome sequence