



# Progress to Control and Eradication of Peste des Petits Ruminants in the Southern African Development Community Region

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In southern Africa, small ruminants are an important source of nutrition and income to resource-poor small holder farmers. After spreading from West to Central and Eastern Africa, peste des petits ruminants (PPR) emerged in the United Republic of Tanzania in 2008 and has since been reported in Angola, the Democratic Republic of the Congo, and the Comoros. The disease can cause considerable morbidity and mortality in naïve sheep and goat populations and severely impact rural livelihoods, particularly those of women. Gaps in the knowledge of PPR epidemiology still exist, particularly around the role of small-ruminant movement and the role of the abundant wildlife in southern Africa. The capacity of veterinary services to undertake surveillance and control PPR is heterogeneous within the region, with vaccination being limited. The Pan African strategy for the control and eradication of PPR mirrors the Global Strategy and provides the framework for the Southern African Development Community (SADC) region to meet the 2030 goal of eradication. Five countries and one zone within Namibia are officially PPR free according to OIE Standards. Most countries have developed national strategies for the control and eradication of PPR. To strengthen national and regional PPR eradication programme goals, there is a need for a regional risk-based surveillance adapted to infected, high-risk and lower-risk countries that will enable targeted and efficient control, rapid response to incursions and prevention of spread as well as improved preparedness. Continued international and national support will be necessary including laboratory diagnostics and enhancing surveillance capacity to prevent further spread southwards on the continent.

**Keywords:** peste des petits ruminants, Southern African Development Community, surveillance, risk-based approaches, small ruminants

## INTRODUCTION

Peste des petits ruminants (PPR) is a World Organization for Animal Health (OIE) listed disease (1) caused by a morbillivirus resulting in variable respiratory and enteritis associated clinical disease in sheep and goat populations. PPR can also infect cattle, camels, domestic buffaloes, and wild ruminants (2). Given the high morbidity and mortality of PPR infection in immune-naïve small ruminants, the economic and food security impact of outbreaks is large for small-holder farmers. Women's livelihoods and resilience are particularly affected by PPR as women predominately rear small ruminants primarily for income generation and food security (3). The annual cost

of PPR-associated sheep and goat deaths for worldwide infected countries is estimated between 794 million and 2.7 billion US dollars (4). This contagious viral disease has steadily expanded its geographical distribution from West into Eastern Africa and more recently to the Southern African Development Community (SADC) countries. Given the porous nature of country borders and movement of animals in many African countries, the risk of spread is high for countries bordering PPR infected ones.

Following the successful eradication of rinderpest globally in 2011, the Food and Agriculture Organization of the United Nations (FAO) and the OIE developed the Global Strategy for the Control and Eradication (GSCE) of PPR (5) to enable this plague to be the next eradicated animal disease by 2030. The control and eventual eradication of PPR will contribute significantly to achieving the elimination of poverty [Sustainable Development Goals (SDG 1)] and the end of hunger and malnutrition (SDG2) as well as contributing to other SDGs (3, 5, 8, 11, 12 and 17) (6). The global strategy was endorsed by 45 African Countries and the African Union Inter-African Bureau for Animal Resources (AU-IBAR) voiced its support for the global programme (7). The SADC region had already developed its own PPR control strategy (8). The global conference on “*Partnering and Investing for a Peste des Petits Ruminants Free World*” organized by the OIE and FAO in 2018, hosted by the European Commission was to reaffirm the political will of countries and to mobilize resources (6) to meet the 2030 eradication goal.

With the southern spread of this disease into the SADC region and issues associated with differentiating PPR from other diseases (9), national and regional approaches are urgently needed. SADC is the only region in sub-Saharan Africa with non-infected countries and therefore plays an important role in facilitating the control and eradication of PPR in infected countries which will in turn reduce the risk of disease spread further south on the African continent. An overview of the current situation is presented in this paper and the main constraints and opportunities to control PPR in the SADC regions are discussed.

## SITUATION ANALYSIS

### Current PPR Status in SADC

In southern Africa, PPR has spread into new areas in recent years (Figure 1A). Tanzania was first infected probably from imported animals from Kenya in 2008 and represents an important potential source of PPR viruses for the rest of the region (10). PPR is now considered endemic in Tanzania in small ruminants with PPR lineages II, III, and IV circulating (11). The disease has spread from Tanzania to the Democratic Republic of Congo (DRC) and Comoros (12, 13). Around 2012, Angola was infected probably with imported animals from DRC but these outbreaks have not been officially recorded (14). So far, no clinical disease has been reported in Namibia, Malawi, Mozambique, or Zambia (4). Zambia did detect PPR seropositive goats in recent years, though in the absence of clinical disease, suggesting either that antibodies were from imported vaccinated animals or previously infected (i.e., from Tanzania and/or DRC) or false positives (Bedane personal communication, roadmap meeting). The situation in Mozambique is similar

(15). The borders between Tanzania, DRC, and Angola and neighboring non-infected countries represent important entry gates for PPR into the rest of southern Africa. Namibia (Northern Communal Area), Malawi, Mozambique, and Zambia are therefore considered at high-risk of PPR infection. Botswana, Eswantini, Mauritius and South Africa and the southern zone of Namibia, are declared by the OIE as PPR free. Lesotho and Zimbabwe are also considered at lower-risk of PPR infection (Figure 1B). Madagascar and Seychelles could be considered at risk because of the maritime trade of small ruminants with Comoros (12). However, Madagascar has been declared PPR free by the OIE in 2018, where now efforts for surveillance need to be strengthened to avoid reversal of this status.

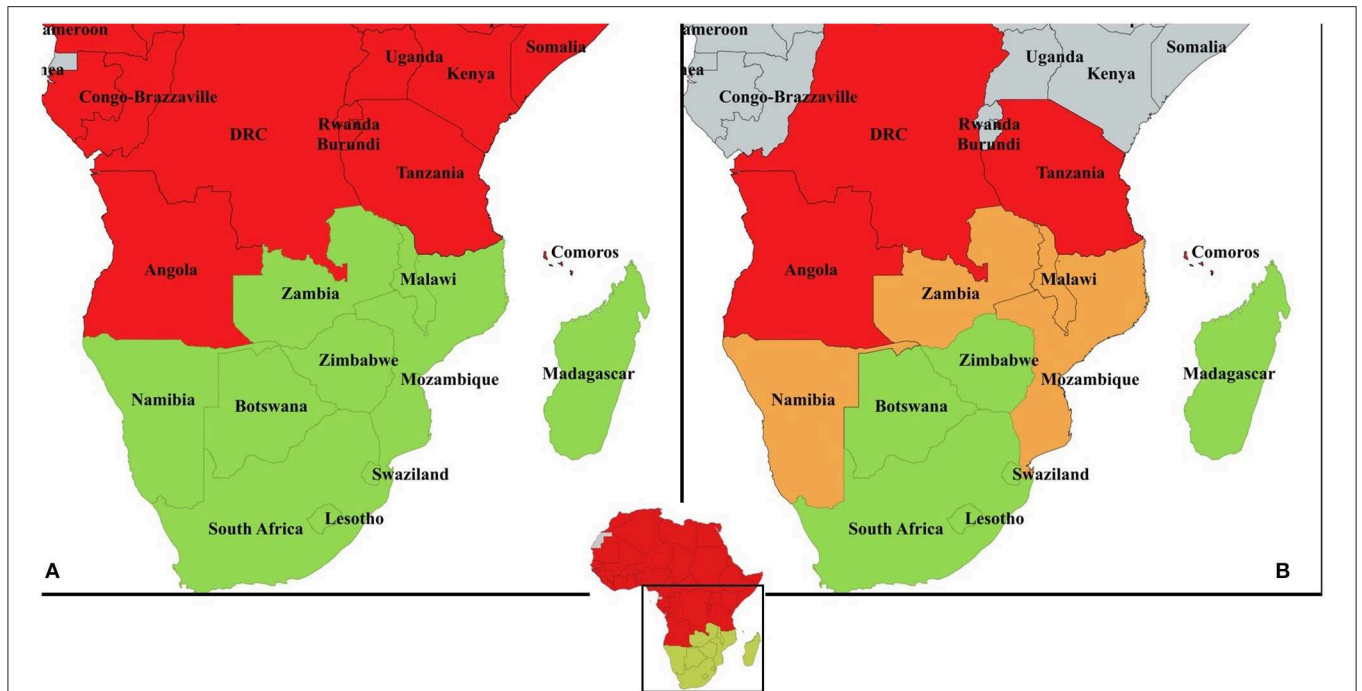
### Epidemiology of PPR in Southern Africa

PPR appears a good candidate for eradication according to criteria for eradication (16). However, the large populations of sheep and goats and their high population turnover (annual turnover rates of up to 30%) necessitate a higher effort (and costs) for control (14). The epidemiology of PPR is relatively well-defined but gaps in knowledge still exist and variability between regions may occur.

Firstly, following initial exposure of small-ruminant populations to PPR, a high mortality and morbidity is expected, which provides a visible clinical picture detectable by passive surveillance systems. However, in African small-scale farming systems, co-infection by multiple pathogens is frequent and could blur the expected clinical picture (17). PPR is also known to be a seasonal disease in some African endemic regions with peak infections usually occurring during the cool, dry season (18). This season in southern Africa starts in April and extends to August in some areas, providing a long environmental window for PPRV transmission. Under these climatic conditions, little is known about the PPRV persistence in urine and feces (14).

Secondly, as in other African regions, the SADC region hosts countries with large small-ruminant populations (e.g., South Africa) and many are still free of PPR unlike all other African countries. Long-range cross-border trade of small ruminants involving two or more countries is frequent in the region (19). The patterns of this trade are largely unknown despite their relevance for the introduction and spread of animal diseases. Short-range trade (involving adjacent districts in neighboring countries), considered also to be illegal, represents an important socio-cultural component of local livelihoods. In addition, climate change is expected to increase extreme climatic events including droughts (20). Those events will affect mainly the poorest populations depending mostly on small ruminant production and living in the most arid areas. Droughts and political instability have been shown already to play their role in PPR spread (21).

Finally, some questions still remain on the host range of PPRV and their role in the local PPR epidemiology (22). In particular, little is known regarding virus excretion in infected camels, cattle, and wildlife (23, 24). In West Africa, cattle seem to be a dead-end host for PPR (25) but the role of local southern African breeds could be different (e.g., these breeds experienced different selective pressures by the rinderpest virus). In Africa, the role



**FIGURE 1 | (A)** PPR country data compiled from official reports and literature between 2008 and 2018. In red, countries with at least one occurrence of disease reported, in green countries with absence of disease or disease never been reported. Angola is considered infected in at least one zone by multiple references (see text) despite no OIE report of clinical disease. Zambia has reported seropositivity in 2015 to OIE but subsequent surveillance failed to prove occurrence of disease; **(B)** suggested risk-based approach for PPR surveillance and control in the Southern African Development Community: in red, “infected countries” with presence of the disease in at least one zone; in orange, “high-risk countries” sharing a border with an infected country; in green, “lower-risk countries” with no border shared with any infected country. Madagascar and Seychelles could be considered at risk because of the maritime trade of small ruminants with Comoros. However, the scale of this trade in intensity and frequency is not quantified. Mauritius is considered lower-risk country given its assumed low level of maritime trade with Comoros.

of most wild ungulate species in PPR epidemiology is largely unknown, as no clinical disease has ever been reported despite exposure (26). Clinical disease has been observed in African ungulates in zoo environments elsewhere (27) and in other wild ungulate species in central Asia (28, 29). The southern African region has large and healthy wildlife populations with relative freedom of movements across borders thanks to the creation of Transfrontier Conservation Areas (TFCAs) (30, 31). In addition, several species are endemic to the region (e.g., springbok) and some countries such as South Africa and Namibia have developed an important wildlife industry where animals maybe bred in conditions in-between natural and zoo settings where they could become particularly susceptible to PPR.

## Regional Capacity for PPR Surveillance and Control

The SADC Secretariat has identified PPR as one of the three major Transboundary Animal Diseases affecting regional and international trade (32). The SADC strategy (8) describes the limited PPR control capacity in SADC region in relation to diagnosis and surveillance, knowledge of virus transmission and susceptible species and differentiation of infected and vaccinated animals. Legislation on the use of PPR vaccines was also noted as an issue in most countries. The Pan African Veterinary Vaccine Center (AU-PANVAC) is mandated to provide quality assurance

of all veterinary vaccines produced or imported into Africa and to coordinate the harmonization of veterinary vaccine registration with the support of the Global Alliance for Livestock Veterinary Medicines (GALVmed) and the OIE, which will be important for many SADC countries should they require PPR vaccine quickly due to an incursion.

Effective vaccination campaigns to ensure sustained herd immunity with 80 percent population coverage, will be pivotal to eradicating PPR as it was for eradicating rinderpest (14) though the high reproductive rate of small ruminants may warrant the need for annual vaccination in some flocks. Vaccination of small ruminants is limited in some areas due to the cost of vaccines, delivery and access to animals. Current vaccines against PPR virus are homologous vaccines (33) and require only one dose for life-long protection. The first vaccine was against lineage II (Nigeria 75/1) Africa PPR virus strain and has been used for 30 years. The impossibility to differentiate between vaccinated and infected animals and its thermolability are some of the limitations of this vaccine. Recent research on freeze-drying these types of live-attenuated vaccines have enabled thermostability and resistance to high temperatures in the field (34). The Botswana Veterinary Institutes (BVI) capacity in establishing and maintaining PPR-VAC<sup>®</sup> was confirmed during a recent FAO supported project. This live-attenuated vaccine has also been assessed recently using an *in-vivo* challenge model in goats (35).

343 Additionally, a thermo-adapted live-attenuated PPR vaccine has  
 344 been trialed in goats in India (36). Assessment of the cross-lineage  
 345 efficacy of different PPR vaccines is important given SADC has  
 346 several lineages circulating (37). Recently comparative studies  
 347 have indicated that the Nigeria 75/1 strain vaccines produce  
 348 stronger antibody responses than the India S96, though the  
 349 Indian strain vaccine elicits a greater cell-mediated immune  
 350 response (38).

351 Given concurrent infection of sheep and goats with PPR and  
 352 other diseases such as FMD or goat-pox (17, 39) a bivalent  
 353 vaccine or concurrent vaccination would be of benefit to livestock  
 354 owners and would be in line with the GSCE targeting other  
 355 small ruminant diseases during the eradication programme.  
 356 Development of vaccines based on Differentiating Infected from  
 357 Vaccinated Animals (DIVA) technology will assist in surveillance  
 358 and eradication (37). Unfortunately, these recombinant vaccines  
 359 require booster doses and cost more than conventional vaccines  
 360 but they have the advantage of temperature stability and their  
 361 DIVA properties.

362 The SADC member states have a network of laboratories  
 363 (provincial and national laboratories) for the surveillance of PPR  
 364 and PPR diagnostic capacity which varies between countries.  
 365 As in most national laboratories, the laboratories in Malawi,  
 366 Mozambique, Namibia, Zambia, and Zimbabwe have been  
 367 using c-ELISA assays to conduct PPR sero-surveillance in  
 368 high-risk areas to detect the presence or absence of PPRV  
 369 (Country reports, 2019). To increase the test sensitivity and  
 370 affordability, AU-PANVAC has developed a blocking (b)-ELISA  
 371 test (40). Some SADC countries have participated in the  
 372 validation trial of this test (e.g., Malawi), others have been  
 373 supplied kits (e.g., Mozambique) and others have requested  
 374 them (Bedane personal communication). Most of the national  
 375 laboratories have molecular PPR diagnostic capacity (e.g., PCR  
 376 or qPCR). The capacity to conduct virus neutralization tests—  
 377 OIE gold standard—and virus isolation and sequencing is absent  
 378 from most national laboratories in SADC. Consequently, PPR  
 379 confirmatory diagnostics of doubtful results requires countries to  
 380 send samples to OIE reference laboratories for PPR (e.g., CIRAD  
 381 or Pirbright Institute) (41) or to AU-PANVAC.

382 New field surveillance strategies may assist in the early  
 383 diagnosis of disease and provide increased sensitivity and  
 384 specificity of tests by targeting PPR virus specific antibodies,  
 385 antigens, or genetic material (41–43). The direct detection of  
 386 PPR virus genetic material and antigen in fecal samples could  
 387 be used in small ruminant and wildlife surveillance (41). A pen-  
 388 side test using quantum dots with a lateral-flow test strip has  
 389 been evaluated in the field with similar results to c-ELISA (43).  
 390 Such, a pen-side test that could confirm several small ruminant  
 391 pneumo-enteritis diseases would be useful (34).

## 392 **FAO and OIE Guidance and Support**

393 FAO and OIE have established a Global Secretariat, which  
 394 coordinates efforts for the PPR Global Eradication Programme  
 395 (GEP) (44) based on a Progressive Control Pathway (PCP). The  
 396 Global Secretariat is conducting Regional Roadmap workshops  
 397 for the PCP implementation. In southern Africa, two Regional  
 398 Roadmap meetings took place in October 2016 in Harare,  
 399

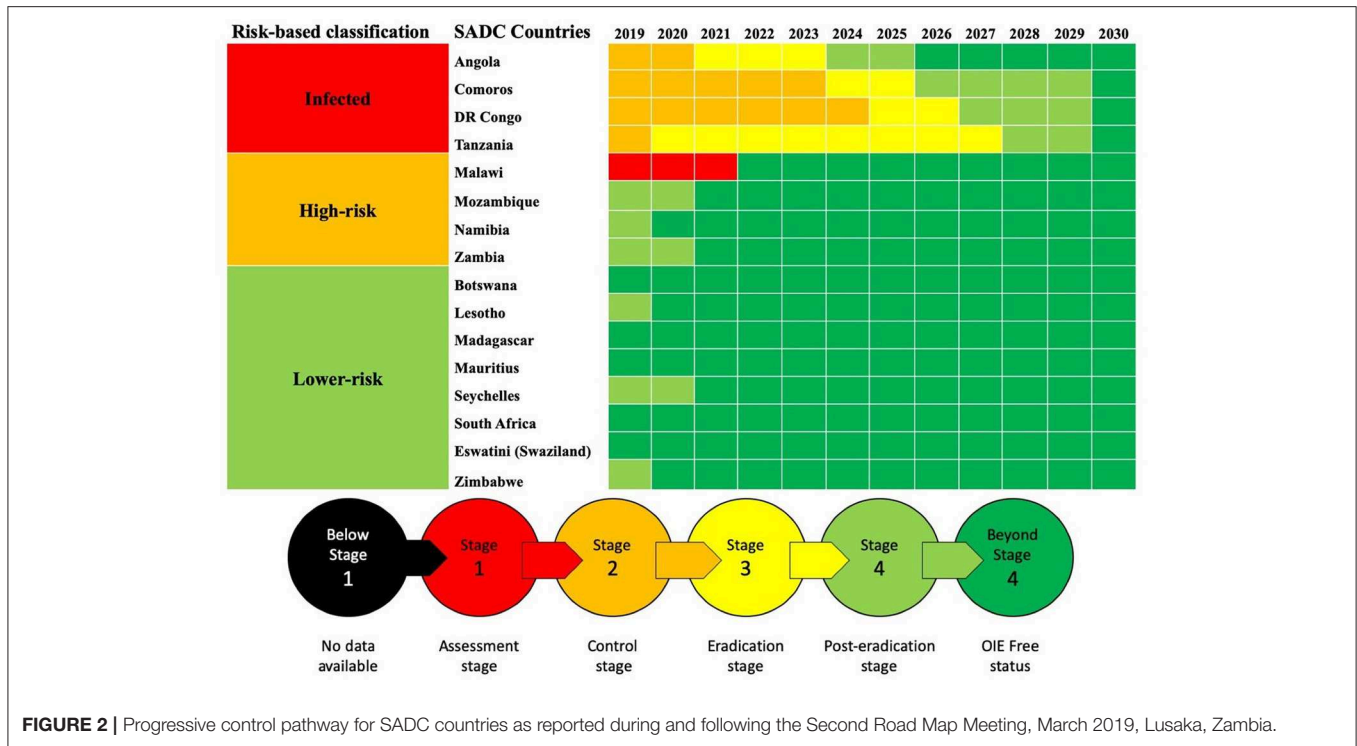
400 Zimbabwe and in March 2019 in Lusaka, Zambia (45). The  
 401 PPR Roadmap meetings ensure continuous evaluation and  
 402 monitoring of the PPR situation and help in harmonizing policies  
 403 and strategies among countries, as well as with other regions,  
 404 for the implementation of the PPR GSCE. This strategy follows  
 405 three core components advocating a risk-based approach to  
 406 disease control to better target “virus hotspots.” The progressive  
 407 stepwise approach—no available data (stage 1) to OIE free status  
 408 (stage 5)—(Figure 2) and the PPR Monitoring and Assessment  
 409 Tool (PMAT) are used in these meetings and correspond to  
 410 a combination of decreasing levels of epidemiological risk and  
 411 increasing levels of prevention and control capabilities.

412 The support provided by OIE and FAO both directly  
 413 and indirectly assists SADC countries to progress along  
 414 their respective PPR roadmap pathways (Figure 2). The  
 415 FAO has been actively building capacity to prevent PPR  
 416 introduction into Malawi, Mozambique, and Zambia through a  
 417 Technical Cooperation Project 2013–2015 involving serological  
 418 surveillance, local stakeholders awareness, and building rapid  
 419 diagnostic capacity and national contingency and preparedness  
 420 plans (TCP/SFS/3403) (46). Additional support by FAO  
 421 provided to Madagascar and Lesotho enabled the former to  
 422 obtain “Freedom from PPR certification” in 2018 while Lesotho  
 423 will soon submit the documentation for its OIE freedom  
 424 following FAO project TCP/LES/3604 (Pers. Com. Bedane).  
 425 PPR control and eradication became one of the components  
 426 of a recently launched SADC-based project financed by EU  
 427 (“Support Toward the Operationalization of the SADC Regional  
 428 Agricultural Policy”—GCP /SFS/004/EC). Additionally, OIE  
 429 Performance of Veterinary Services (PVS) tool (47) will greatly  
 430 support the assessment of the 47 Critical Competencies of  
 431 Veterinary Services in countries and of areas specific to PPR  
 432 control and eradication (7). Better control and diagnosis of other  
 433 small ruminant diseases is also necessary for improving farmer  
 434 participation. OIE has also been assisting to build PPR diagnostic  
 435 capacity through focal point training, including fifth cycle  
 436 workshop “Wildlife Health Information Management” 2018 and  
 437 laboratory twinning projects between reference laboratories and  
 438 SADC laboratories (e.g., in Tanzania).

## 440 **THE WAY FORWARD**

441 Progress toward the control and eradication of PPR in SADC is  
 442 now well-planned by many southern African countries. However,  
 443 there is a need to coordinate efforts at the regional level. Three  
 444 risk-based categories can be identified (infected, high-risk, lower-  
 445 risk countries, Figure 1B). Better coordination between countries  
 446 within the same category and between categories should improve  
 447 harmonized surveillance and targeted control.  
 448

449 Following the PCP, a better understanding of the  
 450 epidemiology of PPR in the region and its contributing factors  
 451 will be necessary for eradication and this will require funding  
 452 for field epidemiology research. Urgent active surveillance  
 453 is required to establish the extent of PPR sero-positive areas  
 454 in infected (across country) and high-risk (border areas)  
 455 countries. In parallel, sheep and goat movements need to be  
 456



**FIGURE 2 |** Progressive control pathway for SADC countries as reported during and following the Second Road Map Meeting, March 2019, Lusaka, Zambia.

better understood across the region. Studies on legal animal movement (e.g., by trucks or other vehicles) and other more informal cross-border movements will require participatory methods to enable mapping. A better understanding of cultural and social practices around small ruminant production systems of small-holder farmers in southern Africa is necessary in order to optimize surveillance and control of PPR (and other diseases). Women are known to be managing small ruminant production systems in Africa and, through communication and training tools, they should be empowered with the primary level of passive surveillance systems and control tools as identified in a study on gendered barriers to livestock vaccine uptake and ongoing gender inclusive vaccine study in Kenya (48, 49). Clarifying the role of wildlife and wildlife/livestock interfaces is also of paramount importance for SADC.

Risk-based approaches should be used to better understand the risks of introduction from infected to high-risk countries; the risk of disease spread once introduced into a new country and from there to other lower-risk countries. Spatial epidemiology can include different types of data layers such as the presence of wildlife populations, roads, density of small ruminants, each weighted by expert knowledge (50). These risk assessments are important to inform policy development, contingency planning, and for allocating scarce resources to high-risk areas within countries (13).

Veterinary services' capacity building is necessary in order to survey, control and eradicate PPR from SADC. In infected countries, going through stage 2–4 should be done through good communication with neighboring non-infected countries in order for them to survey for PPR with the most updated information. In high-risk countries, controlling animal movements is difficult with porous borders. Therefore, strategic

passive surveillance for early detection (e.g., clinical and laboratory surveillance in markets or cross-border trade hubs) and early-response (e.g., vaccination) is needed to prevent outbreaks in new areas. The specific epidemiological context of SADC countries implies that surveillance systems should be prepared to expect non-conventional disease expression as the incursion of PPR in the Maghreb region showed moderate clinical signs and low rates of mortality. Improving biosecurity and sanitary protection through Public Private Partnerships will also be necessary (51) and FAO and OIE can help by facilitating donor agency-country relationships. Capacity building and experience sharing between infected and non-infected countries are important as demonstrated in FAO/OIE workshops. Countries at lower-risk of PPR introduction should get prepared using risk-based approaches at reacting to PPR outbreaks on their territory given their specific context (in particular given the size of the wildlife industry in some countries). Vaccines that are thermotolerant, produced in large quantities and if possible have DIVA abilities are needed. Further, PPR molecular diagnostic training and laboratory equipment and reagents are also needed in the region.

## CONCLUSION

The support of international organizations (i.e., FAO and OIE) and SADC technical committees will be of paramount importance to ensure effective regional collaboration. The experience from meetings and trainings organized by these groups has shown that trust and sustainable relationships between stakeholders and veterinary services is crucial to facilitate information flow within the region. The updated SADC strategy for the control and eradication of PPR will further

guide regional coordination and provide leadership to meet the 2030 goal.

## DATA AVAILABILITY STATEMENT

All datasets generated for this study are included in the manuscript/supplementary files.

## AUTHOR CONTRIBUTIONS

AB and AC contributed equally to the design and writing of the manuscript. BB contribution to FAO activities, situation analysis of PPR in the region, laboratory capacity, and overall review of the manuscript.

## REFERENCES

- OIE. *OIE-Listed Diseases, Infections and Infestations in Force in 2019*. Paris (2019).
- OIE. *Chapitre 14.7. Infection With Peste des Petits Ruminants Virus*. Paris: OIE (2016)
- FAO, editor. *Supporting Livelihoods and Building Resilience Through Peste des Petits Ruminants (PPR) and Small Ruminant Diseases Control*. Rome: FAO (2013).
- Jones BA, Rich KM, Mariner JC, Anderson J, Jeggo M, Thevasagayam S, et al. The economic impact of eradicating peste des petits ruminants: a benefit-cost analysis. *PLoS ONE*. (2016) 11:e0149982. doi: 10.1371/journal.pone.0149982
- FAO, OIE. *Global Strategy for the Control and Eradication of PPR*. Paris: OIE and FAO (2015).
- FAO, OIE, editors. *Partnering and Investing for a peste des petits ruminants-free world*. In: *Partnering and Investing for a Peste des Petits Ruminants-Free World*. Brussels: FAO and OIE (2018).
- AU-IBAR. *The Pan African Strategy for Control and Eradication of Peste des Petits Ruminants*. Nairobi: AU-IBAR (2015).
- Southern African Development Community. *SADC Control Strategy for Peste de Petit Ruminants (PPR)*. Gaborone: Southern African Development Community (2012). p. 1–24 Available online at: [https://www.sadc.int/files/7413/5542/4349/PPR\\_Strategy.pdf](https://www.sadc.int/files/7413/5542/4349/PPR_Strategy.pdf)
- Torsson E, Berg M, Misinzo G, Herbe I, Kgotlele T, Paarni M, et al. Seroprevalence and risk factors for peste des petits ruminants and selected differential diagnosis in sheep and goats in Tanzania. *Infect Ecol Epidemiol*. (2017) 7:1368336. doi: 10.1080/20080686.2017.1368336
- Kgotlele T, Chota A, Chubwa CC, Nyasebwa O, Lyimo B, Torsson E, et al. Detection of peste des petits ruminants and concurrent secondary diseases in sheep and goats in Ngorongoro district, Tanzania. *Comp Clin Pathol*. (2018) 28:755–9. doi: 10.1007/s00580-018-2848-5
- Kivaria FM, Kwiatek O, Kapaga AM, Swai ES, Libeau G, Moshy W, et al. The incursion, persistence and spread of peste des petits ruminants in Tanzania: epidemiological patterns and predictions. *Onderstepoort J Vet Res*. (2013) 80:593. doi: 10.4102/ojvr.v80i1.593
- Cetre-Sossah C, Kwiatek O, Faharoudine A, Soule M, Moutroifi YO, Vrel MA, et al. Impact and epidemiological investigations into the incursion and spread of peste des petits ruminants in the Comoros archipelago: an increased threat to surrounding Islands. *Transbound Emerg Dis*. (2014) 63:452–9. doi: 10.1111/tbed.12296
- Chazya R, Muma JB, Mwacalimba KK, Karimuribo E, Mkandawire E, Simuunza M. A qualitative assessment of the risk of introducing peste des petits ruminants into northern zambia from Tanzania. *Vet Med Int*. (2014) 2014:202618. doi: 10.1155/2014/202618
- Baron MD, Diallo A, Lancelot R, Libeau G. Peste des petits ruminants virus. *Adv Virus Res*. (2016) 95:1–42. doi: 10.1016/bs.aivir.2016.02.001
- Mapaco L, Monjane I, Fafetine J, Arone D, Caron A, Chilundo A, et al. Peste des Petits Ruminants virus surveillance in domestic small ruminants, Mozambique (2015 and 2017). *Front Vet Sci*. (Submitted this issue).

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- Yekutieli P. *Eradication of Infectious Diseases. A Critical Study*. Basel: S. Karger AG (1980).
- Adedeji AJ, Dashe Y, Akanbi OB, Woma TY, Jambol AR, Adole JA, et al. Co-infection of peste des petits ruminants and goatpox in a mixed flock of sheep and goats in Kanam, North Central Nigeria. *Vet Med Sci*. (2019) 5:412–8. doi: 10.1002/vms3.170
- Abubakar M, Jamal SM, Arshed MJ, Hussain M, Ali Q. Peste des petits ruminants virus (PPRV) infection; its association with species, seasonal variations and geography. *Trop Anim Health Prod*. (2009) 41:1197–202. doi: 10.1007/s11250-008-9300-9
- Nkamwesiga J, Coffin-Schmitt J, Ochwo S, Mwiine FN, Palopoli A, Ndekezi C, et al. Identification of peste des petits ruminants transmission hotspots in the Karamoja subregion of Uganda for targeting of eradication interventions. *Front Vet Sci*. (2019) 6:221. doi: 10.3389/fvets.2019.00221
- IPCC. *Global Warming of 1.5°C. An IPCC Special Report on the Impacts of Global Warming of 1.5°C Above Pre-industrial Levels and Related Global Greenhouse Gas Emission Pathways, in the Context of Strengthening the Global Response to the Threat of Climate Change, Sustainable Development, and Efforts to Eradicate Poverty*. In Masson-Delmotte V, Zhai P, Pörtner HO, Roberts D, Skea J, Shukla PR, et al., editors. Geneva: IPCC (2018).
- Spiegel KA, Havas KA. The socioeconomic factors surrounding the initial emergence of peste des petits ruminants in Kenya, Uganda, and Tanzania from 2006 through 2008. *Transbound Emerg Dis*. (2019) 66:627–33. doi: 10.1111/tbed.13116
- Torsson E, Kgotlele T, Berg M, Mtui-Malamsha N, Swai ES, Wensman JJ, et al. History and current status of peste des petits ruminants virus in Tanzania. *Infect Ecol Epidemiol*. (2016) 6:32701. doi: 10.3402/iee.v6.32701
- Parida S, Muniraju M, Mahapatra M, Muthuchelvan D, Buczkowski H, Banyard AC. Peste des petits ruminants. *Vet Microbiol*. (2015) 181:90–106. doi: 10.1016/j.vetmic.2015.08.009
- Munir M. Role of wild small ruminants in the epidemiology of peste des petits ruminants. *Transbound Emerg Dis*. (2014) 61:411–24. doi: 10.1111/tbed.12052
- Couacy-Hymann E, Koffi MY, Kouadio VK, Mossoum A, Kouadio L, Kouassi A, et al. Experimental infection of cattle with wild type peste-des-petits-ruminants virus - Their role in its maintenance and spread. *Res Vet Sci*. (2019) 124:118–22. doi: 10.1016/j.rvsc.2019.02.011
- Mahapatra M, Sayalel K, Muniraju M, Eblate E, Fyumagwa R, Shilinde L, et al. Spillover of peste des petits ruminants virus from domestic to wild ruminants in the Serengeti ecosystem, Tanzania. *Emerg Infect Dis*. (2015) 21:2230–4. doi: 10.3201/eid2112.150223
- Kinne J, Kreutzer R, Kreutzer M, Wernery U, Wohlsein P. Peste des petits ruminants in Arabian wildlife. *Epidemiol Infect*. (2010) 138:1211–4. doi: 10.1017/S0950268809991592
- Marashi M, Masoudi S, Moghadam MK, Modirrousta H, Marashi M, Parvizifard M, et al. Peste des petits ruminants virus in vulnerable wild small ruminants, Iran, 2014–2016. *Emerg Infect Dis*. (2017) 23:704–6. doi: 10.3201/eid2304.161218

- 685 29. Aguilar XF, Fine AE, Pruvot M, Njeumi F, Walzer C, Kock R, et  
686 al. PPR virus threatens wildlife conservation. *Science*. (2018) 362:165–6.  
687 doi: 10.1126/science.aav4096
- 688 30. Osofsky SA, Cumming HM, Kock MD. Transboundary management of  
689 natural resources and the importance of a “One Health” approach. In Fearn E,  
690 editor. *State of the Wild: A global Portrait of Wildlife, Wildlands, and Oceans*.  
691 London: Island Press (2008), p. 88–98.
- 692 31. Caron A, Cornelis D, Foggini C, Hofmeyr M, de Garine-Wichatitsky M.  
693 African buffalo movement and zoonotic disease risk across transfrontier  
694 conservation areas, Southern Africa. *Emerg Infect Dis*. (2016) 22:277–80.  
695 doi: 10.3201/eid2202.140864
- 696 32. SADC, editor. *The Trans-boundary Animal Diseases (TADs) Project*.  
697 Gaborone (2012) Available online at: [https://www.sadc.int/themes/  
698 agriculture-food-security/livestock-production/#TADS](https://www.sadc.int/themes/agriculture-food-security/livestock-production/#TADS) Consulted May 2019
- 699 33. Sen A, Saravanan P, Balamurugan V, Rajak KK, Sudhakar SB, Bhanuprakash  
700 V, et al. Vaccines against peste des petits ruminants virus. *Expert Rev Vaccines*.  
701 (2010) 9:785–96. doi: 10.1586/erv.10.74
- 702 34. Mariner JC, Gachanja J, Tindih SH, Toye P. A thermostable presentation of  
703 the live, attenuated peste des petits ruminants vaccine in use in Africa and  
704 Asia. *Vaccine*. (2017) 35:3773–9. doi: 10.1016/j.vaccine.2017.05.040
- 705 35. Enchery F, Hamers C, Kwiatek O, Gaillardet D, Montange C, Brunel  
706 H, et al. Development of a PPR challenge model in goats and its  
707 use to assess the efficacy of a PPR vaccine. *Vaccine*. (2019) 37:1667–73.  
708 doi: 10.1016/j.vaccine.2019.01.057
- 709 36. Balamurugan V, Sen A, Venkatesan G, Bhanuprakash V, Singh RK.  
710 Protective immune response of live attenuated thermo-adapted peste  
711 des petits ruminants vaccine in goats. *Virus Dis*. (2014) 25:350–7.  
712 doi: 10.1007/s13337-014-0208-x
- 713 37. Kardjadj M, Luka PD. Factors affecting PPRV in African  
714 countries and their countermeasures. *Br J Virol*. (2016) 3:63–76.  
715 doi: 10.17582/journal.bjv/2016.3.3s.63.76
- 716 38. Hodgson S, Moffat K, Hill H, Flannery JT, Graham SP, Baron MD, et al.  
717 Comparison of the immunogenicities and cross-lineage efficacies of live  
718 attenuated peste des petits ruminants virus vaccines PPRV/Nigeria/75/1 and  
719 PPRV/Sungri/96. *J Virol*. (2018) 92:e01471–18. doi: 10.1128/JVI.01471-18
- 720 39. Mansoor MK, Al-Rawahi AH, El-Tahir HA, Al-Farai B, Hussain MH, Asi  
721 MN, et al. Concurrent vaccination of goats with foot and mouth disease  
722 (FMD) and peste des petits ruminants (PPR) booster vaccines. *Trop Anim  
723 Health Prod*. (2018) 50:1–3. doi: 10.1007/s11250-017-1391-8
- 724 40. Bodjo SC, Baziki J-d-D, Nwankpa N, Chitsungo E, Koffi YM, Couacy-Hymann  
725 E, et al. Development and validation of an epitope-blocking ELISA using an  
726 anti-haemagglutinin monoclonal antibody for specific detection of antibodies  
727 in sheep and goat sera directed against peste des petits ruminants virus. *Arch  
728 Virol*. (2018) 163:1745–56. doi: 10.1007/s00705-018-3782-1
- 729 41. Bataille A, Kwiatek O, Belfkhi S, Mounier L, Parida S, Mahapatra M,  
730 et al. Optimization and evaluation of a non-invasive tool for peste  
731 des petits ruminants surveillance and control. *Sci Rep*. (2019) 9:4742.  
732 doi: 10.1038/s41598-019-41232-y
- 733 42. Berguido FJ, Bodjo SC, Loitsch A, Diallo A. Specific detection of peste  
734 des petits ruminants virus antibodies in sheep and goat sera by the  
735 luciferase immunoprecipitation system. *J Virol Methods*. (2016) 227:40–6.  
736 doi: 10.1016/j.jviromet.2015.10.008
- 737 43. Cheng S, Sun J, Yang J, Lv J, Wu F, Lin Y, et al. A new immunoassay of  
738 serum antibodies against Peste des petits ruminants virus using quantum  
739 dots and a lateral-flow test strip. *Anal Bioanal Chem*. (2017) 409:133–41.  
740 doi: 10.1007/s00216-016-9972-3
- 741 44. FAO, OIE. *Peste des Petits Ruminants. Global Eradication Programme:  
742 Contributing to Food Security, Poverty Alleviation and Resilience. Five Years  
743 2017–2021*. (2016). Available online at: <http://www.fao.org/3/a-i6316e.pdf>
- 744 45. OIE, FAO, AIEA. *Workshop on PPR Prevention and Control*. Dar es  
745 Salaam (2013).
- 746 46. FAO, editor. *Terminal Report Capacity Building to Prevent Peste des Petits  
747 Ruminants Introduction into Malawi, Mozambique and Zambia*. Rome:  
748 FAO (2016).
- 749 47. OIE, editor. *The OIE Tool for the Evaluation of Performance of Veterinary  
750 Services (OIE PVS Tool)*. Paris (2019).
- 751 48. Mutua E, de Haan N, Tumusiime D, Jost C, Bett B. A qualitative study  
752 on gendered barriers to livestock vaccine uptake in Kenya and Uganda  
753 and their implications on Rift Valley Fever control. *Vaccine*. (2019) 7:E86.  
754 doi: 10.3390/vaccines7030086
- 755 49. IDRC. *Gender Inclusive Vaccine Distribution and Delivery Systems for  
756 Newcastle Disease and Peste des Petits Ruminants among Smallholder Farmers  
757 in Kenya*. IDRC (2019).
- 758 50. Tran A, Trevenne C, Lutwama J, Sserugga J, Gely M, Pittiglio C, et  
759 al. Development and assessment of a geographic knowledge-based model  
760 for mapping suitable areas for rift valley fever transmission in eastern  
761 Africa. *PLoS Negl Trop Dis*. (2016) 10:e0004999. doi: 10.1371/journal.pntd.0  
762 004999
- 763 51. OIE, editor. *The OIE PPP Handbook: Guidelines for Public-Private-  
764 Partnerships in the Veterinary Domain*. Paris: OIE (2019).
- 765 **Conflict of Interest:** The authors declare that the research was conducted in the  
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