



# Plant Development and Genetic Improvement Research Unit



## Regulation of the expression of ethylene biosynthesis genes *in Hevea brasiliensis* shoots

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**IRRDB Workshop Biotec 2009  
on Hevea Genome & Transcriptome**

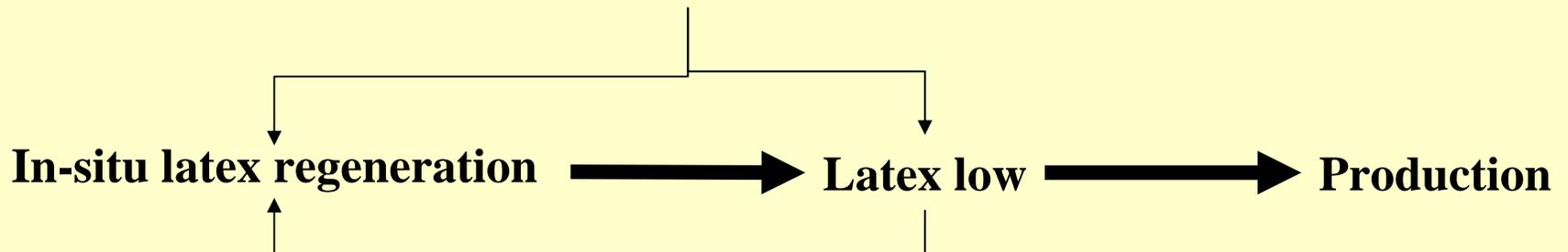
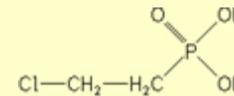
**3-5 June 2009 - Montpellier, France**

# Natural Rubber Production: Tapping & Ethephon Stimulation



## Ethephon

(2-chloroethyl-phosphonic acid)

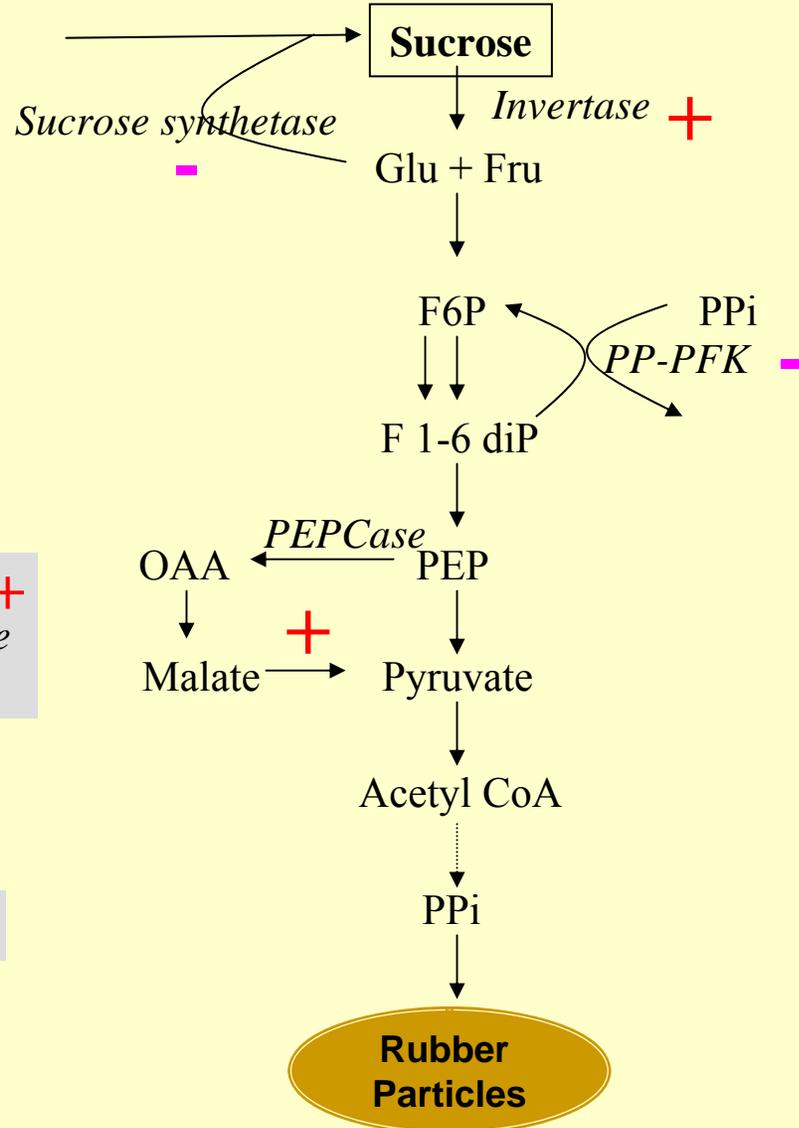
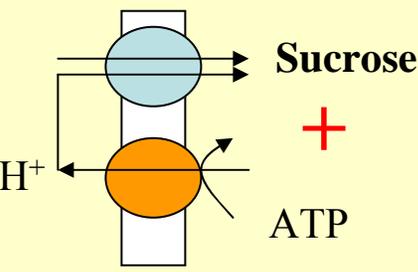


# Tapping Panel Dryness: dry cut and brown bast



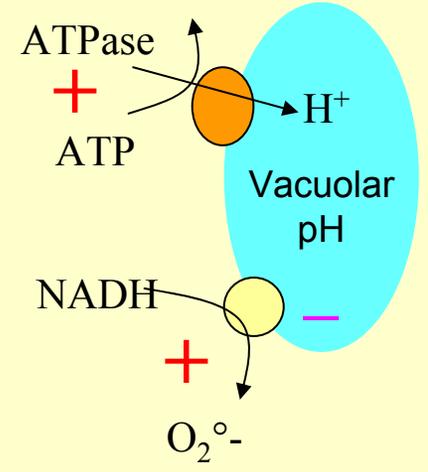
# Stimulation of the laticifer metabolism by ethylene

(from J. d'Auzac)



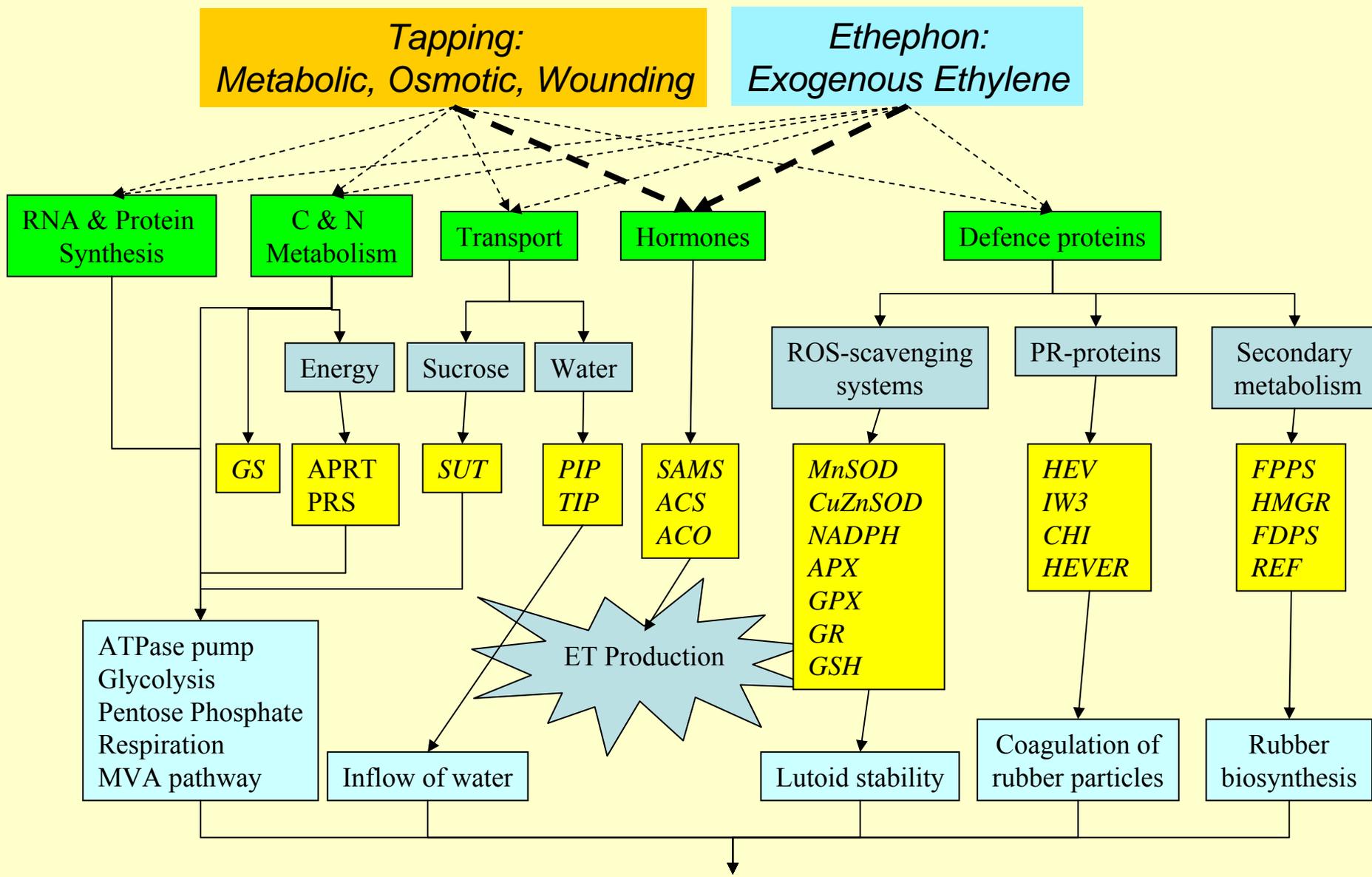
- cytosolic pH +
- Glu + NH<sub>4</sub> → Gln (+) via *Glutamine synthetase*
- Calmoduline +
- Phosphorylation
- Nucleotides adenines +
- ATP/ADP -
- Latex biosynthesis +

Wounding	
Chitinase	
Hevein	+
HRGP	
SOD	



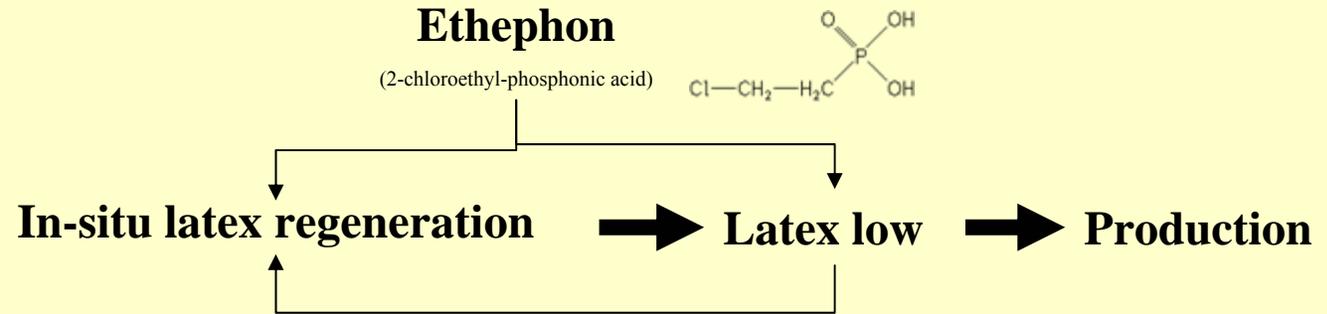
Senescence	
NADH oxydase	+
Peroxydase	
Catalase	-
SOD	+
Thiols	

# Wounding & ethylene regulation of the gene expression in laticifers



*Latex production vs TPD*

# Exogenous & endogenous ethylene production in bark tissue



**Siwey:** Endogenous ethylene production in bark tissue after tapping and after ethephon treatment

**Audley (1975):** ethephon could not induce ethylene production in 2-year-old stem

**Sivakumaran (1984):** Endogenous ethylene concentration in bark were higher in wound-susceptible clones than wound-resistant clones

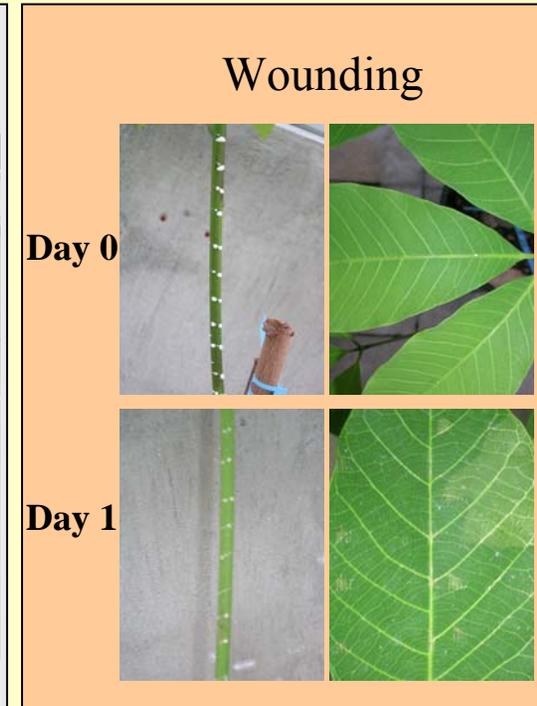
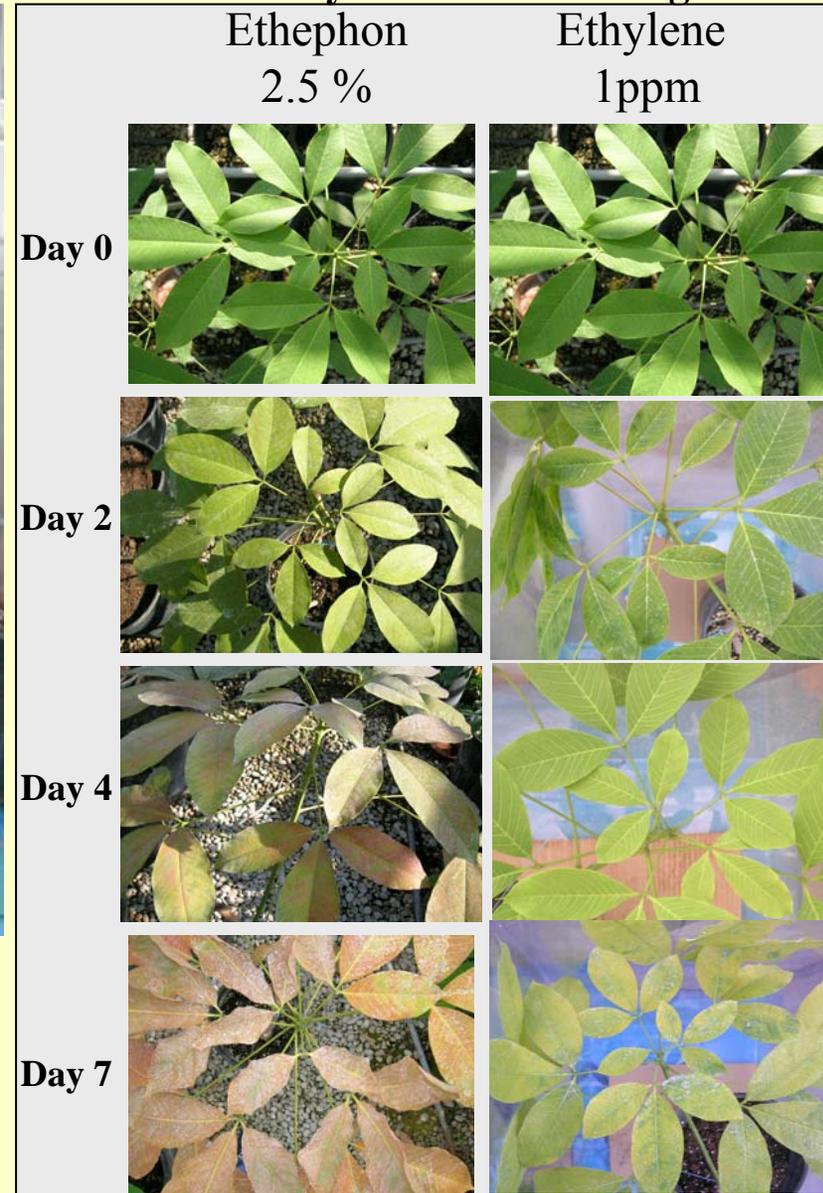


Tapping	Apparent	Ethylene (nl/g/h)		
		Exogenous	Endogenous	Endog./Exogenous
1	334	124	210	169 %
2	66	31	35	111 %
3	118	62	56	91 %
4	94	64	31	49 %

**Releasing ethylene of bark tissues after ethephon treatment**

# A model to study the regulation of the gene expression by ethylene and wounding.

Application and symptoms observed after ethylene and wounding treatments in 3-month-old shoots

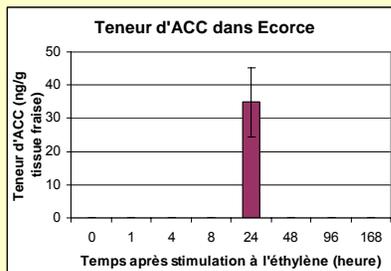
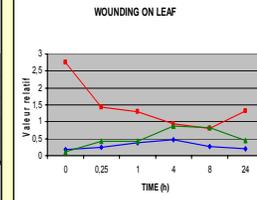
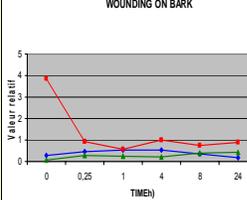
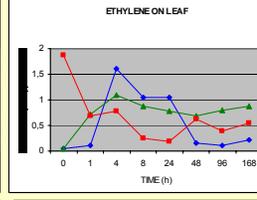
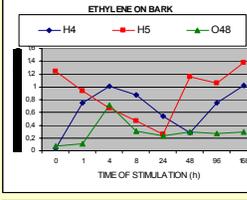
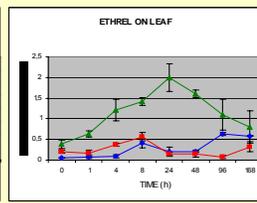
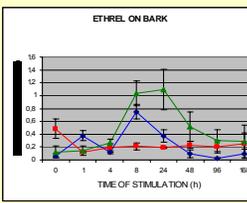
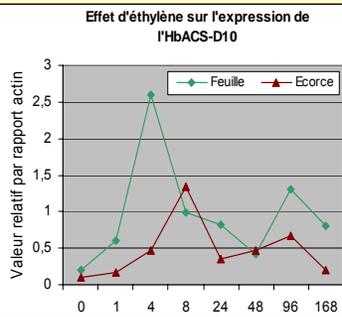
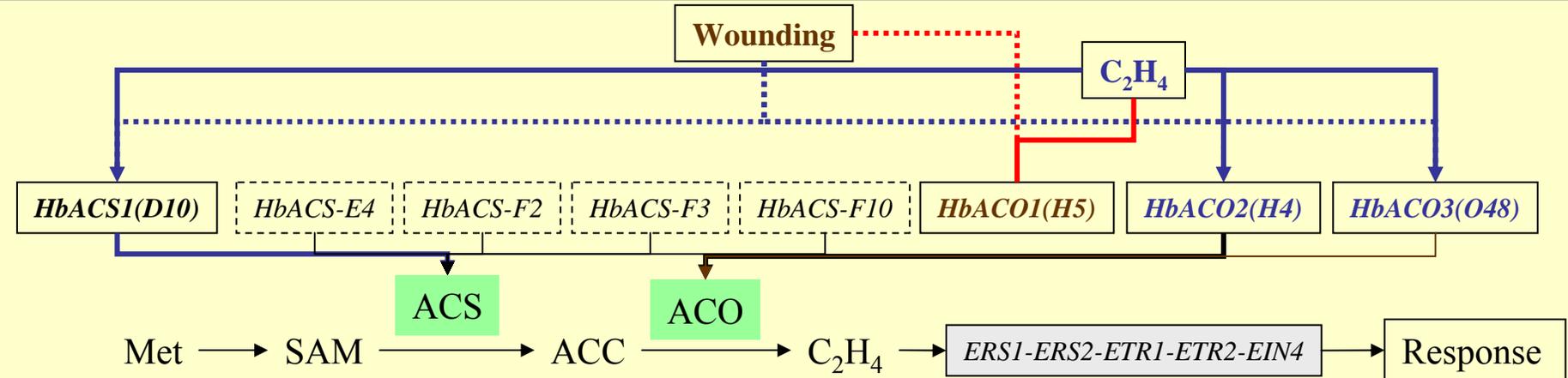


# Isolating & characterization of ACS and ACO genes involved in ethylene biosynthesis in *Hevea brasiliensis*

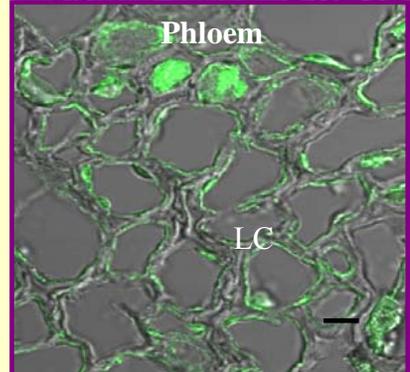
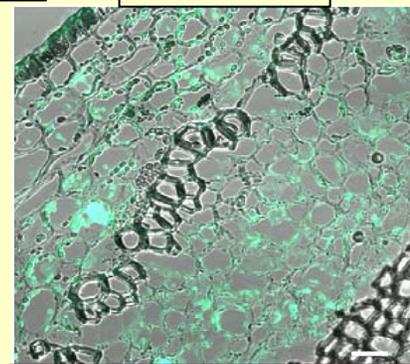
KUSWANHADI<sup>1</sup> (PhD, 2006), J. Leclercq<sup>2</sup>, L. Alemanno<sup>2</sup>, M. Rio<sup>2</sup>, J. Tregear<sup>3</sup>, M.-N. Ducamp-Collin<sup>4</sup>, P. Montoro<sup>2\*</sup>

<sup>1</sup> Sembawa Research Centre, Indonesian Rubber Research Institute, P.O. box 1127, Palembang, Indonesia - <sup>2</sup> UMR DAP, CIRAD, France

<sup>3</sup> IRD, UR 192, France - <sup>4</sup> UMR Qualisud, France

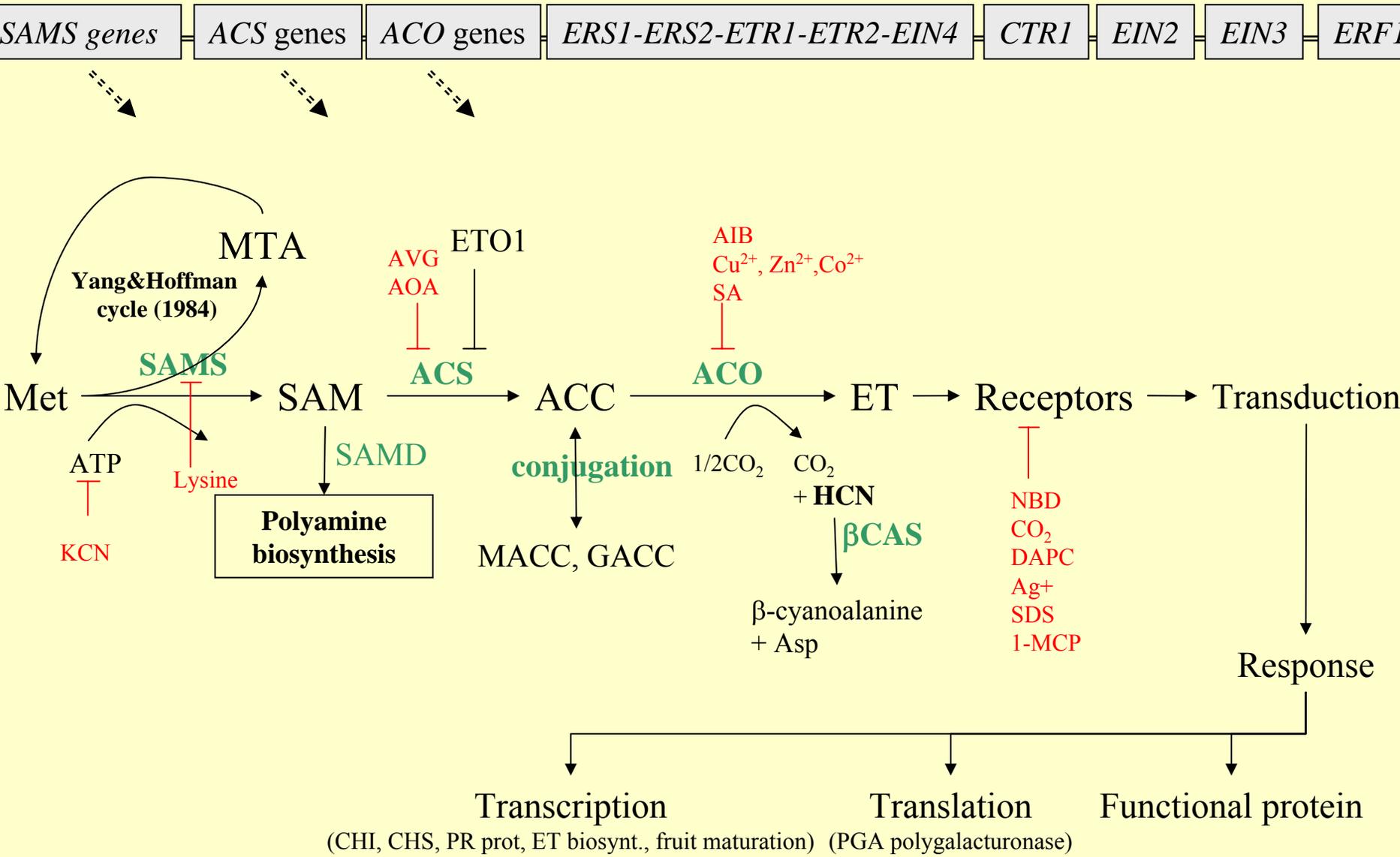


**1-MCP**

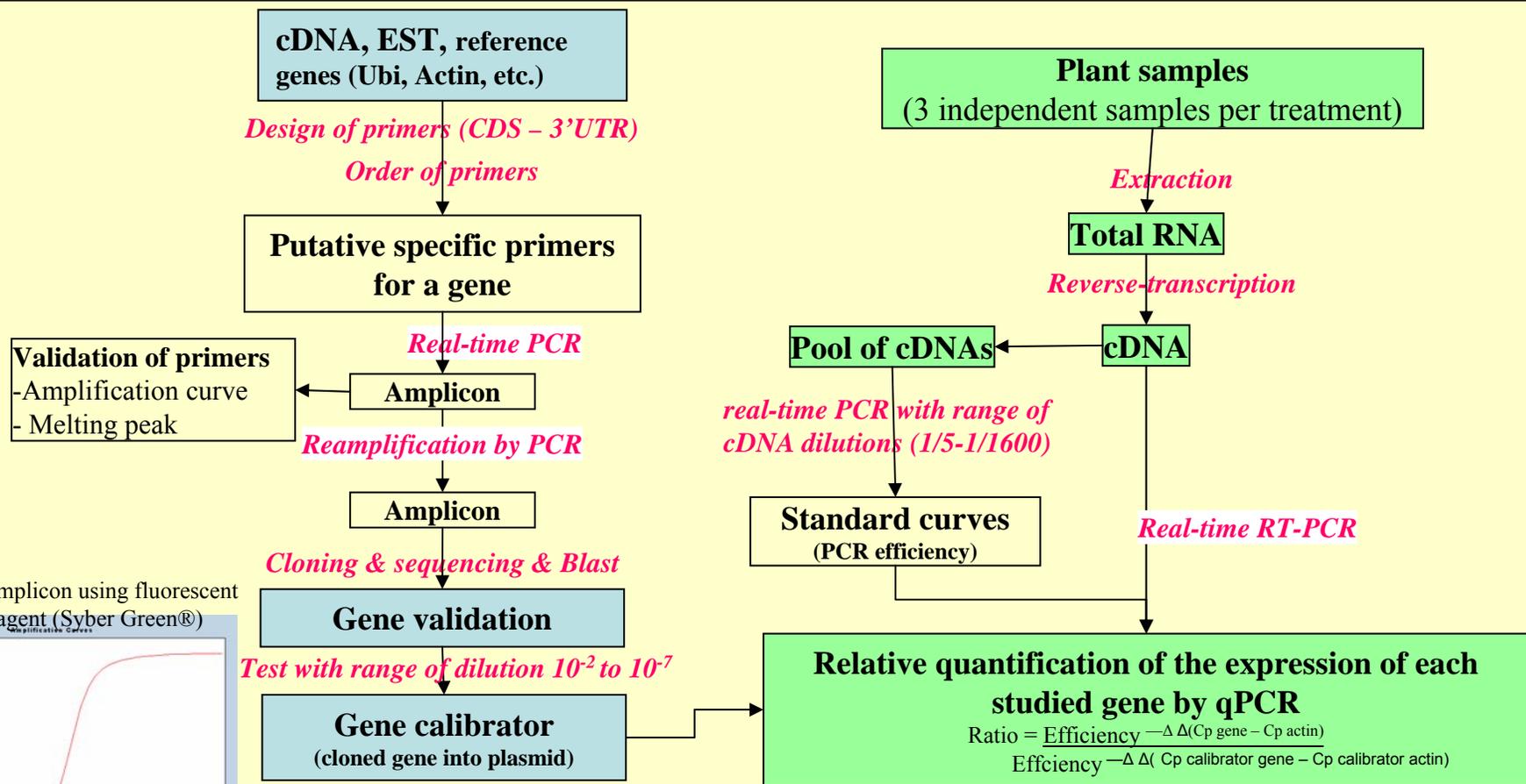


ACS amino-transferases (cofacteur : pyroxydal phosphate)  
 ACO super famille Fe<sup>2+</sup>/Ascorbate oxydase.

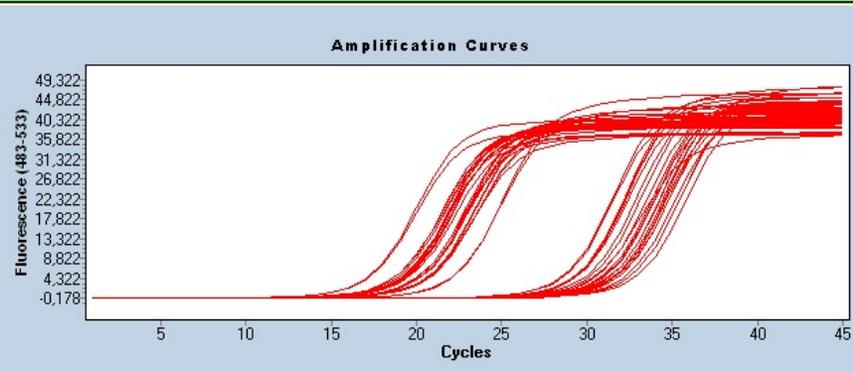
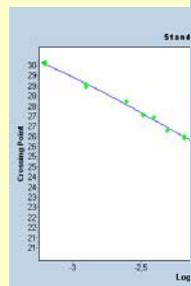
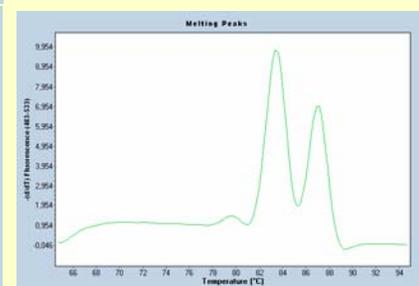
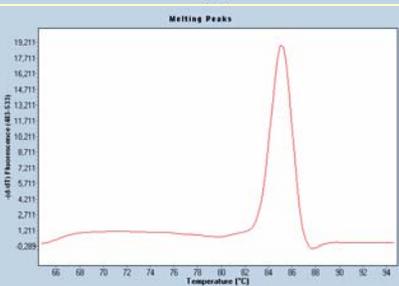
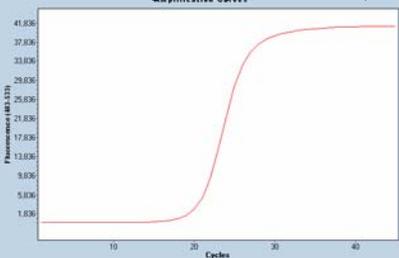
# Effect of wounding & ethylene on the expression of genes involved in ethylene biosynthesis and related pathways in 3 clones with contrasting metabolism



# Step for the analysis of the gene expression using real-time RT-PCR Light Cycler 480 (Roche)



Quantity of amplicon using fluorescent intercalant reagent (Syber Green®)



# High-throughput gene expression analysis using 384-well plate on real-time PCR Light Cycler 480 (Roche)

## 1. Manual preparation of 96-well plates

Primers, water & SyberGreen



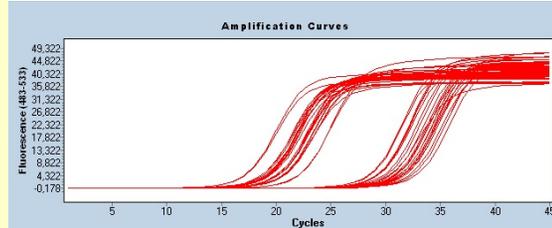
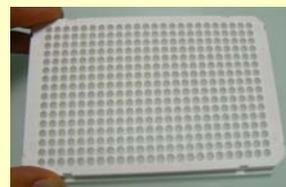
DNA templates



## 2. Preparation of 384-well plates using Robot Beckman Coulter



## 3. Real-time PCR analysis using 384-well plate on PCR Light Cycler 480 (Roche)

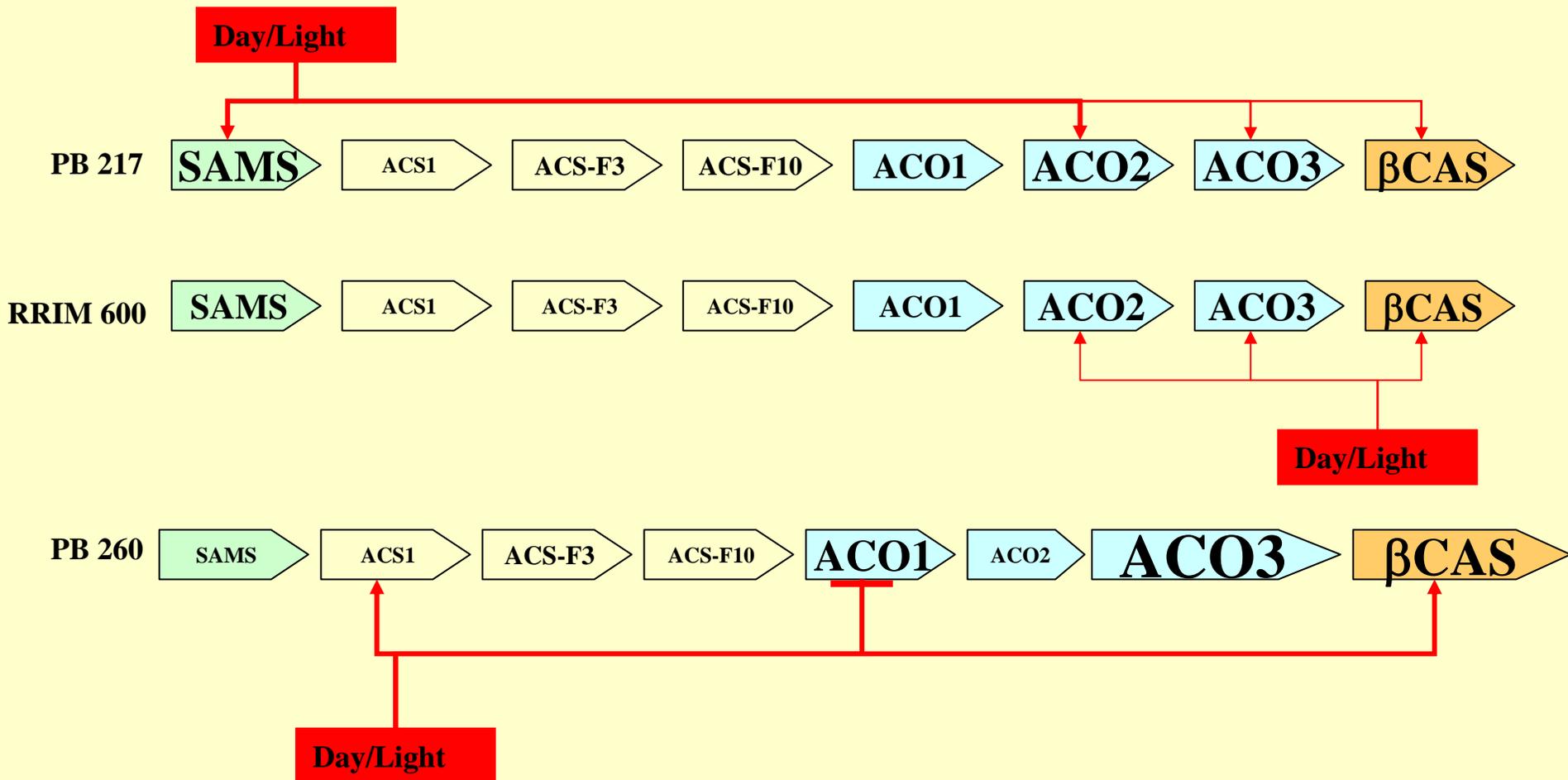


## Comparison of the expression of genes involved in ethylene biosynthesis during daytime in three clones with constrasting metabolism.

Value must be multiplied by  $10^{-4}$ . (\*) a-b Fisher test (LSD). All values with the same letter are not significantly different ( $P < 0.05$ ). Each value is the mean of 3 replications.

Clone	Daytime	SAMS	ACS1	ACSF10	ACO1	ACO2	ACO3	CAS
PB 217	8h00	1953 ± 110 <sup>b</sup>	0.6 ± 0.5 <sup>bc</sup>	20 ± 8 <sup>a</sup>	35 ± 26 <sup>bc</sup>	817 ± 533 <sup>b</sup>	216 ± 113 <sup>b</sup>	819 ± 220 <sup>bc</sup>
	12h00	2997 ± 409 <sup>a</sup>	0.5 ± 0.3 <sup>c</sup>	19 ± 5 <sup>a</sup>	60 ± 54 <sup>bc</sup>	1031 ± 529 <sup>b</sup>	257 ± 110 <sup>b</sup>	1339 ± 343 <sup>b</sup>
	16h00	3499 ± 1547 <sup>a</sup>	3.9 ± 4.9 <sup>bc</sup>	21 ± 5 <sup>a</sup>	41 ± 13 <sup>bc</sup>	4874 ± 3673 <sup>a</sup>	514 ± 247 <sup>b</sup>	1062 ± 118 <sup>bc</sup>
RRIM 600	8h00	413 ± 31 <sup>c</sup>	0.1 ± 0 <sup>c</sup>	1.4 ± 0.7 <sup>b</sup>	6 ± 3 <sup>c</sup>	55 ± 18 <sup>b</sup>	40 ± 21 <sup>b</sup>	214 ± 41 <sup>c</sup>
	12h00	509 ± 116 <sup>c</sup>	0.2 ± 0.2 <sup>c</sup>	1.9 ± 1.9 <sup>b</sup>	9 ± 9 <sup>c</sup>	118 ± 66 <sup>b</sup>	62 ± 58 <sup>b</sup>	293 ± 119 <sup>c</sup>
	16h00	496 ± 61 <sup>c</sup>	0.1 ± 0 <sup>bc</sup>	2.0 ± 0.7 <sup>b</sup>	4 ± 1 <sup>c</sup>	101 ± 19 <sup>b</sup>	107 ± 102 <sup>b</sup>	579 ± 268 <sup>bc</sup>
PB260	8h00	16 ± 7 <sup>c</sup>	4.1 ± 2.5 <sup>bc</sup>	12 ± 7 <sup>ab</sup>	507 ± 419 <sup>a</sup>	4 ± 3 <sup>b</sup>	273270 ± 128410 <sup>a</sup>	1531 ± 317 <sup>b</sup>
	12h00	16 ± 2 <sup>c</sup>	5.4 ± 1.1 <sup>b</sup>	24 ± 19 <sup>a</sup>	282 ± 162 <sup>ab</sup>	7 ± 9 <sup>b</sup>	354130 ± 188870 <sup>a</sup>	3803 ± 1266 <sup>a</sup>
	16h00	13 ± 4 <sup>c</sup>	14.7 ± 6.8 <sup>a</sup>	14 ± 11 <sup>ab</sup>	186 ± 80 <sup>bc</sup>	8 ± 2 <sup>b</sup>	314300 ± 260920 <sup>a</sup>	3794 ± 570 <sup>a</sup>

# Day/light regulation of the accumulation of transcripts for genes encoding enzymes related to ethylene biosynthesis & cyanide detoxification in 3 clones with contrasting metabolism



→ The effect of a treatment must be related to the expression of a control sampled at the same daytime

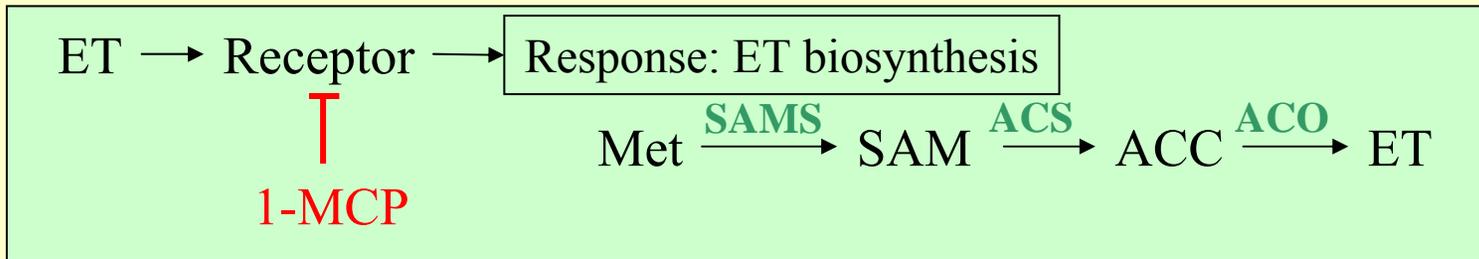
## Up-regulated genes (p-Value&lt;0.25)

## Non-regulated genes

## Down-regulated genes

Treatment		Clone	SAMS	ACS1	ACSF3	ACSF10	ACO1	ACO2	ACO3	CAS	UBI
Wounding	15 min	PB217	1,6	41,4	4,4	9,6	0,6	2,4	0,8	1,0	0,3
		RRIM 600	1,1	30,5	0,6	11,3	0,5	1,5	0,4	0,8	1,2
		PB 260	1,8	42,8	0,1	9,8	0,2	2,0	0,5	0,3	1,9
MCP/Wounding	15 min	PB217	1,2	4,3	0,2	5,9	0,3	0,4	0,0	0,8	0,6
		RRIM 600	1,0	8,5	3,4	1,3	0,1	1,2	0,2	1,2	0,8
		PB 260	1,2	52,1	19,6	9,2	1,2	2,5	0,8	1,4	1,7
Wounding	4h	PB217	1,4	7,0	65,5	3,3	0,7	7,1	1,5	1,1	1,1
		RRIM 600	4,2	3,0	4,0	2,8	0,5	3,0	1,4	0,9	0,8
		PB 260	1,3	5,6	4,9	0,5	0,2	4,8	1,4	1,5	0,6
MCP/Wounding	4h	PB217	0,8	2,3	15,5	0,8	1,6	7,5	53,2	1,6	1,1
		RRIM 600	0,7	1,0	1,4	0,7	1,5	1,2	0,8	1,1	0,9
		PB 260	1,7	3,7	5,2	1,3	2,1	2,6	1,5	1,3	1,7
Ethylene	4h	PB217	1,0	1,0	7,0	0,5	0,4	0,9	0,4	0,6	0,5
		RRIM 600	0,9	1,7	17,5	2,1	1,9	1,3	0,9	1,0	0,8
		PB 260	1,7	1,8		0,5	1,3	1,6	1,7	2,8	1,4
MCP/Ethylene	4h	PB217	0,6	1,6	0,4	1,1	1,6	1,7	0,4	0,9	0,9
		RRIM 600	1,3	0,6	1,3	1,2	0,9	0,8	1,0	1,1	1,0
		PB 260	1,3	5,9		1,7	1,4	1,1	1,7	1,4	0,8
Ethylene	24h	PB217	1,1	1,1	3,1	1,4	0,3	1,4	0,4	1,5	1,7
		RRIM 600	1,0	5,1	0,4	1,2	0,5	1,6	2,0	1,7	1,5
		PB 260	0,7	1,2		0,5	0,9	0,9	0,8	0,9	0,5
MCP/Ethylene	24h	PB217	0,9	5,5	0,7	0,2	0,4	1,1	0,2	0,2	1,8
		RRIM 600	1,3	0,8	33,1	1,5	1,2	0,7	0,8	0,9	1,4
		PB 260	1,4	2,2		1,9	0,5	1,6	1,1	0,5	1,3

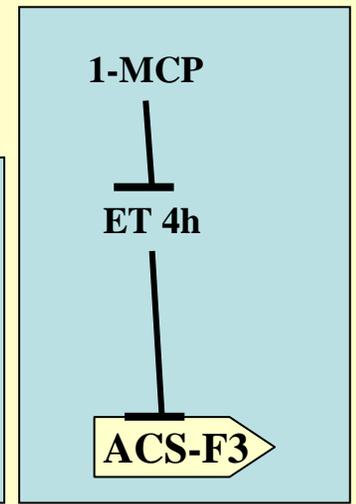
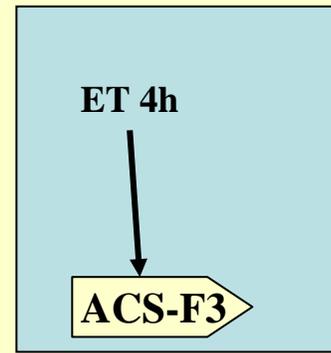
# Ratio of up- or down-regulation calculated with the level of gene expression in treated & control tissues sampled at the same daytime



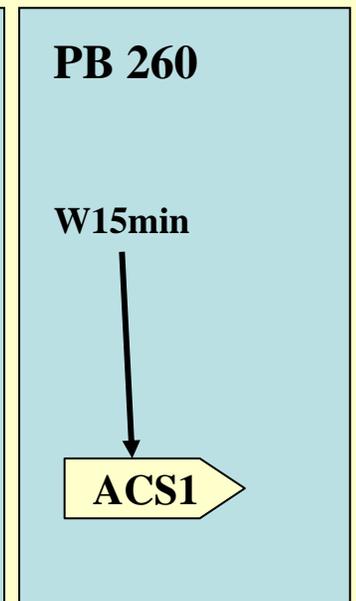
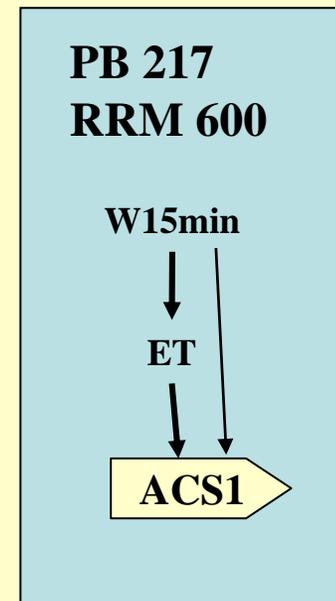
## Regulation of the expression of gene *ACS-F3* in clone RRIM 600 upon ethylene treatment. Three biological replicates

Pretreatment	Treatment		Expression ACSF3
<b>None</b>	<b>Control (8h00-12h00)</b>	Mean	0.00261
<b>None</b>	<b>Ethylene 4h (8h00-12h00)</b>	Mean	0.04560
		Ratio	17,5
		p-value	0,21
<b>1-MCP 16h</b>	<b>Control (8h00-12h00)</b>	Mean	0.01040
<b>1-MCP 16h</b>	<b>Ethylene 4h (8h00-12h00)</b>	Mean	0.01335
		Ratio	1,3
		p-value	0,69

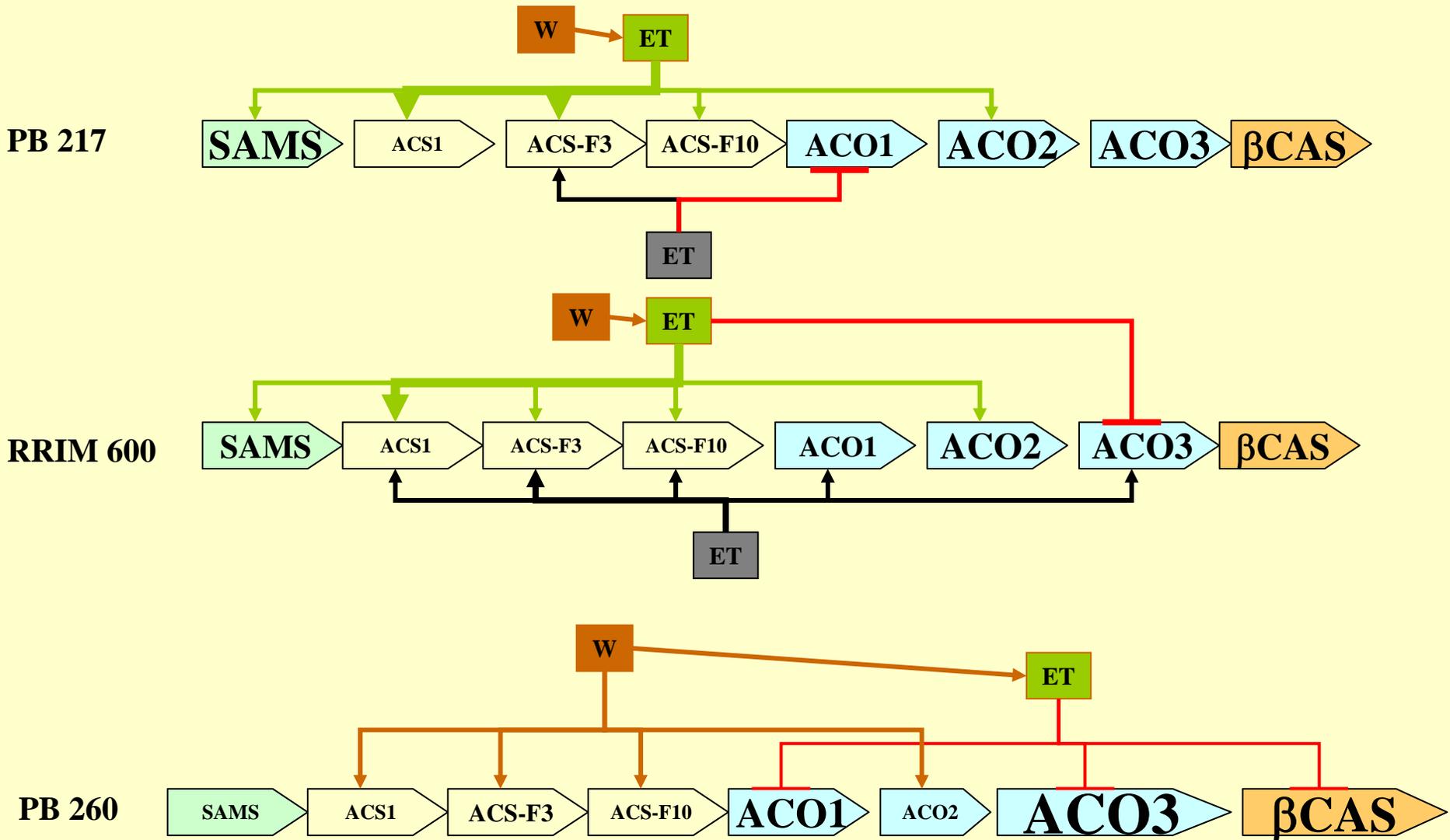
				ACSF3
<b>Ethylene</b>	4h	RRIM 600	ratio	17,5
			p-value	0,21
<b>MCP/Ethylene</b>	4h	RRIM 600	ratio	1,3
			p-value	0,69



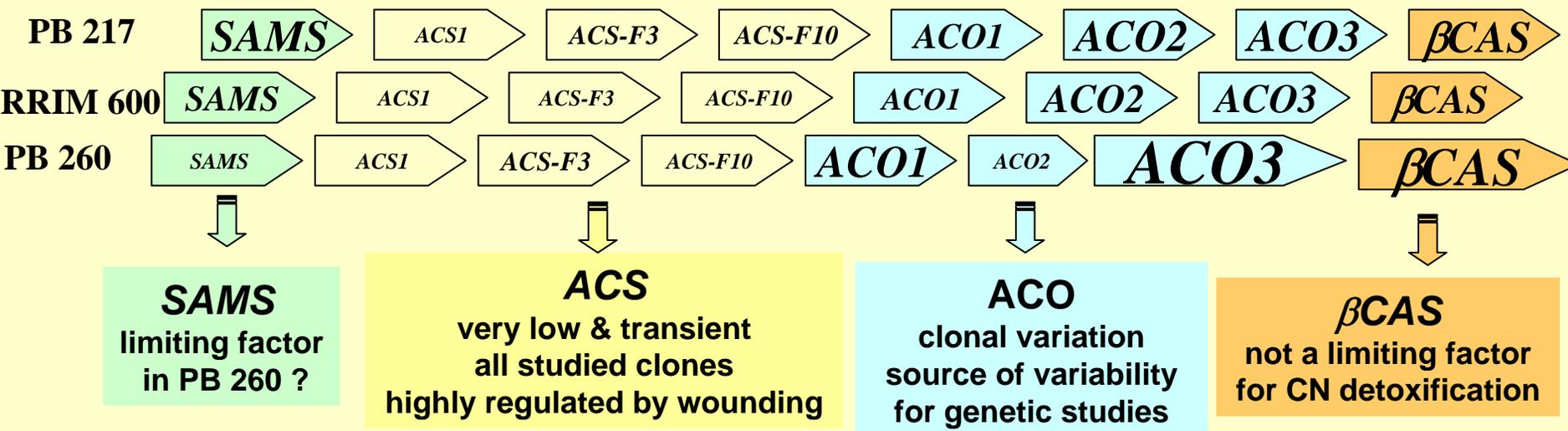
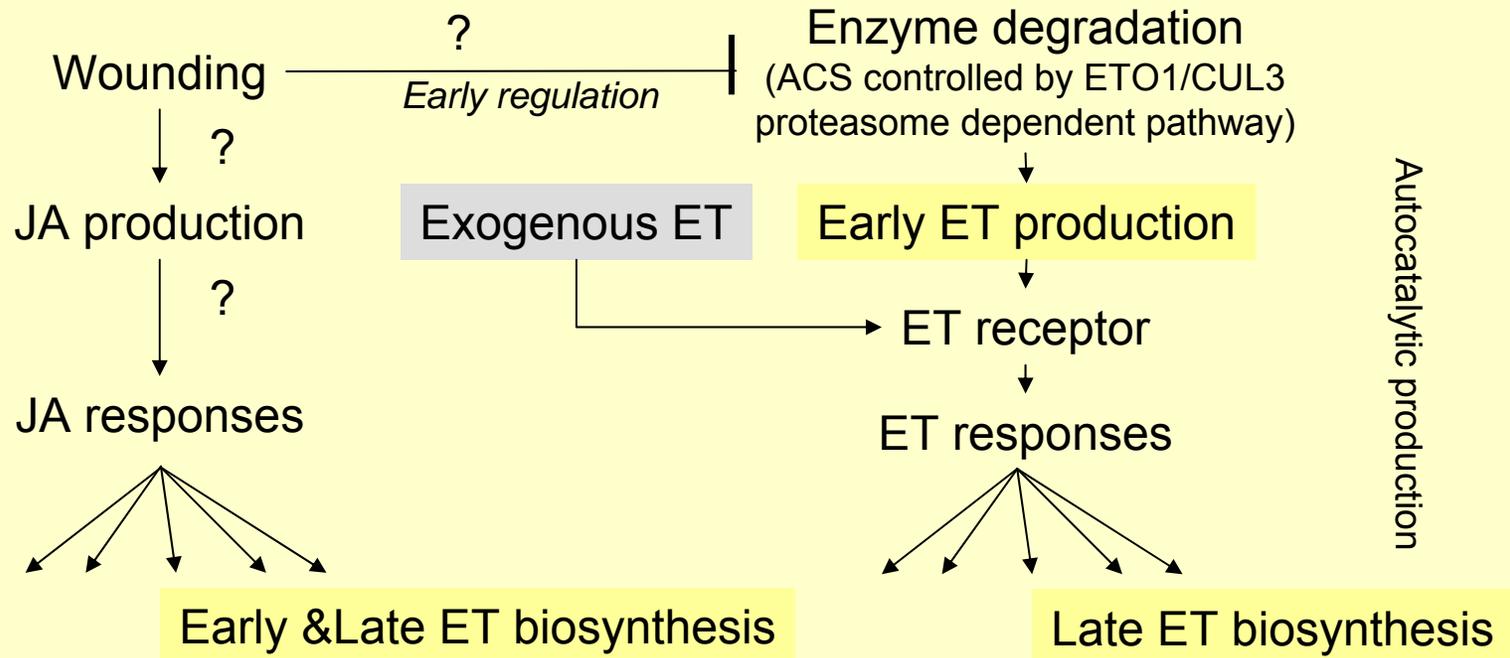
Treatment		Clone		ACS1
<b>Wounding</b>	15 min	PB217	ratio	41,4
			p-value	0,10
		RRIM 600	ratio	30,5
			p-value	0,24
		PB 260	ratio	42,8
			p-value	0,21
<b>MCP/Wounding</b>	15 min	PB217	ratio	4,3
			p-value	0,18
		RRIM 600	ratio	8,5
			p-value	0,13
		PB 260	ratio	52,1
			p-value	0,21



# Early and late wounding & ethylene effects on the accumulation of transcripts for genes encoding enzymes related to ethylene biosynthesis & cyanide detoxification



# Regulation of the ethylene biosynthesis





# Plant Development and Genetic Improvement Research Unit



## Acknowledgements

**Plant Development and Genetic Improvement Research Unit**  
**CIRAD [www.cirad.fr](http://www.cirad.fr)**

**BURST Group – Biology of Responses to STresses in tropical perennial species**  
**Leclercq, Montoro, Oliver, Rio**

**GS Group – Gene & Selection – **Xavier SABAU****

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