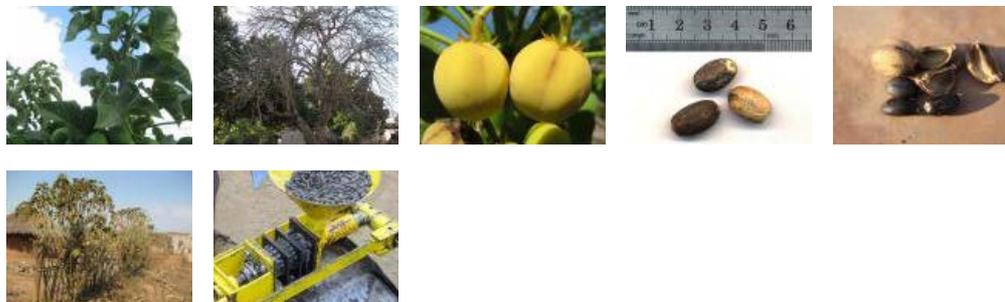


Jatropha (*Jatropha* sp.) kernel meal and other jatropha products

Description Nutritional aspects Nutritional tables References

Click on the "Nutritional aspects" tab for recommendations for ruminants, pigs, poultry, rabbits, horses, fish and crustaceans



Common names

Jatropha, physic nut, purging nut, pulza, nettlespurge [English]; pourghère, pignon d'Inde, médicinier [French]; coquillo, template, piñon, piñoncillo, piñol, tártago [Spanish]; mundubi-assu, turgueira [Portuguese]; mupuluka [Angola]; fagiola d'India [Italian]; Chi Dầu mè [Vietnamese]; [Amharic]; جنروفا [Arabic]; i, [Bengali]; 麻疯树属 [Chinese]; [Hindi]; سانगोप्रागिरी [Japanese]; , [Marathi]; Ятрофа [Russian]; [Tamil]; [Telugu]

Products: jatropha kernel meal, jatropha oil meal, jatropha oil cake, jatropha seed meal, jatropha shells, jatropha hulls

Species

Jatropha curcas L. ; *Jatropha platyphylla* Müll. Arg. ; *Jatropha tanjorensis* Ellis & Saroja [Euphorbiaceae]

Synonyms

Synonym for *Jatropha platyphylla* Müll. Arg.: *Jatropha peltata* Sassé

Feed categories

- Oil plants and by-products
- Toxic plants

Related feed(s)

Description

Jatrophas are oil plants belonging to the Euphorbiaceae family. The *Jatropha* genus includes more than 175 species, among which *Jatropha curcas* L. and *Jatropha platyphylla* Müll. Arg. have drawn particular attention as potential biofuel and animal feed sources. *Jatropha curcas* has both toxic and non-toxic genotypes while *Jatropha platyphylla* is a non-toxic species (Makkar et al., 2012).

The jatropha plant

Jatrophas are drought-resistant perennial and multipurpose shrubs or trees, similar to the cassava plant (Elbehri et al., 2013). *Jatropha* is a deciduous tree, shedding its leaves during the dry season. It can grow to a height of 3-5 m, and remains productive for 30 to 50 years. *Jatropha* has a deep taproot and four shallow lateral roots. The taproot anchors the plant in the soil, stabilizing the soil against landslides, and the lateral roots prevent soil erosion. The trunk is covered with a smooth grey bark that exudes watery and sticky latex when cut. The leaves are smooth, 4-6 lobed, 10-15 cm long and wide, and are usually pale green in colour (Raheman, 2012). Inflorescences develop at the apex of the branches and bear approximately 10 or more ovoid fruits (pods) (Makkar et al., 2012). Flowers and fruits develop during the rainy season or year-round in permanently humid regions. The pods contain many elliptic seeds and become yellow when they mature. Dry *Jatropha* fruits contain about 38% husks and 62% seeds. The seeds look like castor seeds in shape, and are black in colour (Raheman, 2012). They consist of 30-40% testa (shells) and 60-70% kernels. The kernels contain 44-62% oil (King et al., 2009).

Most *Jatropha* species, including *Jatropha curcas*, contain numerous toxic components and the plant and its by-products, when not detoxified, are deleterious to humans and livestock. One species, *Jatropha platyphylla* is non-toxic and is eaten by indigenous people in the Sinaloa State of Mexico, who call it "sangregado". This species has thick succulent branches and its leaves, borne on long petioles, are glabrous and larger (25-35 cm) than those of *Jatropha curcas*. Its seeds are almost circular and contain 50-60% edible oil (Makkar et al., 2011).

Jatropha utilisation

Jatrophas are multipurpose plants. The name jatropha was derived from the Ancient Greek words *iatros* (doctor) and *trophos* (feed) because of its many potential medicinal applications (Elbehri et al., 2013).

- **Jatropha seed oil** is used to make soap, lubricants, varnish, insecticides or medicines (Raheman, 2012). It has good feedstock qualities for biodiesel production. *Jatropha* biodiesel meets European Union (EN14214) and North American standards (ASTM D6751) (Makkar et al., 2012).

Automatic translation

Sélectionner une langue

Feed categories

All feeds

Forage plants

- ▶ Cereal and grass forages
- ▶ Legume forages
- ▶ Forage trees
- ▶ Aquatic plants
- ▶ Other forage plants

Plant products/by-products

- ▶ Cereal grains and by-products
- ▶ Legume seeds and by-products
- ▶ Oil plants and by-products
- ▶ Fruits and by-products
- ▶ Roots, tubers and by-products
- ▶ Sugar processing by-products
- ▶ Plant oils and fats
- ▶ Other plant by-products

Feeds of animal origin

- ▶ Animal by-products
- ▶ Dairy products/by-products
- ▶ Animal fats and oils
- ▶ Insects

Other feeds

- ▶ Minerals
- ▶ Other products

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- **Jatropha oil cake** (oil kernel meal) results from the oil extraction of decorticated seeds. It can be used as fertilizer or as feedstock in biogas production. Jatropha kernel meal is a protein-rich product, but only kernel meal obtained from non-toxic genotypes of *Jatropha curcas*, from the non-toxic species *Jatropha platyphylla*, or detoxified kernel meal from toxic genotypes of *Jatropha curcas* can be safely used as feed resources (Makkar et al., 2012). Jatropha seed meal is obtained after the extraction of non-dehulled seeds.
- **Jatropha plants** are used to make fences and shelter to protect from animals, wind or erosion (Elbehri et al., 2013).
- **Jatropha roots** yield an oil that has anthelmintic properties.
- **Jatropha leaves** are used as feed in the rearing of silkworms, and in human nutrition as a vegetable for their antimicrobial and anti-inflammatory properties (Makkar et al., 2009).
- **Whole seeds** of the non-toxic genotype are sometimes boiled or roasted and eaten as a snack by humans.
- **Jatropha latex** is used as dye or as a pesticide/molluscicide.
- **The fruit hulls** can be used as green manure or to produce biogas.

The rise and fall of jatropha as a source of biofuel

In the 2000s, the cultivation of jatropha was encouraged in many developing countries in order to provide both renewable energy and supplementary incomes (Promode Kant et al., 2011). By 2010, about 250 jatropha projects were running worldwide on a total area of around 900 000 ha (Elbehri et al., 2013). In India, millions of marginal farmers and landless people were encouraged to plant jatropha. In 2006, the Chinese Government decided to grow jatropha over 1 million ha of marginal lands. In Tanzania, more than 10,000 small farmers established jatropha plantations and many more have done so elsewhere in East Africa (Promode Kant et al., 2011). However, the results have been disappointing. In India, 85% of the jatropha farmers have discontinued growing it (Axelsson et al., 2011). In China, there is very little production of biodiesel from Jatropha seeds. In Tanzania, the results are very unsatisfactory: the net present value of a five-year investment in a jatropha plantation was negative, with a loss of US\$ 65 per ha on lands with yields of 2 t/ha of seeds when the expected jatropha seed yield on poor barren soils is only 1.7 to 2.2 t/ha (Promode Kant et al., 2011). This "extraordinary collapse of jatropha as a global biofuel" has been ascribed by analysts to an overestimation of jatropha capabilities, as the species is not yet sufficiently adapted to cultivation to provide sustainably high yields and economic returns, together with a lack of diligence in implementing nationwide cultivation plans (Axelsson et al., 2011; Promode Kant et al., 2011).

Distribution

Jatropha is thought to have originated from tropical America (Mexico). It could have been introduced to other regions from the Caribbean by Portuguese sailors travelling to Cape Verde and Guinea Bissau. It is now available in many arid, semiarid, tropical and subtropical regions of the world as it is able to thrive in many climatic zones with a wide range of rainfall (200 to 1200 mm annual rainfall) (Raheman, 2012). Jatropha is well adapted to arid and semiarid conditions. It can grow on marginal lands with limited water and agrochemical supply but will not yield enough seeds in such conditions (Elbehri et al., 2013; Raheman, 2012; Brittain et al., 2010). Jatropha does better in areas with 1000-1500 mm annual rainfall, temperatures of 20-28°C with no frost, and where the soils are free-draining sands and loams with no risk of waterlogging (Elbehri et al., 2013; Raheman, 2012). However, it grows on almost any type of soil including gravelly, sandy, stony or rocky soils and in the crevices of rocks where soil depth and fertility are very low. It responds positively to fertilizer or manure with added calcium, magnesium and sulfur. For economic returns, a soil with moderate fertility is preferred (Raheman, 2012).

Processes

After harvesting, jatropha fruits are dehusked and the seeds are sun-dried and then shelled. The kernels are pressed for oil extraction, resulting in a protein-rich jatropha kernel meal.

Detoxification

Because jatropha kernel by-products contain many toxic and antinutritional substances, different processes have been developed to mitigate toxicity or poor feed efficiency. These processes have been extensively reviewed by Makkar et al., 2012. Further methodological information on solvent and meal:solvent ratio can also be found in Makkar et al., 2012.

Chemical treatments	Effects	Reference
Ethanol (80%) or methanol (92%)	Saponins and phorbol esters reduced by 95% after four extractions	Makkar et al., 1997
Moist heating at 121°C for 20 minutes	Total inactivation of trypsin inhibitors	Makkar et al., 1997
Alkaline methanol + heat treatment	Phorbol ester reduction and trypsin inhibitor inactivation	Makkar et al., 2010b
Methanol	Phorbol ester reduced by 100%	Pighinelli et al., 2012
Ethanol (unknown dose) followed by NaHCO ₃ treatment	Lectin inactivation and phorbol esters reduced by 97.9%	Makkar et al., 2012
Ethanol (95%) at 35°C + heating with pressurized steam at 90°C for 30 minutes + sun-drying and further autoclaving at 121°C for 30 minutes	Phorbol esters and antinutrients were still active	Chivandi et al., 2006
Petroleum ether extraction	Phorbol esters reduced by 67.7% in kernels	Chivandi et al., 2004
Solid-liquid extraction with a sequential combination of hexane and methanol	Phorbols esters reduced by 99.6%	Gaur, 2009
Double solvent extraction with methanol/ethanol (50/50 for 8h)	Phorbol esters reduced by 97.30% (down to 0.10 mg/g) in cake	Guedes et al., 2014
Double solvent extraction followed by moist heat treatment	Phorbol esters reduced by 70.8% in kernels	Chivandi et al., 2004
Double solvent extraction + wet extrusion, re-extraction with hexane and moist heat treatment	Phorbol esters reduced by 87.7% in kernels	Chivandi et al., 2004
Heat treatment + bentonite + zinc oxide + NaHCO ₃ (4%)	Phorbol ester content reduced to 0.05-0.04 mg/g (lower than in non toxic jatropha varieties)	Sadubthummarak et al., 2013
CaOH (2%)	Phorbol esters reduced by 90% in meal	Rakshit et al., 2007
NaHCO ₃ or NaOH (3%)	Phorbol esters reduced by 55% in meal	Elangovan et al., 2013
NaHCO ₃ or NaOH (3%)	Curcin reduced by 100%	Elangovan et al., 2013
Biological treatments	Effects	Reference
Solid state fermentation of seed cake with white-rot fungi <i>Bjerkandera adusta</i> or <i>Phlebia rufa</i>	Phorbol esters reduced by 91% and 97% respectively	de Barros et al., 2011
Solid state fermentation of seed cake with	Phorbol esters reduced to undetectable level	Joshi et al., 2011

Pseudomonas aeruginosa PseA strain
within 9 days under 30°C, pH =7 and 65%
relative humidity

In 2010, a process involving meal heat treatment, alkali treatment (up to pH=11), acid treatment (down to pH=8) with short-chain alcohol (for precipitation) was able to reduce phorbol esters below 3 mg/kg, thus allowing the use of the detoxified jatropha kernel meal in animal feeding and the production of a detoxified jatropha protein isolate (Makkar et al., 2010b).

Forage management

Jatropha is a fast-growing plant that can be propagated either by seedlings (generative method) or direct planting of cuttings (vegetative method) (Raheman, 2012).

Cultivation

Jatropha cuttings can be planted directly in the field, in nursery beds, or in polyethylene bags, during the rainy season, at 10-20 cm depth in the soil. When jatropha plants develop from cuttings, they produce many branches but yield fewer seeds and have not enough time to develop their taproot, which makes them sensitive to wind erosion. Presoaked seeds (24 h in cold water), directly sown in wet soil or in seed beds under nursery conditions at the beginning of the rainy season are advisable for seed production. Jatropha can be easily intercropped with shade-tolerant plants, legume seeds such as black gram (*Vigna mungo*), and vegetables, such as tomato, bitter melon (*Momordica charantia*), pumpkin, and cucumber can be profitably grown under jatropha plants for the first two years. Irrigation and fertilizers are beneficial during the establishment period. After establishment, irrigation is not necessary but fertilizer should be applied every year during the productive period. Pruning should be done to promote branching and subsequent fruit yield (Raheman, 2012).

Harvest and yields

Depending on climatic conditions, jatropha may be harvested twice in a year under semiarid conditions, or continuously in more humid areas. Fruits have to be handpicked and harvest is time-consuming. Jatropha seed yields are variable, depending on growing conditions: in degraded land with low inputs, yields were reported to range from 1 to 2.5 t/ha/year; in fertile soils with high inputs jatropha could produce 2 to 5 t/ha/year (Elbehri et al., 2013). It has been noted that even on adequately fertile soils, jatropha is no match for sunflower and that the hugely variable yields have contributed to its decline in the early years of the present century (Promode Kant et al., 2011).

Environmental impact

Soil erosion control and wind shelter

Jatrophas, because of their taproot and lateral roots, can play two important roles in soil preservation. Lateral roots protect soil from heavy rains and provide soil cohesion, decreasing soil erodibility (Reubens et al., 2011; Henning, 2009). The deep taproot extracts subsurface soil moisture and enhances vegetative cover or allows intercropping, even in very dry environments. Planting jatropha in lines to form contour bunds improves rainwater infiltration. Jatropha hedges reduce wind erosion by reducing wind velocity and by binding the soil with their surface lateral roots (Henning, 2009).

Live fence

Jatropha is used as a living fence in many tropical and subtropical countries. Jatropha leaves are not eaten by livestock: goats would die of starvation rather than browsing jatrophas. When jatropha is intended for live fences, jatropha cuttings are planted as a hedge to protect homes, gardens and fields from wandering animals. Hedges planted very close together (5 cm) form a barrier that is impenetrable even by chickens (Henning, 2009).

Datasheet citation

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English correction by Tim Smith (Animal Science consultant) and Hélène Thiollet (AFZ)

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Jatropha (*Jatropha* sp.) kernel meal and other jatropha products

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Nutritional attributes

Jatropha seeds and kernels

Jatropha seeds contain highly variable amounts of shells (30-40%) and kernels (60-70%) (King et al., 2009). Their crude protein and oil content is 16-18% and 34-37%, respectively (Makkar et al., 2009), while the kernels contain 22-35% protein and 55-58% oil (DM basis). Jatropha oil is mainly composed of unsaturated fatty acids, notably oleic acid (35-41% of fatty acids) and linoleic acid (35-43% of fatty acids) (Makkar et al., 2011).

Jatropha oil by-products

The composition of jatropha oil by-products depends on the extraction process. The scientific literature contains references to different jatropha products containing variable amounts of residual oil and shells. However, the potentially most valuable product is the defatted jatropha kernel meal, due to its high protein content.

Jatropha kernel meal

The main product is the jatropha kernel meal, obtained from the complete oil extraction of dehulled seeds. Jatropha kernel meal contains from 40% to more than 75% crude protein (DM basis). Well defatted jatropha kernel meal contains less than 2% oil. It is relatively low in fibre (crude fibre less than 10% DM). Starch content ranges from 7 to 12% DM. Jatropha kernel meal contains good amounts of phosphorus, potassium, calcium, and magnesium. The amino acid composition of jatropha kernel meal and soybean meal are similar, except for lysine content, which is lower, and for the sulphur-containing amino acids cystine and methionine contents, which are higher in the jatropha kernel meal. Jatropha kernel meal contains low levels of non-protein nitrogen (9% of total nitrogen), suggesting a high level (91%) of true protein (Makkar et al., 1998; Makkar et al., 2009).

Jatropha seed meal

Several papers mention a jatropha seed meal obtained from the extraction of whole (non-dehulled) seeds. This product is much lower in protein (about 16% DM) and much higher in fibre (crude fibre more than 33% DM) than the kernel meal (Abd El-Rahman et al., 2011; Abdel-Shafy et al., 2011; Chivandi et al., 2006; Guedes et al., 2014; Sumiati et al., 2009).

Jatropha shells

Jatropha shells are a low-protein (5% DM), high-fibre (ADF more than 75% DM) by-product of low nutritive value.

Potential constraints

Raw jatropha products tend to cause morbidity or mortality in different animal species. Toxicity of jatropha as a feedstuff has been studied extensively in several species including mice, rats, goats, sheep, and fish (Adam, 1974; Adam et al., 1975; Ahmed et al., 1979a; Ahmed et al., 1979b; Makkar et al., 1999). This toxicity is mainly due to phorbol esters and curcin (a lectin) (Nesseim et al., 2012). Others substances, such as trypsin inhibitors, lectins and phytate restrict the use of jatropha products in animal feeding. *Jatropha platyphylla* is free of phorbol esters and hence non-toxic. However, its kernels and kernel meal contain trypsin inhibitors, lectin and phytate (Makkar et al., 2012).

Phorbol esters

Jatropha species contain lipophilic tetracyclic diterpenoids called phorbol esters, that are present in all parts of the plant, including the seeds, leaves, stems, roots and flowers. The largest concentration (between 2 and 6 g/kg DM) is in the seed kernel. Non-detoxified kernel meal contains from 1 to 3 g/kg phorbol esters, and the oil 2 to 7 g/kg (DM basis) (Makkar et al., 2012). A detoxification process, for example by combining an alkali (NaOH) and an alcohol (methanol) can remove most or all of the phorbol esters (Makkar et al., 2010b). There are low-phorbol genotypes and species, such as *Jatropha platyphylla*, that may provide a meal where phorbol esters are below the detection level (Makkar et al., 2012).

Phorbol esters are known for their inflammatory action resulting in irritation and toxicity in insect, fish and mammalian species (Goel et al., 2007; Makkar et al., 2009; Nesseim et al., 2012). *Jatropha curcas* contains one of the most potent of the phorbols: phorbol-12-myristate-13-acetate, which is recognized for its tumor promoting activity (as a co-carcinogen) and is used as a biomedical research tool in models of carcinogenesis. Phorbol esters are toxic to livestock even at very low concentrations (Goel et al., 2007). In calves, mortality and severe liver, digestive, kidney and lung lesions were observed when they were fed diets containing only 0.25 g/kg *Jatropha curcas* seeds (Ahmed et al., 1979b). The same occurred in goats fed similar doses of *Jatropha curcas* (Adam et al., 1975). In mice, intra gastric absorption of phorbol esters resulted in acute haemorrhagic gastro-enteritis with LD₅₀ = 27.34 mg/kg live weight (Li et al., 2010).

Curcin

Jatropha contains lectins or haemagglutinins. These proteins represent 6 to 11% of the seed proteins and they play an important role as deterrents to herbivores (Nesseim et al., 2012). In *Jatropha curcas*, the lectin curcin has irritative properties. Curcin belongs to a group of proteins called ribosome inactivating proteins, which inactivate ribosomes and, therefore, reduce protein synthesis (Makkar et al., 2011). However, curcin is also found in edible crops such as spinach, beet root and cereal grains where it does not seem to alter protein metabolism (Nesseim et al., 2012).

Trypsin inhibitors

Jatropha contains trypsin inhibitors that prevent protein digestion and have deleterious effects on growth performance, notably in monogastric animals (Rackis et al., 1981).

Automatic translation

Sélectionner une langue

Feed categories

All feeds

Forage plants

- ▶ Cereal and grass forages
- ▶ Legume forages
- ▶ Forage trees
- ▶ Aquatic plants
- ▶ Other forage plants

Plant products/by-products

- ▶ Cereal grains and by-products
- ▶ Legume seeds and by-products
- ▶ Oil plants and by-products
- ▶ Fruits and by-products
- ▶ Roots, tubers and by-products
- ▶ Sugar processing by-products
- ▶ Plant oils and fats
- ▶ Other plant by-products

Feeds of animal origin

- ▶ Animal by-products
- ▶ Dairy products/by-products
- ▶ Animal fats and oils
- ▶ Insects

Other feeds

- ▶ Minerals
- ▶ Other products

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Saponins

Jatropha curcas seeds contain saponins, which are bitter, have haemolytic activity and are toxic, particularly to fish (Cseke et al., 1999). Raw *Jatropha curcas* kernel meal contains 2-3.4% DM saponins (Makkar et al., 1998; Martinez-Herrera et al., 2006), which is lower than in soybean meal (4.7% DM) and have very mild haemolytic activity (Makkar, personal communication).

Phytates

Phytates are present in jatropha kernel meal (6-10% DM, Makkar et al., 1998; Martinez-Herrera et al., 2006), but their effect can be alleviated with phytase enzymes (Makkar et al., 2012).

Ruminants

Ruminants should be fed only with detoxified jatropha products or with those from non-toxic genotypes or species.

Jatropha leaves

In vitro organic matter digestibility (apparent and true) and metabolisable energy (gas method) for *Jatropha curcas* leaves were respectively 60%, 82% and 7 MJ/kg. These were among the highest estimated values compared to other agricultural by-products, and drought-tolerant plants and browses in Syria (Al-Masry, 2003).

Jatropha seeds

Raw jatropha seeds were found extremely toxic to goats (Adam et al., 1975), sheep and calves (Ahmed et al., 1979a; Ahmed et al., 1979b).

Jatropha kernel meal

Phorbol esters cannot be degraded by rumen microbes and cause severe toxic symptoms in ruminants (Makkar et al., 2012). It is, therefore, advisable to use only detoxified jatropha kernel meal or kernel meals obtained from non-toxic genotypes. Detoxified jatropha kernel meal has been studied as a protein source for goats and lambs and can be used safely, economically and successfully in these species. Jatropha kernel meal from non-toxic genotypes or species have not been investigated in *in vivo* studies for ruminants.

In vitro studies

Based on *in vitro* studies, well-detoxified jatropha kernel meal or meal from non-toxic jatropha genotypes can be used as a good quality protein source in ruminants. Jatropha kernel meal (heated to 121°C at 66% moisture for 30 minutes) from toxic and non-toxic genotypes was found to have lower *in vitro* OM digestibility and metabolizable energy (gas method) than soybean meal (78-77 vs. 88% and 10.9-10.3 vs. 13.3 MJ/kg, respectively). The *in vitro* (pepsin + trypsin) protein digestibility was similar to that of heated soybean meal (89-90 vs. 91%), whereas the 24h *in vitro* rumen protein degradability from toxic and non-toxic jatrophas were much lower than that of soybean meal (43 and 29% vs. 81%). The particularly low value of N degradability of non-toxic jatropha meal suggests that it has substantial amounts of rumen-undegradable protein that can be used post-rationally (Makkar et al., 2009; Makkar et al., 2012). Other studies have shown that *in vitro* organic matter digestibility of jatropha kernel meal was lower than that of soybean meal (49-61 vs. 64% Triastuty et al., 2009) and may be reduced by detoxification processes (from 51 to 61%, Ouskoueian et al., 2011). Moist heating (66% moisture, 121°C for 15 min) is much more effective in inactivating trypsin inhibitor (Makkar et al., 2012).

Detoxification by fungal fermentation improved the protein content by 10-15%. Fungal fermentation also enhanced total lipid and fatty acids (Belewu et al., 2011).

Sheep

In Egypt, detoxified jatropha cake replaced up to 70% of soybean meal in a concentrate feed mixture fed to lambs without any adverse effect on performance. Economic efficiency was slightly improved with the jatropha supplement compared to the control ration (El-Zelaky et al., 2011). Jatropha kernel meal detoxified with lactic acid bacteria replaced up to 50%, and even up to 70%, of soybean meal in concentrate feed mixture without any adverse effect on lamb performance (Abo El Fadel et al., 2011).

Goats

In Brazil, the inclusion of 20% (DM) of fungal detoxified jatropha kernel meal in the diets of growing goats did not result in changes in blood parameters or clinical symptoms of poisoning (Kasuya et al., 2012). In Nigeria, fungal detoxified jatropha kernel meal replaced 50% soybean meal in the diet of West African Dwarf Goats, increasing DM intake, weight gain (1380 vs. 708 g/d) and tissue deposition. Full replacement of soybean meal by jatropha kernel meal was less efficient (weight gain of 860 g/d) (Belewu et al., 2011).

Pigs

Non-detoxified jatropha kernel meal is toxic to pigs. When the detoxification procedure fails to remove completely and/or to neutralize phorbol esters and/or the other toxic anti-nutritional factors, dietary jatropha meal was found to cause severe adverse effects (diarrhea, death, etc.) in growing-finishing pigs (Chivandi et al., 2006). It is recommended to incorporate jatropha kernel meal in pig feeds only when it is certain that the product has been thoroughly detoxified.

Detoxified jatropha kernel meal supplemented with lysine replaced up to 50% of the soybean meal protein in diets for growing pigs without any negative effects on health and growth performance (Haifeng Wang et al., 2011). In finishing pigs, a linear increase of inclusion rate of detoxified and extruded jatropha kernel meal (from 0 to 8%) resulted in a significant reduction in the growth rate of finishing pigs linked to a decreased voluntary feed intake. These effects were attributed to the presence of traces of phorbol esters (Berenchtein et al., 2014).

Poultry

Raw jatropha seeds and kernel meal are toxic to poultry, mainly due to the presence of phorbol esters (Makkar et al., 1997). This acute toxicity can depress growth even at low inclusion rates, and may cause mortality at higher levels (Annongu et al., 2010; Ojo et al., 2013; Sumiati et al., 2011). Several other antinutritional factors present in jatropha are also harmful to poultry (Makkar et al., 1997). Raw jatropha products should, therefore, not be used. It is also not advisable to use jatropha products in poultry feeds, except in the case of industrially detoxified products, following a confirmation trial.

The detoxification of jatropha products has led to better performance in most cases when compared to raw jatropha, but performance was still depressed in broilers (jatropha seeds: [Annongu et al., 2010](#); jatropha kernel meal: [Aguirre Jamarillo, 2011](#); [Pasaribu et al., 2010](#); [Sumiati et al., 2011](#)), and layers ([Sumiati et al., 2012](#)) when compared to the control diets. In some cases the detoxification processes led to an actual improvement and to performance close to that from the control diets ([Granda Paz, 2012](#); [Boguhn et al., 2010](#); [Sumiati et al., 2009](#)). More sophisticated detoxification methods and the use of non-toxic varieties or species of jatropha may facilitate the use of jatropha products in poultry diets in the future.

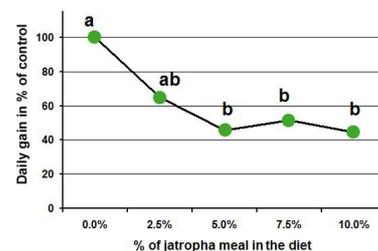
Rabbits

Jatropha kernel meal

The use of *Jatropha curcas* kernel meal is not yet recommended in rabbit feeding. Untreated *Jatropha curcas* kernel meal introduced in rabbits diets resulted in a reduction of feed intake and in a significant growth rate reduction of more than 30%, as shown in the following figure (mean values with different superscripts are significantly different at $P < 0.05$) ([Abdel-Shafy et al., 2011](#)).

Detoxification by fermentation with *Bifidobacterium* (culture for 15 days at 37°C) suppressed haematological changes but feed intake remained significantly reduced, by 28%, when fermented jatropha kernel meal was introduced at 6% or 12% in rabbit diets ([Widiyastuti et al., 2014](#)).

Using jatropha kernel meal may be a non-chemical method to treat ectoparasites in rabbits. Rabbits experimentally infected with the tick *Hyalomma marginatum marginatum* showed a drastic reduction in tick reproduction after being fed a diet containing 10% jatropha kernel meal with no serious effects on rabbits liver and kidney functions ([Nasr et al., 2011](#)), though some haematological alterations were recorded ([Abdel-Shafy et al., 2011](#)).



Jatropha tanjorensis leaves

Dried *Jatropha tanjorensis* leaves introduced in diets for growing rabbits at up to 25% caused erratic growth (both positive and negative weight changes), with no clear relation to the inclusion rate though some haematological parameters were improved ([Orhue et al., 2008](#)). The renal function also improved with a significant reduction of serum urea concentration. However, there were also signs of hepatic toxicity ([Akhigbe et al., 2009](#)). Until proven harmless, *Jatropha tanjorensis* leaves cannot be recommended as a feed ingredient in rabbit nutrition.

Fish

Common carp (*Cyprinus carpio*)

Detoxified jatropha kernel meal

Detoxified jatropha kernel meal replaced 50% of the fish meal in common carp diets without altering health parameters (no changes in blood parameters or haematocrit) and without negative effects on feeding time, palatability or fish growth. Feeding common carp with detoxified jatropha kernel meal increased amino acid availability. Protein digestibility was over 90% and lipid digestibility was in the range of 74-90%. However, lower digestive enzyme activity was reported, possibly due to phytates but this problem could be alleviated by the use of phytase. Feed conversion ratio remained unchanged as well as the protein productive value ([Makkar et al., 2012](#); [Kumar et al., 2011a](#)). Feeding common carp on detoxified jatropha kernel meal resulted in higher lipid deposition that could be due to higher carbohydrates and lower cholesterol contents in plasma and fish muscle ([Kumar et al., 2010](#)). Feeding detoxified jatropha kernel meal also resulted in more whole body crude protein ([Kumar et al., 2011a](#)).

Detoxified jatropha protein isolate

Detoxified protein isolate of *Jatropha curcas* is very palatable to common carp ([Kumar et al., 2012b](#)). Detoxified protein isolate replaced 75% fish meal in carp diets without compromising health or growth, but the best results were obtained when it replaced only 50% fish meal ([Kumar et al., 2012b](#)). A diet containing detoxified jatropha protein isolate (at 75% fish meal replacement level) had protein and energy digestibilities similar to those of the control diet, which may be due to the removal of antinutritional factors during processing and lower non starch polysaccharides. Fish fed on detoxified jatropha protein isolate had greater lipid retention and this was positively correlated with the inclusion rate of the isolate. Protein retention was significantly higher in carp fed detoxified jatropha protein isolate. This could be explained by a particularly well balanced amino acid profile in detoxified jatropha protein isolate ([Kumar et al., 2012b](#)).

Jatropha kernel meal from non-toxic genotypes

Kernel meal produced with non-toxic genotypes of jatropha were used to replace 50% fish meal in the diets of common carp fingerlings. The results obtained with untreated kernel meal (body weight gain, feed conversion ratio, etc.) were statistically similar to those obtained with the control diet though they tended to be lower. Heating kernel meal was found to be useless as it reduced the digestibility of protein and energy due to a Maillard reaction ([Makkar et al., 1999](#)).

Rainbow trout (*Onchorynchus mykiss*)

Detoxified jatropha kernel meal replaced 50% fish meal in rainbow trout without affecting health, growth rate, feed conversion ratio, protein efficiency, protein productive value and energy retention. A higher inclusion rate (62.5% fish meal replacement) resulted in poorer growth, probably due to the high phytate content of jatropha, also a relatively high carbohydrate content not well digested by carnivorous fish or to an insufficient supply of sulphur-containing amino acids (11.3 g/kg vs. a requirement of 13 g/kg). Detoxified jatropha kernel meal reduced cholesterol in plasma and muscle of rainbow trout ([Kumar et al., 2011b](#)).

Nile tilapia (*Oreochromis niloticus* L.)

Heat-treated kernel meal of *Jatropha platyphylla*

Kernel meal of the non-toxic species *Jatropha platyphylla* heated at 121°C for 15 minutes under 66% moisture replaced 50 or 62.5% fish meal in Nile tilapia diets fed for 12 weeks without altering fish health. Most blood parameters were unchanged or remained within normal ranges; haematocrit, red blood cell counts and blood glucose were higher, while cholesterol decreased in plasma. Growth performance of Nile tilapia fed on heated *Jatropha platyphylla* kernel meal was similar to the control. Nutrient utilization, energy expenditure and metabolizable energy were not different from the control diet indicating that heated *Jatropha platyphylla* kernel meal could be fed as a protein feed for Nile tilapia ([Akinleye et al., 2012](#); [Makkar et al., 2011](#);

[Kumar et al., 2012a](#)).

Crustaceans

White leg shrimp (*Litopenaeus vannamei*)

Detoxified jatropha kernel meal successfully replaced 25 or 50% of fish meal in white leg shrimp diets. It had excellent nutrient and energy digestibilities, resulting in higher protein and energy deposition, thus leading to greater shrimp performance. Though cholesterol is necessary to shrimp health and performance, it was shown that feeding hypocholesterolemic detoxified jatropha kernel meal had no deleterious effect on shrimp ([Harter et al., 2011](#))

Datasheet citation

Heuzé V., Tran G., Edouard N., Renaudeau D., Bastianelli D., Lebas F., 2016. *Jatropha (Jatropha sp.) kernel meal and other jatropha products*. Feedipedia, a programme by INRA, CIRAD, AFZ and FAO. <http://www.feedipedia.org/node/620> Last updated on April 15, 2016, 10:17

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Jatropha (*Jatropha* sp.) kernel meal and other jatropha products

Description Nutritional aspects Nutritional tables References

Tables of chemical composition and nutritional value

- Jatropha (*Jatropha* sp.) kernel meal, defatted, detoxified or non-toxic
- Jatropha (*Jatropha* sp.) seed meal, defatted
- Jatropha (*Jatropha* sp.) kernels
- Jatropha (*Jatropha* sp.) shells
- Jatropha (*Jatropha curcas*) leaves, dried

Avg: average or predicted value; SD: standard deviation; Min: minimum value; Max: maximum value; Nb: number of values (samples) used

Jatropha (*Jatropha* sp.) kernel meal, defatted, detoxified or non-toxic

From detoxified *Jatropha curcas*, or non-toxic *J. curcas* genotypes or non-toxic *J. platyphylla*



Main analysis	Unit	Avg	SD	Min	Max	Nb
Dry matter	% as fed	92.4	3.1	87.6	97.1	25
Crude protein	% DM	67.8	4.0	62.4	77.5	35
Crude fibre	% DM	6.2	1.7	3.4	9.1	25
NDF	% DM	13.4	3.5	9.8	20.0	21
ADF	% DM	6.8	1.8	5.3	11.5	9
Ether extract	% DM	1.9	1.8	0.4	5.2	27
Ash	% DM	10.7	1.5	7.9	13.6	35
Starch (polarimetry)	% DM	9.6	1.7	6.8	12.0	16
Total sugars	% DM	3.4	2.7	0.1	8.4	16
Gross energy	MJ/kg DM	18.3	0.6	17.3	19.1	24

Minerals	Unit	Avg	SD	Min	Max	Nb
Calcium	g/kg DM	11.4		8.7	14.0	2
Phosphorus	g/kg DM	21.5		17.9	25.2	2

Amino acids	Unit	Avg	SD	Min	Max	Nb
Alanine	% protein	4.3	0.3	4.0	4.9	8
Arginine	% protein	11.5	0.9	10.4	13.0	8
Aspartic acid	% protein	10.6	1.0	9.3	12.2	8
Cystine	% protein	1.6	0.4	0.8	1.8	6
Glutamic acid	% protein	16.2	0.8	14.7	17.0	8
Glycine	% protein	4.4	0.2	4.2	4.8	8
Histidine	% protein	2.9	0.2	2.7	3.3	7
Isoleucine	% protein	3.9	0.3	3.4	4.3	8
Leucine	% protein	6.7	0.5	5.9	7.2	8
Lysine	% protein	3.4	0.2	3.0	3.6	8
Methionine	% protein	1.6	0.1	1.4	1.7	8
Phenylalanine	% protein	4.4	0.4	3.8	4.9	8
Proline	% protein	4.6	0.5	3.9	5.4	8
Serine	% protein	4.3	1.1	2.4	5.3	8
Threonine	% protein	3.4	0.3	3.1	3.8	8
Tryptophan	% protein	1.1	0.0	1.1	1.1	5
Tyrosine	% protein	3.4	1.0	2.5	4.8	7
Valine	% protein	5.1	1.3	4.0	7.2	8

Secondary metabolites	Unit	Avg	SD	Min	Max	Nb
Tannins (eq. tannic acid)	g/kg DM	0.8	0.8	0.2	1.7	3
Tannins, condensed (eq. catechin)	g/kg DM	0.0	0.0	0.0	0.0	3

Ruminant nutritive values	Unit	Avg	SD	Min	Max	Nb
ME ruminants (gas production)	MJ/kg DM	9.6	0.4	9.1	10.5	8

The asterisk * indicates that the average value was obtained by an equation.

Automatic translation

Sélectionner une langue

Feed categories

- All feeds
- Forage plants
 - Cereal and grass forages
 - Legume forages
 - Forage trees
 - Aquatic plants
 - Other forage plants
- Plant products/by-products
 - Cereal grains and by-products
 - Legume seeds and by-products
 - Oil plants and by-products
 - Fruits and by-products
 - Roots, tubers and by-products
 - Sugar processing by-products
 - Plant oils and fats
 - Other plant by-products
- Feeds of animal origin
 - Animal by-products
 - Dairy products/by-products
 - Animal fats and oils
 - Insects
- Other feeds
 - Minerals
 - Other products

Latin names

- Plant and animal families
- Plant and animal species

Resources

- Broadening horizons
- Literature search
- Image search
- Glossary
- External resources
 - Literature databases
 - Feeds and plants databases
 - Organisations & networks
 - Books
 - Journals

References

Abo El Fadel et al., 2011; Aguirre Jamarillo, 2011; Akinleye et al., 2012; Boguhn et al., 2010; Chivandi et al., 2006; Kumar et al., 2010; Makkar et al., 2011; Martinez-Herrera et al., 2006; Nithyananthama et al., 2013; Ouskoueian et al., 2011

Last updated on 31/07/2014 21:54:51

Jatropha (Jatropha sp.) seed meal, defatted

Includes raw and detoxified seed meals



Main analysis	Unit	Avg	SD	Min	Max	Nb
Dry matter	% as fed	90.1	1.1	88.5	93.3	19
Crude protein	% DM	16.9	4.1	13.7	26.9	19
Crude fibre	% DM	36.1	1.8	33.1	38.3	18
NDF	% DM	44.2				1
ADF	% DM	37.5				1
Ether extract	% DM	6.0	2.8	1.6	13.5	19
Ash	% DM	5.8	0.6	5.2	7.8	19
Gross energy	MJ/kg DM	19.7	2.3	17.7	22.2	3

Minerals	Unit	Avg	SD	Min	Max	Nb
Calcium	g/kg DM	9.3	2.3	7.2	11.8	3
Phosphorus	g/kg DM	8.5	4.2	3.7	11.6	3

Amino acids	Unit	Avg	SD	Min	Max	Nb
Alanine	% protein	5.3				1
Arginine	% protein	9.6				1
Aspartic acid	% protein	8.6				1
Glutamic acid	% protein	18.0				1
Glycine	% protein	3.0				1
Histidine	% protein	3.2				1
Isoleucine	% protein	3.7				1
Leucine	% protein	7.2				1
Lysine	% protein	3.8				1
Methionine	% protein	1.2				1
Phenylalanine	% protein	5.2				1
Serine	% protein	4.0				1
Threonine	% protein	3.2				1
Tyrosine	% protein	2.6				1

The asterisk * indicates that the average value was obtained by an equation.

References

Abd El-Rahman et al., 2011; Abdel-Shafy et al., 2011; Chivandi et al., 2006; Guedes et al., 2014; Sumiati et al., 2009

Last updated on 31/07/2014 21:49:55

Jatropha (Jatropha sp.) kernels



Main analysis	Unit	Avg	SD	Min	Max	Nb
Dry matter	% as fed	95.5	1.1	94.0	96.9	12
Crude protein	% DM	27.9	2.1	25.6	33.1	19
Crude fibre	% DM	3.0	0.3	2.7	3.4	5
NDF	% DM	4.0	0.3	3.5	4.5	11
ADF	% DM	2.8	0.2	2.4	3.0	7
Lignin	% DM	0.1	0.1	0.0	0.2	6
Ether extract	% DM	57.0	0.7	55.5	57.8	18
Ash	% DM	4.3	0.9	3.6	6.8	15
Starch (polarimetry)	% DM	5.5	0.4	4.9	5.9	4
Total sugars	% DM	4.0	0.5	3.4	4.4	4
Gross energy	MJ/kg DM	30.7	0.8	29.1	31.5	11

Secondary metabolites	Unit	Avg	SD	Min	Max	Nb
Tannins (eq. tannic acid)	g/kg DM	0.2	0.2	0.0	0.4	4

Tannins, condensed (eq. catechin) g/kg DM 0.0 0.0 0.0 0.0 4

The asterisk * indicates that the average value was obtained by an equation.

References

Aderibigbe et al., 1997; Makkar et al., 1998; Makkar et al., 2009; Makkar et al., 2011; Martinez-Herrera et al., 2006; Ojo et al., 2013; Ouskoueian et al., 2011

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Jatropha (*Jatropha* sp.) shells



Main analysis	Unit	Avg	SD	Min	Max	Nb
Dry matter	% as fed	90.5	0.7	89.8	91.9	6
Crude protein	% DM	4.6	0.6	4.3	5.8	6
NDF	% DM	86.4	2.5	83.9	89.6	6
ADF	% DM	76.4	2.1	74.6	79.8	6
Lignin	% DM	46.4	1.2	45.1	47.5	6
Ether extract	% DM	0.9	0.4	0.5	1.4	6
Ash	% DM	5.3	1.3	2.8	6.1	6
Gross energy	MJ/kg DM	19.4	0.1	19.3	19.5	6

Secondary metabolites	Unit	Avg	SD	Min	Max	Nb
Tannins (eq. tannic acid)	g/kg DM	23.3	3.9	20.0	29.0	4
Tannins, condensed (eq. catechin)	g/kg DM	0.0	0.0	0.0	0.0	4

Ruminant nutritive values	Unit	Avg	SD	Min	Max	Nb
OM digestibility, ruminants (gas production)	%	27	0	26	27	4
ME ruminants (gas production)	MJ/kg DM	2.5	0.2	2.4	2.8	4

The asterisk * indicates that the average value was obtained by an equation.

References

Aderibigbe et al., 1997; Makkar et al., 1998

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Jatropha (*Jatropha curcas*) leaves, dried



Main analysis	Unit	Avg	SD	Min	Max	Nb
Dry matter	% as fed	94.0				1
Crude protein	% DM	15.8				1
Crude fibre	% DM	15.3				1
NDF	% DM	33.3				1
ADF	% DM	28.8				1
Ether extract	% DM	5.3				1
Ash	% DM	21.8				1
Gross energy	MJ/kg DM	16.2				*

Ruminant nutritive values	Unit	Avg	SD	Min	Max	Nb
ME ruminants (gas production)	MJ/kg DM	6.9				1

The asterisk * indicates that the average value was obtained by an equation.

References

Al-Masry, 2003

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Datasheet citation

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Jatropha (*Jatropha* sp.) kernel meal and other jatropha products

Description Nutritional aspects Nutritional tables References

References

- Abd El-Rahman, H. H. ; Abedo, A. A. ; Salman, F. M. ; Mohamed, M. I. ; Shoukry, M. M., 2011. Partial substitution of cumin seed meal by *Jatropha* meal as a potential protein source for feed. *African J. Biotech.*, 10 (68): 15456-15461
- Abdel Gadir, W. S. ; Onsa, T. O. ; Ali, W. E. M. ; El Badwi, S. M. A. ; Adam, S. E. I., 2003. Comparative toxicity of *Croton macrostachys*, *Jatropha curcas* and *Piper abyssinica* seeds in Nubian goats. *Small Rum. Res.*, 48 (1): 61-67
- Abdel-Shafy, S. ; Nasr, S. M. ; Abdel-Rahman, H. H. ; Habeeb, S. M., 2011. Effect of various levels of dietary *Jatropha curcas* seed meal on rabbits infested by the adult ticks of *Hyalomma marginatum marginatum* I. Animal performance, anti-tick feeding and haemogram. *Trop. Anim. Health Prod.*, 43 (2): 347-357
- Abo El-Fadel, M. H. ; Hussein, A. M. ; Mohamed, A. H., 2011. Incorporation *Jatropha Curcas* meal on lambs ration and it's effect on lambs performance. *Nature & Sci.*, 7 (2): 129-132
- Adam, S. E. I. ; Magzoub, M., 1975. Toxicity of *Jatropha curcas* for goats. *Toxicology*, 4 (3): 347-354
- Adam, S. E. I., 1974. Toxic effects of *Jatropha curcas* in mice. *Toxicology*, 2 (1): 67-76
- Aderibigbe, A. O. ; Johnson, C. O. L. E. ; Makkar, H. P. S. ; Becker, K. ; Foidl, N., 1997. Chemical composition and effect of heat on organic matter- and nitrogen-degradability and some antinutritional components of jatropha meal. *Anim. Feed Sci. Technol.*, 67 (2-3): 223-243
- Aguirre Jaramillo, R. J., 2011. Use of the meal residue of oil extraction of the *Jatropha* (*Jatropha curcas*), for the improvement of feeding of broiler chicks 0-21 days in Pronaca S.A. Cantón Quito company. Tesis de ingeniero, Universidad de las americas, Quito, Ecuador
- Ahmed, O. M. M., Adam, S. E. I., 1979. Toxicity of *Jatropha curcas* in sheep and goats. *Research Vet. Sci.*, 27 (1): 89-96
- Ahmed, O. M. M.; Adam, S. E. I., 1979. Effects of *Jatropha curcas* on calves. *Vet. Pathol.*, 16 (4): 476-482
- Akhigbe, A. O. ; Idu, M. ; Orhue, E. S. ; Ataman, J. E. ; Ehimwenman, S. O., 2009. Effect of *Jatropha tanjorensis* J.L. Ellis and Soroja leaves in rabbits: biochemistry and ultrasonography. *Res. J. Med. Plant*, 3 (1): 29-33
- Akinleye, A. ; Kumar, V. ; Makkar, H. P. S. ; Angulo-Escalante, M. A. ; Becker, K., 2012. *Jatropha platyphylla* kernel meal as feed ingredient for Nile tilapia (*Oreochromis niloticus* L.): growth, nutrient utilization and blood parameters. *J. Anim. Physiol. Anim. Nutr.*, 96 (1): 119-129
- Al-Masri, M. R., 2003. An *in vitro* evaluation of some unconventional ruminant feeds in terms of the organic matter digestibility, energy and microbial biomass. *Trop. Anim. Health Prod.*, 35 (2): 155-167
- Annonu, A. A.; Belew, M. A.; Joseph, J. K., 2010. Potentials of *Jatropha* seeds as substitute protein in nutrition of poultry. *Res. J. Anim. Sci.*, 4 (1): 1-4
- Axelsson, L. ; Franzén, M. ; Ostwald, M. ; Berndes, G. ; Ravindranath, N., 2011. Performance of *Jatropha* biodiesel production and its environmental and socio-economic impact – A case study in Southern India. *Linköping Electronic Conference Proceedings*, 57 (10): 2470-2477
- Barri, M. E. S. ; Onsa, T. O. ; Elawad, A. A. ; Elsayed, N. Y. ; Wasfi, I. A. ; Bari, E. M. A. ; Adam, S. E. I., 1983. Toxicity of five Sudanese plants to young ruminants. *J. Comp. Pathol.*, 93 (4): 559-575
- Belew, M. A. ; Belew, K. Y. ; Lawal, I. A., 2011. Cocktail of fungi blend on *Jatropha curcas* kernel cake: effect on feed intake and blood parameters of goat. *Libyan Agric. Res. Cent. J. Int.*, 2 (3): 138-143
- Berenchtein, B. ; Abdalla, A. L. ; Paim, T. do P. ; Sbardella, M. ; Louvandini, H. ; Filho, A. L. A. ; Dhanasekaran, D. ; Santos, P. P. dos, 2014. Effects of detoxified *Jatropha curcas* kernel meal in finishing pig diets on their performance, carcass traits, meat quality and intoxication. *Livest. Sci.*, 118 (1/2): 157-165
- Boguhn, J.; Rodehutschord, M.; Makkar, H. P. S.; Becker, K., 2010. Amino acid digestibility of detoxified *Jatropha* kernel meal in turkeys. Abstract book of the 13th European Poultry Conference, August 23 - 27, 2010, Tours, France. *World Poult. Sci. J.*, 66 (Supplement), 476
- Brittaine, R.; NeBambi Lutaladio, 2010. *Jatropha*: a small-holder bioenergy crop; The potential for pro-poor development. *Integrated Crop Management n°8*. IFAD / FAO
- Chivandi, E. ; Mtimuni, J. P. ; Read, J. S. ; Makuza, S. M., 2004. Effect of processing method on phorbol ester concentration, total phenolics, trypsin inhibitor activity and the proximate composition of the Zimbabwean *Jatropha curcas* provenance: A potential livestock feed. *Pakistan J. Biol. Sci.*, 7 (6): 1001-1005
- Chivandi, E. ; Erlwanger, K. H. ; Makuza, S. M. ; Read, J. S. ; Mtimuni, J. P., 2006. Effects of dietary *Jatropha curcas* meal on percent packed cell volume, serum glucose, cholesterol and triglyceride concentration and alpha-amylase activity of weaned fattening pigs. *Res. J. Anim. Vet. Sci.*, 1 (1): 18-24
- Cseke, L. J. ; Kaufman, P. B. ; Warber, S. ; Duke, J. A. ; Briemann, H. L., 1999. *Natural products from plants*. CRC Press LLC., Boca Raton, USA.
- de Barros, C. R. M. ; Ferreira, M. M. L. ; Nunes, F. M. ; Bezerra, R. M. F. ; Dias, A. A. ; Guedes, C. V. ; Cone, J. W. ; Marques, G. S. M. ; Rodrigues, M. A. M., 2011. The potential of white-rot fungi to degrade phorbol esters of *Jatropha curcas* L. seed cake. *Eng. Life Sci.*, 11 (1): 107-110
- EFSA, 2015. Scientific opinion on the Risks for human and animal health related to the presence of phorbol esters in *Jatropha* kernel meal. EFSA Panel on Contaminants in the Food Chain (CONTAM), *EFSA J.*, 13 (12): 4321
- EFSA, 2015. Risks for human and animal health related to the presence of phorbol esters in *Jatropha* kernel meal. EFSA Panel on Contaminants in the Food Chain (CONTAM); *EFSA Journal*, 13 (12): 4321-4401
- El-Zelaky, O. A. ; Khalifa, E. I. ; Mohamed, A. H. ; Bahera, K. M. ; Hussein, A. M., 2011. Productive and reproductive

Automatic translation

Sélectionner une langue

Feed categories

All feeds

Forage plants

- ▶ Cereal and grass forages
- ▶ Legume forages
- ▶ Forage trees
- ▶ Aquatic plants
- ▶ Other forage plants

Plant products/by-products

- ▶ Cereal grains and by-products
- ▶ Legume seeds and by-products
- ▶ Oil plants and by-products
- ▶ Fruits and by-products
- ▶ Roots, tubers and by-products
- ▶ Sugar processing by-products
- ▶ Plant oils and fats
- ▶ Other plant by-products

Feeds of animal origin

- ▶ Animal by-products
- ▶ Dairy products/by-products
- ▶ Animal fats and oils
- ▶ Insects

Other feeds

- ▶ Minerals
- ▶ Other products

Latin names

Plant and animal families

Plant and animal species

Resources

Broadening horizons

Literature search

Image search

Glossary

External resources

- ▶ Literature databases
- ▶ Feeds and plants databases
- ▶ Organisations & networks
- ▶ Books
- ▶ Journals

- performance of Rahmani male lambs fed rations containing *Jatropha* cake. *Egyptian J. Sheep & Goat Sci.*, 6 (2): 15-24
- Elangovan, A. V. ; Gowda, N. K. S. ; Satyanarayana, M. L. ; Suganthi, R. U. ; Rao, S. B. N. ; Sridhar, M., 2013. *Jatropha* (*Jatropha curcas*) seed cake as a feed ingredient in the rations of sheep. *Anim. Nut. Feed. Technol.*, 13 (1): 57-67
- Elbehri, A.; Segerstedt, A.; Liu, P., 2013. A global assessment of sustainability Biofuels and sustainability challenge : issues, trends and policies for biofuels and related feedstocks. FAO, Rome, Italy
- Gaur, S., 2009. Development and evaluation of an effective process for the recovery of oil and detoxification of meal from *Jatropha curcas*. MSc. Thesis, Missouri University of Science and Technology, USA
- Goel, G. ; Makkar, H. P. S. ; Francis, G. ; Becker, K., 2007. Phorbol esters: Structure, biological activity, and toxicity in animals. *Int. J. Toxicol.*, 26 (4): 279-288
- Granda Paz, V. A., 2012. Formulation of an optimal diet for growing broilers base on the bioconversion of jatropha oilmeal (*Jatropha curcas*) with fibrolytic enzymes. ESPE university, Dept. Ciencias de la vida, Sangolqui, Ecuador
- Guedes, R. E. ; Cruza, F. de A. ; Lima, M. C. de ; Sant'Ana, L. d'O.; Castro, R. N., Mendes, M. F., 2014. Detoxification of *Jatropha curcas* seed cake using chemical treatment: analyses with a central composite rotatable design. *Ind. Crops Products*, 52: 673-679
- Haifeng Wang ;Yi Chen ; Yi'nuo Zhao ; Hongyun Liu ; Jianxin Liu; Harinder P. S. Makkar; Klaus Becker, 2011. Effects of replacing soybean meal by detoxified *Jatropha curcas* kernel meal in the diet of growing pigs on their growth, serum biochemical parameters and visceral organs. *Anim. Feed Sci. Technol.*, 170 (1-2): 141-146
- Harter, T. ; Buhrke, F. ; Kumar, V. ; Focken, U. ; Makkar, H. P. S. ; Becker, K., 2011. Substitution of fish meal by *Jatropha curcas* kernel meal: effects on growth performance and body composition of white leg shrimp (*Litopenaeus vannamei*). *Aquacult. Nutr.*, 17 (5): 542-548
- Henning, R. K., 2009. The *Jatropha* System – an integrated approach of rural development. In: Henning, The *Jatropha* Book, Weissenberg, Germany
- Joshi, C. ; Mathur, P. ; Khare, S. K., 2011. Degradation of phorbol esters by *Pseudomonas aeruginosa* PseA during solid-state fermentation of deoiled *Jatropha curcas* seed cake. *Biores. Technol.*, 102 (7): 4815-4819.
- Kasuya, M. C. M. ; Luz, J. M. R. da ; Pereira, L. P. da Silva ; Silva, J. S. da ; Montavani, H. C. ; Rodrigues, M. T., 2012. Bio-detoxification of jatropha seed cake and its use in animal feed: chapter 11. In: Fang, Z. (Ed.). *Biodiesel - Feedstocks, Production and Applications*. pp. 309-330.
- King, A. J. ; He, W. ; Cuevas, J. A. ; Freudenberger, M. ; Ramiaramananana, D. ; Graham, I. A., 2009. Potential of *Jatropha curcas* as a source of renewable oil and animal feed. *J. Exp. Bot.*, 60 (10): 2897-2905
- Kumar, V. ; Makkar, H. P. S. ; Amselgruber, W. ; Becker, K., 2010. Physiological, haematological and histopathological responses in common carp (*Cyprinus carpio* L.) fingerlings fed with differently detoxified *Jatropha curcas* kernel meal. *Food Chem. Toxicol.*, 48 (8-9): 2063-2072.
- Kumar, V. ; Makkar, H. P. S. ; Becker, K., 2011. Detoxified *Jatropha curcas* kernel meal as a dietary protein source: Growth performance, nutrient utilization and digestive enzymes in common carp (*Cyprinus carpio* L.) fingerlings. *Aquacult. Nutr.*, 17 (3): 313-326
- Kumar, V. ; Makkar, H. P. S. ; Becker, K., 2011. Nutritional, physiological and haematological responses in rainbow trout (*Oncorhynchus mykiss*) juveniles fed detoxified *Jatropha curcas* kernel meal. *Aquacult. Nutr.*, 17 (4): 451-467
- Kumar, V. ; Akinleye, A. ; Makkar, H. P. S. ; Angulo-Escalante, M. A. ; Becker, K., 2012. Growth performance and metabolic efficiency in Nile tilapia (*Oreochromis niloticus* L.) fed a diet containing *Jatropha platyphylla* kernel meal as a protein source. *J. Anim. Physiol. Anim. Nutr.*, 96 (1): 37-46
- Kumar, V. ; Makkar, H. P. S. ; Becker, K., 2012. Evaluations of the nutritional value of *Jatropha curcas* protein isolate in common carp (*Cyprinus carpio* L.). *J. Anim. Physiol. Anim. Nutr.*, 96 (6): 1030-1043
- Li CaiYan ; Devappa, R. K. ; Liu JianXin ; Lu JianMin ; Makkar, H. P. S. ; Becker, K., 2010. Toxicity of *Jatropha curcas* phorbol esters in mice. *Food Chem. Technol.*, 48 (2): 620-625
- Makkar, H. P. S. ; Becker, K. , 1997. *Jatropha curcas* toxicity: identification of toxic principles. In: Garland, T. ; Barr, A. C. (Eds.) 5th Int. Symp. on Poisonous Plants, San Angelo, Texas, USA, 19-23 May 1997
- Makkar, H. P. S. ; Aderibigbe, A. O. ; Becker, K., 1998. Comparative evaluation of a non-toxic and toxic variety of *Jatropha curcas* for chemical composition, digestibility, protein degradability and toxic factors. *Food Chem.*, 62 (2): 207-215
- Makkar, H. P. S. ; Becker, K., 1999. Nutritional studies on rats and fish (carp, *Cyprinus carpio*) fed diets containing unheated and heated *Jatropha curcas* meal of a non-toxic provenance. *Plant Foods Human Nut.*, 53 (3): 183-192.
- Makkar, H. P. S. ; Francis, G. ; Becker, K., 2007. Bioactivity of phytochemicals in some lesser-known plants and their effects and potential applications in livestock and aquaculture production systems. *Animal*, 1 (9): 1371-1391
- Makkar, H. P. S. ; Becker, K., 2009. *Jatropha curcas*, a promising crop for the generation of biodiesel and value-added coproducts. *Eur. J. Lip. Sci. Technol.*, 111 (8): 773-787
- Makkar, H. P. S. ; Becker, K., 2010. Are *Jatropha curcas* phorbol esters degraded by rumen microbes?. *J. Sci. Food Agric.*, 90 (9):1562-1565
- Makkar, H. P. S. ; Becker, K., 2010. Method for detoxifying plant constituents. WIPO (World Intellectual Property Office), WO/2010/092143
- Makkar, H. P. S. ; Kumar, V. ; Oyeleye, O. O. ; Akinleye, A. O. ; Angulo-Escalante, M. A. ; Becker, K., 2011. *Jatropha platyphylla*, a new non-toxic *Jatropha* species: Physical properties and chemical constituents including toxic and antinutritional factors of seeds. *Food Chem.*, 125 (1): 63-71
- Makkar, H. P. S. ; Kumar, V. ; Becker, K., 2012. Use of detoxified jatropha kernel meal and protein isolate in diets of farm animals: chapter 21. In: Makkar, H.P.S, *Biofuel co-products as livestock feed - opportunities and challenges*, FAO
- Makkar, H. P. S., 2016. State-of-the-art on detoxification of *Jatropha curcas* products aimed for use as animal and fish feed: A review article. *Anim. Feed Sci. Technol.*, 222: 87-99
- Martinez-Herrera, J. ; Siddhuraju, P. ; Francis, G. ; Davila-Ortiz, G. ; Becker K. , 2006. Chemical composition, toxic/anti-metabolic constituents, and effect of different treatments on their levels, in four provenances of *Jatropha curcas* L. from Mexico. *Food Chem.*, 96 (1): 80-89
- Nasr, S. M. ; Abdel-Shafy, S. ; Abdel-Rahman, H. H.; Habeeb, S. M., 2011. Effect of dietary *Jatropha curcas* seed meal on rabbits infested by adult ticks of *Hyalomma marginatum marginatum*. *Toxicol. Environ. Chem.*, 93 (8): 1693-1708
- Nesseim, T. D. T. ; Fillet, M. ; Mergeau, G. ; Dieng, A. ; Hornick, J-L., 2012. Toxic principles, toxicity and technology

of detoxification of *Jatropha curcas* seed: a review. *Base*, 16 (4): 531-540

Nithiyanthama, S. ; Siddhuraju, P. ; Francis, G. , 2013. Proximate composition and functional properties of raw and processed *Jatropha curcas* L. kernel meal. *Int. J. Res. Pharm. Biomed. Sci.*, 4 (1): 183-195

Ojo, R. J.; Oguche, P. I.; Kube, G. D.; Udzer, T. E., 2013. Effect of *Jatropha curcas* supplemented diet on broilers. *Scholars Acad. J. Biosci.*, 1 (6):329-336

Oliveira, A. S. de ; Schwambach, T. I. ; Sinhorin, A. P. ; Oliveira, M. R. C. ; Alessi, K. A. ; Filho, F. A. de O. ; Pina, D. dos S., 2012. Capacity of ensilage of *Jatropha curcas* L. cake to degrade phorbol esters. *Rev. Bras. Zoot.*, 41 (6): 1545-1549

Orhue, E. S. I. ; Idu, M. ; Ataman, J. E. ; Ebite, L. E., 2008. Haematological and histopathological studies of *Jatropha tanjorensis* (J. L. Ellis and Soroja) leaves in rabbits. *Asian J. Biol. Sci.*, 1 (2): 84-89

Ouskoueian, E. ; Abdullah, N. ; Saad, W. S. ; Omar, A. R. ; Bin Puteh, M. ; Yin Wan Ho, 2011. Anti-nutritional metabolites and effect of treated *Jatropha curcas* kernel meal on rumen fermentation *in vitro*. *J. Anim. Vet. Adv.*, 10 (2): 214-220

Panigrahi, S. ; Francis, B. J. ; Cano, L. A. ; Burbage, M. B., 1984. Toxicity of *Jatropha curcas* seeds from Mexico to rats and mice. *Nutr. Rep. Int.*, 29 (5): 1089-1099

Pasaribu, T.; Wina, E.; Tangendjaja, B.; Iskandar, S., 2010. Performance of broiler chicken fed physically and chemically treated jatropha (*Jatropha curcas*) seed meal. *Indon. J. Agric.*, 3 (2): 121-126

Pighinelli, A. L. M. T.; Ferrari, R. A. ; Machado, M. C. N. A. ; Park, K. J., 2012. Study of *Jatropha curcas* L. cake detoxification. Energy, biomass & biol. residues, *Int. Conf. Agric. Eng., CIGR-AgEng 2012. Agric. for a healthier Life*, Valencia, Spain, 8-12 July 2012

Promode Kant, ShuiRong Wu, 2011. The extraordinary collapse of *Jatropha* as a global biofuel. *Environ. Sci. Technol.*, 45 (17): 7114–7115

Rackis, J. J. ; Gumbmann, M. R., 1981. Protease inhibitors: physiological properties and nutritional significance. In: Ory, R. L. (Ed.), *Antinutrients and natural toxicants in foods*. Westport, CT, USA: Food and Nutrition Press

Raheman, H., 2012. *Jatropha*: chapter 14. In: Chittaranjan, K; Joshi, C. P. ; Shonnard, D. R. (Eds). *Handbook of bioenergy crop plants*. CRC Press, Taylor & Francis Group

Rakshit, K. D. ; Bhagya, S., 2007. Effect of processing methods on the removal of toxic and anti-nutritional constituents of *Jatropha* meal: A potential protein source. *J. Food Sci. Technol.*, 3: 88-95

Reubens, B. ; Achten, W. M. J. ; Maes, W. H. ; Danjon, F. ; Aerts, R. ; Poesen, J. ; Muys, B., 2011. More than biofuel? *Jatropha curcas* root system symmetry and potential for soil erosion control. *J. Arid Env.*, 75 (2): 201-205

Sadubthummarak, U. ; Parkpian, P. ; Ruchirawat, M. ; Kongchum, M. ; Delaune, R. D., 2013. Potential treatments to reduce phorbol esters levels in jatropha seed cake for improving the value added product. *J. Env. Sci. and Health. Part B, Pest., Food Cont. Agricult. Wastes*, 48 (11): 974-982

Sumiati; Yusriani, Y. ; Astuti, D. A. ; Suharti, S., 2009. Feeding fermented *Jatropha curcas* L. meal supplemented with cellulase and phytase to kampung chicken. *The 1st International Seminar on Animal Industry, Bogor (Indonesia)*, 23-24 November 2009

Sumiati; Farhanuddin; Hermana, W.; Sudarman, A.; Istichomah, N.; Setiyono, A., 2011. Broiler performances fed diet contained *Jatropha curcas* L. meal fermented with *Rhizopus oligosporus*. *J. Anim. Sci. Technol.*, 34 (2): 117-125

Sumiati; Mutia, R.; Darmansyah, A., 2012. Performance of layer hen fed fermented *Jatropha curcas* L. meal supplemented with cellulose and phytase enzyme. *J. Indon. Trop. Agric.* 37 (2): 108-114

Tjakradidjaja, A. S.; Wiryawan, K. G. ; Afrianti, M., 2012. Fermentability and digestibility of ration containing crude curcumin extract of *Jatropha curcas* seed meal. *2nd Int. Sem. Anim. Ind.*, Jakarta, 2012 : 406 - 412

Triastuty, L. G.; Despal; Permana, I. G., 2009. Nutritional properties of three different origins of Indonesian jatropha (*Jatropha curcas*) meal for ruminant. *J. Agric. Rur. Dev. Tropics & Subtrop.*, Supplement 90: 94-101

Widiyastuti, T. ; Indrajai, M. ; Wibowo, A. ; Hendroko, R., 2014. Biorefinery of jatropha seed cake by lactic acid bacteria and the effects on hematological profile of rex rabbit. *Renew. En. En. Conv. Conf. Exh.*, August 21st – 23rd, 2013 Jakarta, Indonesia., Communication.

78 references found

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