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Evolution of the banana genome (*Musa acuminata*) is impacted by large chromosomal translocations

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Abstract

Most banana cultivars are triploid seedless parthenocarpic clones derived from hybridization between *Musa acuminata* subspecies and sometimes *M. balbisiana*. *M. acuminata* subspecies were suggested to differ by a few large chromosomal rearrangements based on chromosome pairing configurations in inter-subspecies hybrids. We searched for large chromosomal rearrangements in a seedy *M. acuminata* ssp. *malaccensis* banana accession through mate-pair sequencing, BAC-FISH, targeted PCR and marker (DArTseq) segregation in its progeny. We identified a heterozygous reciprocal translocation involving two distal 3 Mb and 10 Mb segments from chromosomes 01 and 04, respectively, and showed that it generated high segregation distortion, reduced recombination and linkage between chromosomes 01 and 04 in its progeny. The two chromosome structures were found to be mutually exclusive in gametes and the rearranged structure was preferentially transmitted to the progeny. The rearranged chromosome structure was frequently found in triploid cultivars but present only in wild *malaccensis* ssp. accessions, thus suggesting that this rearrangement occurred in *M. acuminata* ssp. *malaccensis*. We propose a mechanism for the spread of this rearrangement in *Musa* diversity and suggest that this rearrangement could have played a role in the emergence of triploid cultivars.

Key words: *Musa*, chromosome, translocation, segregation distortion, mate-pair sequencing, genotyping by sequencing

Introduction

Speciation is considered to be the consequence of population divergence due to halted gene flow (Dobzhansky 1937; Mayr 1942). Mechanisms that limit gene flow could involve prezygotic reproductive isolation, which prevents hybrid zygote formation, or postzygotic isolation, which occurs after mating and results in decreased hybrid fitness (Ramsey et al. 2003). Postzygotic reproductive isolation is often associated with speciation genes and/or chromosomal rearrangements (see review by Rieseberg and Blackman 2010). The causal or incidental accumulation of structural variations in relation to speciation events is a matter of debate, with Rieseberg (2001) arguing that large structural variations reduce gene flow more by suppressing recombination and extending the effect of linked isolation genes than by reducing fitness. Regardless of the mechanism, large chromosome structural variations generally cause chromosomal segregation distortion and/or recombination reduction in hybrids (*e.g.* Tadmor et al. 1987; Quillet et al. 1995; Jáuregui et al. 2001; Ostberg et al. 2013), therefore reducing fertility and gene flow. Mechanisms limiting gene flow can be a constraint for breeding programs that exploit genetic resources with the aim of enhancing crop diversity. Conversely, mechanisms reducing fertility have been important factors in the domestication of plants such as bananas with seedless edible fruit.

The *Musa* genus generates bananas, a major starchy staple food and cash crop in tropical and subtropical regions (Lescot 2014), while also providing a valuable model for studying chromosomal rearrangements. Most banana cultivars are derived from *Musa acuminata* (2n=2x=22, A genome), sometimes combined with *Musa balbisiana* (2n=2x=22, B genome). *M. acuminata* is divided into six to nine subspecies (*banksii*, *burmannica*, *malaccensis*, *microcarpa*, *zebrina*, *burmannicoïdes*, *truncata*, *siamea* and *errans*) which diverged following geographical isolation in distinct Southeast Asian continental regions and islands (Daniells 2001; Perrier et al. 2009). The currently accepted domestication scenario suggests that human migrations, probably during the Holocene, led to contacts between these subspecies through the transport of plant material (Perrier et al. 2011). This resulted in the emergence of inter-subspecific hybrids with reduced fertility (Dodds and Simmonds 1948; Fauré et al. 1993a; Shepherd 1999). Early farmers would then have selected parthenocarpic diploid and triploid hybrids producing fruit with high flesh and low seed content.

Cytogenetic studies have shown that chromosomal pairing at meiosis in *Musa acuminata* is generally regular in bivalents within subspecies, but irregular with some multivalents and univalents in hybrids between subspecies (Dodds 1943; Dodds and Simmonds 1948; Dessauw 1987; Fauré et al. 1993a; Shepherd 1999). Chromosomal structural variations between subspecies have been put forward to explain those irregularities. Based on pairing configurations in intersubspecific hybrids, Shepherd (1999) suggested the presence of seven translocation groups, differing from each other by 1 to 4 translocations.

These groups only partly overlap with subspecies delimitation. The Standard group (ST) is the largest one, consisting of *banksii*, *microcarpa* and *malaccensis* spp. accessions. The other groups were named according to the geographic origins of their representatives. The Northern Malayan group (NM) includes some *malaccensis* accessions, the Northern 1 group includes some *burmannicoïdes* and *siamea* accessions, the Northern 2 group includes different *burmannica* and *siamea* accessions, the Malayan Highland group is based on one *truncata* accession, the Javanese group is based on two *zebrina* accession, while the East African group is based on one unclassified accession. Overall, only a limited number of accessions have been studied by cytogenetics and hence little is currently known about the distribution and exact nature of these translocations in *Musa* germplasm.

Genetic mapping studies involving *Musa acuminata* highlighted substantial segregation distortions involving a few linkage groups (Fauré et al. 1993b; Hippolyte et al. 2010; D'Hont et al. 2012; Mbanjo et al. 2012). Chromosomal pairing at meiosis observed in some of the parents has always been irregular, suggesting the presence of chromosomal structural heterozygosity associated with these segregation distortions (Fauré et al. 1993b; Hippolyte et al. 2010). However, no direct links between segregation distortions and the nature of the structural heterozygosity have been established and no large structural variations have been precisely characterized so far in *Musa*.

The aim of the present study was to characterize large structural variations in *Musa* and their impact on chromosome segregation and *Musa* evolution based on the recent availability of a reference genome sequence assembly for *M. acuminata* (D'Hont et al. 2012), while taking advantage of new sequencing potential offered by next-generation sequencing (NGS) technology. We thus focused on a *M. acuminata* ssp. *malaccensis* accession (PT-BA-00267) originally used to produce a genetic map to anchor the *M. acuminata* reference sequence assembly to *Musa* chromosomes and that displayed 17% skewed markers with a high concentration in linkage groups corresponding to chromosomes 01 and 04 (D'Hont et al. 2012).

We sequenced the *M. a.* ssp. *malaccensis* accession PT-BA-00267 through mate-pair sequencing and developed bioinformatics tools to interpret the detected discordant mapping sequences relative to the *M. acuminata* reference sequence. In addition, we refined the analysis of chromosome segregation in the self-progeny of this accession through genotyping by sequencing (DArTseq) and in a biparental cross using SSR markers. A large reciprocal translocation at a heterozygous state was identified in this accession, validated and accurately characterized through PCR and BAC-FISH experiments. The impact of this translocation on chromosome recombination and transmission was also measured and its distribution in accessions representative of *Musa acuminata* germplasm was investigated, enabling us to put forward hypotheses regarding its origin.

Results

Evidence for a large translocation in the PT-BA-00267 accession

Marker segregation analysis revealed high distortion involving two regions on chromosomes 1 and 4

A total of 180 self-progeny individuals of the PT-BA-00267 accession were genotyped using DArTseq technology generating 9,968 SNPs, 7,417 of which were kept after the filtering steps. The markers were well distributed along the 11 reference chromosomes of the *M. acuminata* reference genome assembly, with an average of 1.9 markers per 100 kb (Supporting Information, Table S1) and, as expected, a much higher marker density in gene-rich regions than in repeat-rich pericentromeric regions (Supporting Information, Fig. S1).

The average recombination rate was 0.045 recombinations per Mb (Supporting Information, Table S1), representing around one recombination event per chromosome arm per meiosis. The recombination rate was in general positively correlated with the gene density, with the exception of two gene-rich regions that showed a very limited recombination rate: a 4 Mb region in the median part of metacentric reference chromosome 04 (24.5 Mb to 28.5 Mb) and a 7.5 Mb distal region of the acrocentric reference chromosome 01 (0 Mb to 7.5 Mb) (Supporting Information, Fig. S1). No recombination events were observed between 0 to 2.9 Mb for reference chromosome 01.

Overall, 24% of the markers deviated from the expected Mendelian ratio (0.25: 0.5: 0.25) ($\chi 2$ test, $p < 0.005$) (Supporting Information, Table S1). These markers were mainly located on chromosomes 01 and 04, which each exhibited a large region with very high segregation distortion (Fig. 1, Supporting Information, Fig. S1). Reference chromosomes 01 and 04 displayed 100% and 63% of distorted markers, respectively. Regions of chromosomes 01 and 04 with reduced recombination showed the highest distortion. The segregation bias consisted in an excess of one homozygous genotype (53% observed vs. 25% expected) at the expense of the alternative homozygous (8% vs. 25%) and heterozygous genotypes (39% vs. 50%).

Several clustered markers from reference chromosomes 01 and 04 appeared to be highly linked, with a linkage intensity similar to that observed for physically close markers belonging to one reference chromosome (Fig. 1). These markers belonged to the distal region of acrocentric chromosome 01 and a pericentromeric region of chromosome 04 in the reference assembly. These two regions corresponded to those showing both high segregation distortion and a low recombination rate (Fig. 1).

5 kb mate-pair analysis suggested the presence of a heterozygous reciprocal translocation

Evidence of a structural variation involving chromosomes 01 and 04 was sought based on mapping of 5 kb paired-end reads of the PT-BA-00267 accession on the *M. acuminata* reference sequence assembly and analysis of discordant paired reads. A significant cluster of discordant paired reads was found linking the regions of reference chromosomes 01 and 04 that displayed high distortion and low recombination (Fig. 2a). This linkage, supported by 150 read pairs, suggested a translocation in the PT-BA-00267 accession linking a distal region of reference chromosome 04 (at position 26.7 Mb) to a distal region of reference chromosome 01 (at position 2.9 Mb). Concordant mapped paired reads overlapping the two translocation breakpoints were also observed (Fig. 2b), indicating that the structural variation was at a heterozygous state in the PT-BA-00267 accession.

Analysis of discordant paired reads revealed only one discordant cluster linking reference chromosomes 01 and 04, although the presence of a translocation should generate at least two distinct discordant paired read clusters. Among potential translocation configurations, several required additional clusters of discordant paired reads contiguous to at least one extremity of the detected discordant cluster. We carefully searched for additional links involving these regions and found a cluster (64 paired reads) linking reference chromosome 01 to the extremity of reference chromosome 08 and a second cluster (104 paired reads) that linked the same region of reference chromosome 08 to reference chromosome 04 (Fig. 2c). Coverage of this chromosome 08 region appeared to be excessive (38x) relative to the average coverage on chromosome 08 (14x), thus showing its repeated nature.

Interpretation of these discordant relationships suggested the presence, in PT-BA-00267, of a reciprocal translocation in a heterozygous state involving chromosomes 01 and 04 (Fig. 2d). For convenience, we will refer to chromosomes 01 and 04 for chromosomal structures corresponding to the reference genome sequence and chromosomes 1T4 and 4T1 for chromosomal structures resulting from the reciprocal translocation compared to this reference (Fig. 2d). The presence of a segment of repeated sequences larger than the paired read insert size at the breakpoint of chromosome 4T1 could explain why we did not directly detect discordant reads between the involved regions of chromosomes 01 and 04 (Fig. 2d). This repeated sequence was also present on chromosome 08, but not on chromosomes 01 and 04 in the reference assembly. This explained why an indirect link between chromosomes 01 and 04 through chromosome 08 was detected (Fig. 2d).

PCR and BAC-FISH validation of the heterozygous reciprocal translocation

PCR primers were designed to assess the possible presence of a breakpoint on chromosomes 01, 04 and 1T4 resulting from a translocation, as suggested based on paired read mapping (Fig. 3a,b). However, chromosome 4T1 could not be tested by PCR due to the presence of repeated sequences at the translocation breakpoint, which rendered potential PCR products too long for amplification. The PCR

product sizes confirmed the presence of these three chromosome structures in PT-BA-00267. PCR product sequences confirmed that the amplified region corresponded to the expected one.

BAC-FISH experiments were performed to check the presence of a reciprocal translocation involving chromosomes 01 and 04 and thus the presence of the four chromosome structures. BAC clones from both sides of the breakpoints were selected (Supporting Information, Table S2) and used for BAC-FISH on PT-BA-00267 metaphase chromosome preparations. The results confirmed the presence of the four chromosome structures, reference chromosomes 01 and 04 and also chromosomes 1T4 and 4T1 resulting from reciprocal translocation (Fig. 3c,d and Supporting Information, Fig. S2).

Structural heterozygosity consequences on genotype representation in progeny and gamete viability

In the breakpoint regions of chromosomes 01 and 04 involved in the structural heterozygosity, marker segregation was biased in favor of an excess of one homozygous genotype at the expense of the other. PCR tests were performed on selected PT-BA-00267 self-progeny individuals (5 homozygotes for each chromosome structure and 5 heterozygotes). This showed that the underrepresented homozygous genotypes corresponded to chromosomes 01 and 04 structures, while the overrepresented homozygous genotypes in the progeny corresponded to chromosomes 1T4 and, by extension, to 4T1 structures (Supporting Information, Table S3).

Among the nine possible genotype combinations, only four were found in the most distorted regions in the PT-BA-00267 progeny using DArTseq data (Supporting Information Table S4). Three genotype combinations exhibited a higher than expected frequency in case of Mendelian segregation while a last genotype combination had a lower than expected frequency. The three genotype combinations exceeding expectations were a heterozygous genotype for both chromosome structures (37.78% vs. 25% expected) and the two genotype combinations homozygous for both chromosome structures. Among these homozygous combinations, the 1T4-1T4-4T1-4T1 chromosome combination was highly over-represented (51.67% vs. 6.25% expected) while the 01-01-04-04 chromosome combination was only slightly over-represented (8.33% vs. 6.25% expected). The sole genotype combinations found with under-representation (2.22% vs. 12.5% expected) corresponded to 01-1T4-4T1-4T1 chromosome structures.

To investigate whether these proportions were due to gametic or zygotic selection, we analyzed a second population involving PT-BA-00267 (female parent) and ‘ChicameT’ (male parent), which gave access to gamete transmission ratios. This F1 population was genotyped with 35 SSR markers located on chromosomes 01 and 04. Analysis of PT-BA-00267 alleles in this population revealed high segregation distortion and linkage involving chromosomes 01 and 04, consistent with those observed for the PT-BA-00267 self-progeny population (Supporting Information, Fig. S3). These results suggested differential

gamete viability rather than zygotic selection. Moreover, the F1 population generated information on the allelic composition of maternal gametes transmitted to the offspring. Only PT-BA-00267 maternal gametes having chromosomes 1T4 and 4T1 or gametes having the chromosomes 01 and 04 were transmitted (Supporting Information, Table S5). While gamete viability can be estimated at 100% (best transmission) for gametes with chromosomes 1T4 and 4T1, gamete viability for those with chromosomes 01 and 04 was 28%.

Paternal gamete viability was then estimated based on the PT-BA-00267 selfing population segregations (proportions of genotypes and genotype combinations between chromosomes 01 and 04) using the maternal gamete viability value calculated from the PT-BA-00267 x 'ChicameT' population. The estimated paternal gamete viability in PT-BA-00267 was close to the observed maternal gamete viability (Supporting Information, Table S5).

Distribution of chromosomes 01, 04, 1T4 and 4T1 in* Musa acuminata *germplasm

Mate-pair re-sequencing data were used to search for the presence of chromosomes 01, 04, 1T4 and 4T1 in 14 *M. acuminata* accessions by inspecting paired reads mapped on the reference sequence at the identified translocation breakpoints (Supporting Information, Fig. S4 and Supporting Information, Table S6). Out of the 14 re-sequenced accessions, eight were found to be homozygous for chromosomes 01 and 04 ('Galeo', 'Manang', 'Pisang Madu', 'Banksii', 'Calcutta4', 'DH-Pahang ', 'Bornéo' and 'Maia Oa'), five accessions were found heterozygous with a copy of each chromosome (01, 1T4, 04 and 4T1) ('IDN110', 'Pisang Lilin', 'Akondro Mainty' and 'Paka') and one accession ('Malaccensis Nain') was found to be homozygous for chromosomes 1T4 and 4T1. The triploid 'Grande Naine' accession had at least one copy of each chromosome structure.

The presence of chromosomes 01, 04 and 1T4 was also tested by PCR on 169 *M. acuminata* banana accessions (including the 14 mentioned above) (Supporting Information, Table S6). A total of 119 accessions amplified only chromosomes 01 and 04, 10 accessions amplified only chromosome 1T4, 34 amplified all tested chromosome structures (01, 04 and 1T4), one accession amplified only chromosome 04 and 1T4 and five accessions amplified only chromosome 01. The PCR results were in agreement with the paired read mapping data for the resequenced accessions, with the exception of 'IDN110' which did not amplify the chromosome 01 structure, while paired read mapping identified it as structurally heterozygous. This discrepancy may have been due to local micro-rearrangements or sequence divergence preventing PCR primer hybridization. Chromosome 4T1 could not be tested by PCR, but since nearly all of the observed gametes in progenies were 01-04 or 1T4-4T1 and since the re-sequenced accessions

bearing chromosome 1T4 also had chromosome 4T1, we hypothesize that 4T1 was present when chromosome 1T4 was detected.

The vast majority (45 out of 48) of the diploid wild accession representatives of *M. acuminata* subspecies amplified reference chromosomes 01 and 04 only (Supporting Information, Table S6). Among the three exceptions, 'Malaccensis nain' and 'Pa songkla' amplified only the 1T4 chromosome, while the PT-BA-00267 accession amplified all of the tested chromosomes (01, 04 and 1T4). Among the 73 diploid cultivated accessions tested, 48 amplified only chromosomes 01 and 04, eight amplified only chromosome 1T4 and 17 amplified chromosomes 01, 04, 1T4 (Supporting Information, Table S6).

Among the 31 triploid cultivated accessions tested, around half amplified only chromosomes 01 and 04, including the AAA accessions from the Red, Orotava and Lujujira/Mutika sub-groups, AAB accessions from the Popoulou/Maia Maoli, Mysore, Pisang Kelat and Plantain sub-groups and ABB accessions from the Bluggoe, Pelipita, Peyan and Saba sub-groups (Supporting Information, Table S6). The other half amplified chromosomes 01, 04 and 1T4, including AAA accessions from to the Ambon, Rio, Ibota, Cavendish and Gros Michel sub-groups, AAB accessions from the Silk, Nendra Padaththi and Pome sub-groups and ABB accessions from the Pisang Awak sub-group. The only tetraploid accession tested, i.e. 'Yawa2' (ABBT), also amplified chromosomes 01, 04 and 1T4.

In addition to *Musa acuminata* accessions, nine *Musa* species were tested for the various structures, including *Musa acuminata* close relatives *M. laterita*, *M. ornata*, *M. rosea* and *M. velutina*, along with more divergent species *M. sanguinea*, *M. balbisiana*, *M. maclayi* and *M. textilis*, as well as the *Fe'i* type. In all cases, 01 and 04 structures were found but not the 1T4 structure.

Finally, to refine the position in the *Musa acuminata* classification of key accessions, in particular the two wild diploid accessions homozygous for 1T4 and 4T1 chromosomes, we performed a factorial analysis with *Musa acuminata* diploid accessions for which genotyping by sequencing information was available for 3043 SNP markers. The first two axes that respectively explained 28% and 22% of the diversity allowed us to differentiate the four main *Musa acuminata* subspecies (i.e. *banksii*, *zebrina*, *malaccensis* and *siamea/burmannica*) (Fig. 4). The accession distribution along these axes showed that homozygotes for chromosome 1T4 and 4T1 were present in two distinct genetic groups corresponding to ssp. *malaccensis* and the Sucrier cultivar sub-group. Heterozygous accessions were mostly located in a triangle between *M. acuminata* ssp. *banksii*, *M. acuminata* ssp. *malaccensis* and the Sucrier cultivar subgroup.

Discussion

Characterization of a reciprocal translocation in Musa acuminata

The presence of large chromosome structural variations within *M. acuminata* was proposed by cytogeneticists on the basis of the observation of chromosome pairing irregularities at meiosis in hybrids between *M. acuminata* accessions (Dodds 1943; Dodds and Simmonds 1948; Dessauw 1987; Fauré et al. 1993a; Shepherd 1999). Seven *Musa acuminata* translocation groups, within which accessions were structurally homogeneous, were proposed by Shepherd (1999), with a supposedly ancestral Standard group (ST) and 6 groups suggested to differ by 1 to 4 translocations (Northern Malayan (NM), Malayan Highland, Northern 1, Northern 2, Javanese and East Africa). These groups only partially corresponded to the *Musa acuminata* subspecies classification. Here, for the first time, we used re-sequencing approaches and were able to characterize one of these structural variations in the PT-BA-00267 *M. acuminata* ssp. *malaccensis* accession in the form of a reciprocal translocation involving 3 Mb of the distal region of reference chromosome 01 and 10 Mb of one distal region of reference chromosome 04.

A few of the accessions studied by Shepherd (1999) were included in our study. Among them, we characterized 'P. Lilin' and 'Paka' accessions as structurally heterozygous with chromosomes 01, 04, 1T4 and 4T1 while 'Pahang', 'Selangor', 'Madang' and 'Borneo' accessions were characterized as homozygous for chromosomes 01 and 04. The first two accessions belonged to the NM translocation group and the last four to the ST group. These two groups were proposed to differ by one translocation event (Shepherd 1999) and our results revealed that this event was a reciprocal translocation involving reference chromosomes 01 and 04.

Molecular marker segregation analysis in an F1 population involving 'P. Lilin' as male parent revealed distorted segregation and, based on the pattern of these distortions, Hippolyte et al. (2010) suggested the presence of a duplication of the chromosome 01 distal region in chromosome 04. In the present study, thanks to the availability of a reference genome sequence for *Musa acuminata*, we were able to reinterpret the data and refute the duplication hypothesis but showed that 'P. Lilin' presented a heterozygous reciprocal translocation of distal regions of chromosomes 01 and 04, identical to that identified in PT-BA-00267.

Impact of NM translocation on chromosomal segregation

SNP segregation in PT-BA-00267 self-progeny showed that the reciprocal translocation of distal regions of chromosomes 01 and 04 at a heterozygous state highly distorted chromosome segregation. It also induced a reduction of recombination in regions around breakpoints and generated genetic linkage between the reference chromosome 01 and 04 regions involved. Less expected, we observed no recombination in the 3 Mb translocated region of chromosome 01. In his review, and based on the

observation of an anaphase bridge and a minute chromosome fragment, Shepherd (1999) proposed the presence of a small sub-terminal inversion in one of the translocated fragments between the ST and NM groups (Supporting Information, Fig. S5). This inversion could explain the lack of crossover in this region. Careful searches for inversion between chromosomes 1 and 4T1 in this region in the PT-BA-00267 accession using re-sequencing data did not reveal such a structure. However, if this inversion is terminal and due to the fact that telomeric regions are missing in the assembly such evidence may not yet be detectable with paired read data. Finally, the relatively small size of the translocated fragment (3 Mb, 3-fold smaller than for chromosome 04) combined with the acrocentric nature of chromosome 01 could explain this lack of recombination.

The consequences of this structural heterozygosity on gamete transmission to PT-BA-00267 progeny were found to be similar in paternal and maternal gametes and resulted in the quasi-absence of gametes displaying a combination of Standard and Northern Malayan groups. The lethality of such gametes could be explained by the lack of either of the translocated regions of chromosomes 01 or 04 in these cells. This observation is in agreement with the report on STxNM hybrids by Shepherd (1999), based on chromosome pairing analysis in hybrid progenies.

The observation of differential gamete transmission of the two remaining gamete types with a biased proportion in favor of increased transmission of chromosomes 1T4 and 4T1 compared to chromosomes 01 and 04 was more intriguing. Such bias has also been reported in STxNM hybrids by Dodds and Simmonds (1948), reviewed in Shepherd (1999), based on chromosome pairing observations. Interestingly, in these hybrids, the authors observed a ratio of 57% of pollen mother cells (PMC) displaying 11 bivalents and 39% PMC displaying 9 bivalents, 1 V-shaped trivalent and 1 univalent. Based on these proportions and on the hypothesis that unbalanced gametes are not viable, among 100 PMC, 57 PMC should generate 57 gametes with an ST structure and 57 gametes with an NM structure (Fig. 5). Among the 100 PMCs, 39 showed a tetravalent V-shape and a univalent. Based on the absence of recombination in the translocated region of chromosome 01 associated with its acrocentric nature and the hypothesis of no more than one chiasma per chromosome arm, chromosome 01 was strongly favored as being the univalent observed by Shepherd (1999)(Fig. 5). Note that Shepherd (1999) reached the same conclusion on the nature of the univalent based on the fact that chromosome bridges and univalents were mutually exclusive events. Therefore, the trivalent should involve the two NM chromosomes (1T4 and 4T1) separated by chromosome 04. Based on the V-shape, it could be assumed that during anaphase 1 the two NM chromosomes segregated to the same pole and chromosome 04 segregated to the other. Regarding the fate of the univalent (chromosome 01), it could be expected that it segregated randomly (half with chromosome 04 and half with NM chromosomes), leading to an equal proportion of ST and NM gametes plus unbalanced gametes. However, a few studies have suggested that univalents could be

transmitted in a lower proportion than expected. In *Brassica* addition lines, supernumerary chromosomes (which formed univalent at meiosis) were found at a rate of 1.3 to 30% in gametes, while 50% was expected (Chèvre et al. 1997). Similar results have also been found in wheat addition lines with an average transmission ratio of 25% (Morrison 1953). In the extreme case of no univalent transmission to gametes, the 39 PMCs will give 78 NM gametes and 78 non-viable unbalanced gametes with only chromosome 04. In this context, a total of 57 (30%) ST and 137 (70%) NM gametes would be obtained from 100 PMCs (Fig. 5). Interestingly, these proportions corresponded to that we deduced from DArTseq analysis in PT-BA-00267 self-progeny. This meiosis mechanistic hypothesis could explain the observed segregation. However, other mechanisms involving incompatible gene combinations in hybrids such as those reviewed in Maheshwari and Barbash (2011), Larracuente and Presgraves (2012), Sweigart and Willis (2012) cannot be excluded.

***The Northern Malayan translocation may have emerged in* M. acuminata *ssp*. malaccensis**

All wild diploid *M. acuminata* accessions tested, except three *M. a. malaccensis* accessions, showed the ST structure only. In addition, the ST structure was found in all tested *Musa* species, suggesting that this structure occurred in the *M. acuminata* lineage after its divergence from the other *Musa* species. The NM structure at the homozygous state was only found in a few *M. acuminata* ssp. *malaccensis* wild accessions and in the Sucrier cultivar sub-group. The Sucrier sub-group is composed of diploid AA clones with very low fertility, thus reducing the likelihood that this new structure emerge in the Sucrier sub-group and then spread within *Musa*. Therefore, it is likely that this structure emerged in spp. *malaccensis*, the only other group showing the NM structure in a homozygous state. However, due to its rare occurrence within the *M. a.* spp. *malaccensis* sample tested here, it cannot be completely excluded that this structure could have been present in an untested *Musa* species and was then incorporated into *M. a.* ssp. *malaccensis*. Further phylogenetic analyses are needed to confirm its origin in ssp. *malaccensis.*

Interestingly, the NM structure was found to be over-represented in the heterozygous PT-BA-00267 self-population (70% of haplotypes) as well as in other populations from heterozygous parents (Shepherd 1999). These results suggested that the new emerging structure may progressively replace the ST structure. However, in natural populations, the distinct fertility pattern of homozygotes compared to heterozygotes for these chromosome structures may affect spreading of the new structure.

The Northern Malayan translocation may have favored the emergence of triploid cultivars

The NM structure was found in 39% of the tested banana cultivars (seedless parthenocarpic accessions), mainly in a heterozygous state. Interestingly, this structure was found to be scattered among diverse cultivar groups. These results suggest that the new NM structure could have been incorporated via hybridization and backcrosses in distinct genetic backgrounds. This would imply that there were more hybridization steps in the formation of current cultivars than currently assumed (Perrier et al. 2011), as was already suggested by De Langhe et al. (2010).

The diversity pattern of diploid cultivars with structural heterozygosity in the factorial analysis was interesting as most of them were located between *M. a.* ssp. *malaccensis*, Sucrier and *M. a*. sp. *banksii*, likely as a result of secondary hybridization between these gene pools. This pattern is consistent with the hypothesis of separate South-East Asia and New Guinea domestication events proposed by Sardos et al. (2016).

The NM structure was found in half of the triploid cultivar sub-groups tested, all of them being of the dessert type, highlighting a substantial contribution of this new chromosome structure to polyploid cultivars, while suggesting a role of this structure in dessert banana domestication. In particular, this structure was found in the Cavendish sub-group of dessert bananas, which represents above half of global banana production and also in its suggested 2n gamete donors belonging to the Mlali sub-group (Raboin et al. 2005; Perrier et al. 2009). The reasons underlying the success of this spreading remain to be clarified. The hypotheses include over-representation of the NM structure in gametes from STxNM hybrids, reduced fertility of heterozygous genotypes (important for fruit edibility) or some other important agronomical traits associated with this structure that have been selected by farmers. In addition, as structural heterozygosity perturbs meiosis, it could have favored the production of unreduced gametes (Ramsey and Schemske 1998) and may thus have been an important factor in the formation of triploid cultivars and thus in banana domestication. This hypothesis remains to be tested on larger progenies since no triploids were observed in the selfed PT-BA-00267 progeny analyzed (D'Hont et al. 2012). Triploidy is the most efficient ploidy level for agronomic performance in banana (Bakry et al. 2009). These characteristics have generated more vigorous plants, larger fruits and higher sterility, resulting in a complete absence of seeds in the fruits.

The presence of NM structures in the heterozygous state in accessions used in breeding programs could have important breeding implications. This knowledge could be exploited to either foster recombination or fix allele combinations in the rearranged regions by choosing adequate parental combinations**.** This applies also to genetic studies involving QTLs or GWAS, etc., aimed at identifying chromosome regions involved in agronomic traits since recombination reduction and biased gamete transmission would hamper and reduce the resolution in the vicinity of rearranged regions.

Finally, *Musa* with its combination of vegetative propagation with occasional sexual reproduction, is likely to display ongoing slow-motion genetic differentiation and represents a valuable model for unravelling various facets of the speciation process in plants.

Material and Methods

Plant Material

A total of 169 *Musa* accessions, wild and cultivated, representative of known *Musa acuminata* diversity, were analyzed (see Supporting Information Table S6).

In addition, two populations, *i.e.* a self-progeny population of 180 PT-BA-00267 diploid individuals (D'Hont et al. 2012, Supplementary Information) and a biparental progeny population of 57 triploid individuals from a cross between PT-BA-00267 and a tetraploid derived from chromosome doubling of the parthenocarpic AA cultivar 'Chicame' (PT-BA-00056; 'ChicameT') obtained at the CIRAD research station in Guadeloupe were also analyzed.

Methods

DArTseq genotyping, segregation distortion analysis and recombination rate estimation

PT-BA-00267 self-progeny was genotyped using the DArTseq technology (Cruz et al. 2013) as described in Martin et al. (2016). Only co-dominant DArTseq markers were used for this analysis. DArTseq markers were filtered to reduce the technical genotyping error rate using a similar approach to that described in Spindel et al. (2013) for genotyping by sequencing data. This step was based on the assumption that: i) recombination could not occur several times in a small window of contiguous markers, ii) only markers that could be located on the 11 *M. acuminata* pseudomolecules were preserved, and iii) markers accounting for multiple recombination breakpoints in more than 10% of the progeny were removed. The filtration step was automatically performed with *locOnRef* and *GBS_corrector* tools available on the South Green platform https://github.com/SouthGreenPlatform in the Scaffremodler toolbox (Martin et al. 2016).

Pairwise marker linkage LOD was calculated using JoinMap4.1 and represented along the chromosomes using the *pwd2figure* tool in the Scaffhunter toolbox (Martin et al. 2016) available on the South Green platform https://github.com/SouthGreenPlatform.

Recombination rates were estimated on sliding windows of 500 kb along the 11 chromosomes. Genotyping error should be close to 0% when calculating recombination rates because genotyping errors lead to artefactual recombination breakpoints. Genotyping data were thus corrected based on the same principle applied to marker filtering (*i.e.* if the genotype of an individual for a marker differed from the

strict consensus of the 6 surrounding markers, the genotype of this individual was converted to the consensus genotype).

Mate-pair sequencing and structural variation detection

5 kb insert mate-pair libraries were constructed for 15 *Musa* accessions. Libraries were sequenced using the Illumina Hiseq platform at GENOSCOPE http://www.genoscope.cns.fr and BGI http://www.genomics.cn/en.

Paired reads from the 15 sequenced accessions were aligned against the *Musa acuminata* DH Pahang reference genome sequence (D'Hont et al. 2012) using bowtie2 in very-sensitive mode. Only single hit paired reads were conserved, and redundant paired reads were removed using the MarkDuplicates tool of the Picard toolkit (http://broadinstitute.github.io/picard/). Filtered paired reads were then used to identify discordant read clusters. Re-sequencing data from the reference genome accession 'DH Pahang', was also used. Discordant read clusters detected using paired reads from 'DH Pahang', were considered as resulting from assembly errors and were thus removed in the analysis of other accessions. Discordant reads were searched and interpreted using the Scaffremodler tools previously developed and available on the South Green platform https://github.com/SouthGreenPlatform in the Scaffremodler toolbox (Martin et al. 2016). Discordant read clusters were visualized using CIRCOS software (Krzywinski et al. 2009).

Targeted-PCR validation

Primer pairs were designed at the boundaries of the identified rearrangement breakpoint in the PT-BA-00267 accession. Primers 1LF (5'-TGGAGTTGGCCTGTTAAACC-3') and 1LR (5'-ACTTGCCGTTTGAACCATC-3') on chromosome 1 and 4LF (5'-TGGTGAAAGCATTATCTCTTGG-3') and 4LR (5'-AGACGCAGCATTTGGATG-3') on chromosome 4 were used to validate the reference genome structure. Primers 1T4F (5'-CGCACTTGGAGCTTGTTCTT-3') and 1T4R (5'-AACTTGCCGTTTGAACCATC-3') were used to validate the alternative structure 1T4. The alternative 4T1 structure could not be tested by PCR due to the presence of repeated sequences at the translocation breakpoint, which rendered potential PCR products too long for amplification.

BAC-FISH validation

Chromosome preparations were performed as described in D'Hont et al. (2000). Seven BAC clones (MAMB_34N11, MAMB_17B03, MAMB_51M04, MAMH_47D06, MAMB_01M16, MAMB_51J24 and MAMH_66D03) from both sides of the breakpoints were selected from a BamH1 and HindIII BAC libraries of accession DH-Pahang (D'Hont et al. 2012); http://banana-genome.cirad.fr/).

BAC clones were labelled by random priming with biotin-14-dUTP (Invitrogen, Life Technologies) or Alexa 488-5-dUTP (Invitrogen, Life Technologies). *In situ* hybridization was performed as described in D'Hont et al. (1996) with the following modifications. Chromosome preparations were incubated in RNAse A (100 ng/µL), pepsin (100 mg/ml) in 0.01M HCl and fixed with paraformaldehyde (4%). Biotinylated probes were immunodetected by Texas Red avidin DCS (Vector Laboratories) and the signal was amplified with biotinylated anti-avidin D (Vector Laboratories). Fluorescence images were captured using a CoolSnap HQ camera (Photometrics, Tucson, Ariz) via an Axioplan 2 microscope (Zeiss, Oberkochen, Germany) and analyzed using MetaVueTM (Universal Imaging Corporation, Downington, PA).

Estimation of PT-BA-00267 maternal and paternal gamete transmission ratios

PT-BA-00267 x 'ChicameT' biparental progeny was genotyped with 35 SSR markers located along chromosomes 01 and 04. SSR genotyping was performed with the Applied Biosystems® 35006L Genetic Analyzer. PT-BA-00267 maternal gamete transmission ratios were then estimated in the most distorted region of chromosomes 01 and 04.

PT-BA-00267 selfing progeny could not directly give access to the gamete transmission ratios. However, for this type of cross, the expected genotype combination proportions for the two chromosomal structures in case of a reciprocal translocation can be expressed as a function of parental gamete transmission ratios for the 01 and 04 chromosome regions around the translocation breakpoints (Supporting Information, Fig. S6). As maternal gametes can be fixed using the biparental cross, paternal gamete transmission ratios were estimated by searching quadruplets (a value for each chromosome 01 and 04 combination) complying with both observed genotype proportions and genotype combination proportions in the population. A quadruplet was kept if, for each genotype proportion, the deviation from the observed values was less than 0.02. A total of 10000 quadruplets were searched and the most probable quadruplets were then identified based on their distribution and mean value (Supporting Information, Fig. S7).

Diversity analysis and genotyping by sequencing

To investigate the occurrence of the two chromosomal structures in a panel of *Musa* diversity, PCR amplification was performed on 169 *Musa* accessions using the 1LF/1LR, 4LF/4LR and 1T4F/1T4R PCR primers pairs (Supporting Information, Table S6).

For a part of the studied accessions, Illumina sequencing data (RNAseq and DNAseq) were available through various ongoing projects (Supporting Information, Table S6). RNAseq and DNAseq reads were aligned against version 2 of the 'DH Pahang' *Musa acuminata* reference genome sequence

(Martin et al. 2016) using STAR (Dobin et al. 2012) and BWA (Li and Durbin 2010) respectively. Reads were locally realigned around indels using the IndelRealigner tool of GATK software, version 3.3 (McKenna et al. 2010). For each accession, at each covered position, all mapping bases that had a mapping quality equal to or greater than 10 were counted with the bam-readcount program (https://github.com/genome/bam-readcount). For each accession and at each variant site, a genotype was called based on the maximum likelihood of the genotype, calculated based on a binomial distribution assuming a sequencing error rate of 0.005. The variant calling file was formatted in VCF format. The VCF file was then filtered according to the following criteria: i) data points covered by less than 10 reads were converted to missing data, ii) data points with a minor allele frequency inferior to 3 reads were converted to missing data. Accessions and sites (available at http://banana-genome-hub.southgreen.fr/download) were then selected to have no more than 50% missing sites per accession. The final VCF file, composed of 3043 polymorphous sites for 75 *Musa acuminata* wild and cultivar accessions, was used to calculate a dissimilarity matrix using custom python scripts. The dissimilarity index between two accessions was calculated as the proportion of unmatching alleles. The dissimilarities matrix was used to perform a factorial analysis using R (v3.2.4) software (http://www.r-project.org). Considering that cultivar accessions originated from the wild banana genepools, the factorial analysis was performed with the 35 wild accessions. The 40 cultivar accessions were then projected along the synthetic axes.

Acknowledgements and funding information

This work was supported by the Centre de cooperation Internationale en Recherche Agronomique pour le Développement (CIRAD). The authors thank the CGIAR Research Program on Roots, Tubers and Bananas (RTB) and Agropolis Fondation (ARCAD project) for financial support for data acquisition. We thank the South Green Bioinformatics Platform (http://www.southgreen.fr) for providing us with computational resources. We thank Christophe Jenny (CIRAD research station, French West Indies) for providing the PT-BA-00267 self-progeny population, Lionel Toubi (CRB Plantes tropicales, French West Indies) for providing roots and leaves samples and Jeff Daniells (Department of Agriculture, Fisheries and Forestery, Australia) for providing some leaf samples.

Availability of supporting data

Illumina 5 kb reads mapping in the breakpoint regions of chromosomes 01, 04 and 08, the VCF file comprising 3043 high-quality polymorphic sites for 75 diploid *Musa acuminata* accessions, the PT-BA-00267 selfing population genotyping matrix and the PT-BA-00267 x ‘ChicameT’ genotyping matrix are available in the download section of the Banana Genome Hub (http://banana-genome-

hub.southgreen.fr/download) under *Transloc_1-4_reads.tar.gz*, *Transloc_1-4_vcf.tar.gz*, *AF-Pahang marker matrix file* and *PT-BA-00267_x_ChicameT.txt* names, respectively.

Authors' contributions

G.M., F.C., F.C.B. and A.D.H. designed the study

G.M. performed bioinformatics and genetic analyses

P.D. designed some bioinformatics tools

O.C performed BAC FISH analyses

F.S and D.R. provided the genetic material

C.H. and C.C. performed PCR analyses

K.L., J.S. and M.R. produced or provided part of the sequencing data

F.C. and F.C.B. contributed to the analysis and edited the manuscript

G.M. and A.D.H wrote the manuscript

ADH: coordinated the study.

Figure legends

Fig. 1 Representation of marker linkage, recombination rates and segregation distortion in PT-BA-00267 self-progeny along the 11 *Musa acuminata* chromosomes. Each dot represents linkage between two markers. Marker linkage is represented by a color gradient from red to dark blue for strong and weak linkages, respectively. The black curve represents marker segregation distortions calculated as $-\log_{10}$ (p-value of the chi-square test testing the deviation of the expected Mendelian segregation ratio). The red curve represents the recombination rate.

Fig. 2 Paired read evidence for a reciprocal translocation involving chromosomes 01 and 04. (**a**) Circos representation of significant discordant read clusters from PT-BA-00267 identified in the targeted regions of reference chromosomes 01 (0 to 8.5 Mb) and 04 (23.5 to 29.5 Mb). (**b**) Circos with focus on paired reads in a 25 kb region around the discordant read cluster detected in chromosomes 01 and 04 in (**a**). Grey lines correspond to concordant pairs (correct orientation and insert size), orange and red lines correspond to discordant pairs with smaller and greater insert sizes, respectively. Purple lines correspond to pairs showing a reverse-reverse orientation, green lines a forward-forward orientation, and blue lines correspond to pairs with a complete reverse orientation relative to the paired library construction. (**c**) Circos with focus on paired read clusters detected in the targeted region of chromosomes 01 and 04 and with chromosome 08. (**d**) Hypothesized chromosome structures for PT-BA-00267 based on the paired read mapping interpretation. Linked colored arrows correspond to the read pairs shown in Figure 2C. Centromeres are indicated by circles.

Fig. 3 Validation of PT-BA-00267 structural heterozygosity through PCR and BAC-FISH. **(a)** PCR amplification of breakpoints using primers located along the reference and hypothesized chromosome structures **(b)**. **(c)** BAC-FISH on a PT-BA-00267 chromosome preparation using BACs MAMB_34N11 (red) and MAMB_51M04+MAMH_47D06+MAMB_01M16 (green). **(d)** Location of BACs along reference and hypothesized chromosome structures.

Fig. 4 Factorial analysis performed on 35 wild *Musa acuminata* accessions with projection of 40 cultivars along the synthetic axes. The dissimilarity matrix was based on genotyping by sequencing data. Pink dots indicate accessions homozygous for chromosomes 1T4 and 4T1, black dots indicate accessions homozygous for chromosomes 01 and 04 and purple dots indicate structurally heterozygote accessions.

Fig. 5 Schematic representation of chromosomal pairing within a heterozygous accession for chromosomes 01, 04, 1T4 and 4TI (ST x NM hybrids) and the hypothesis that led to the observed gamete frequencies.

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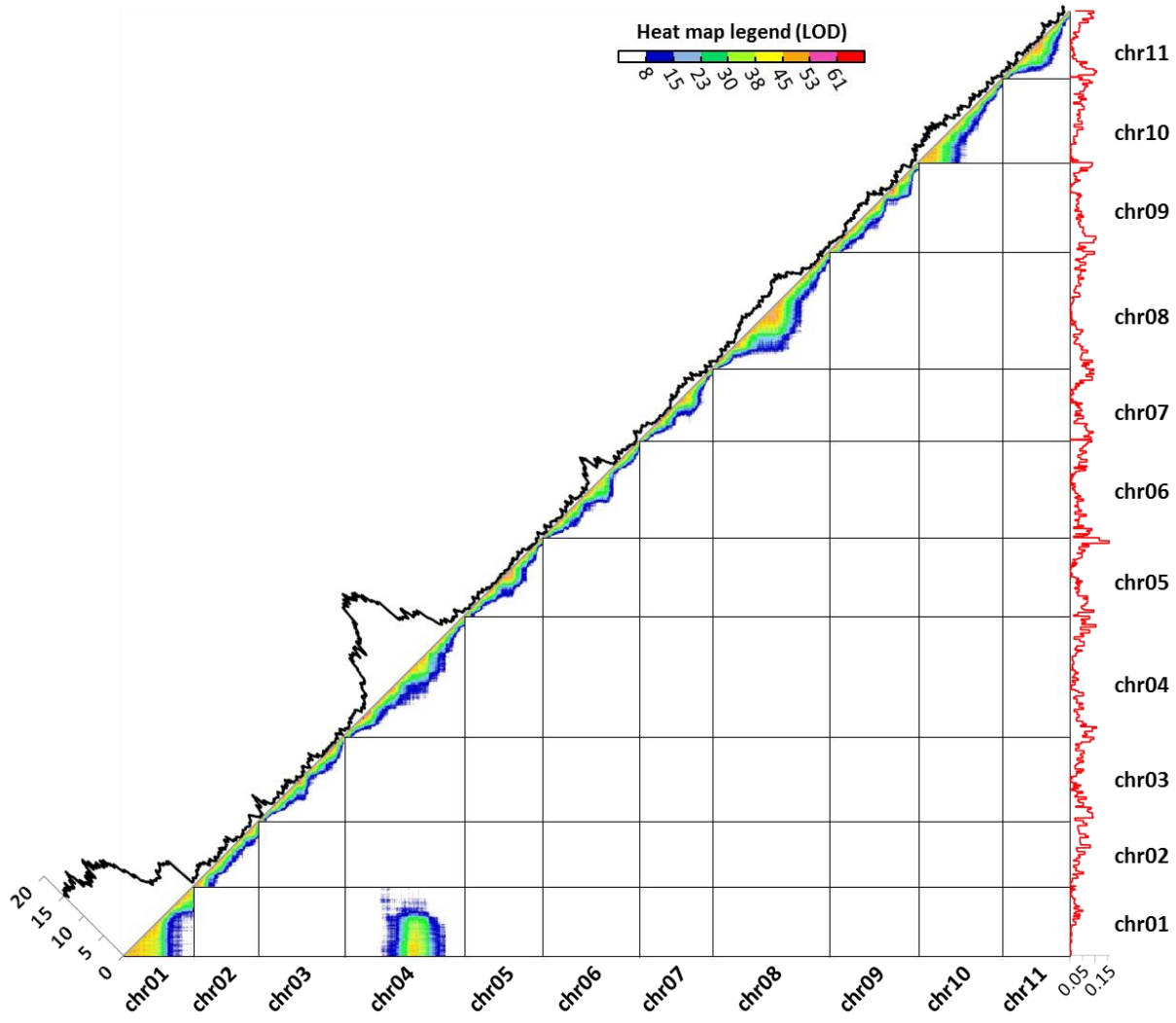
Figures

Fig. 1



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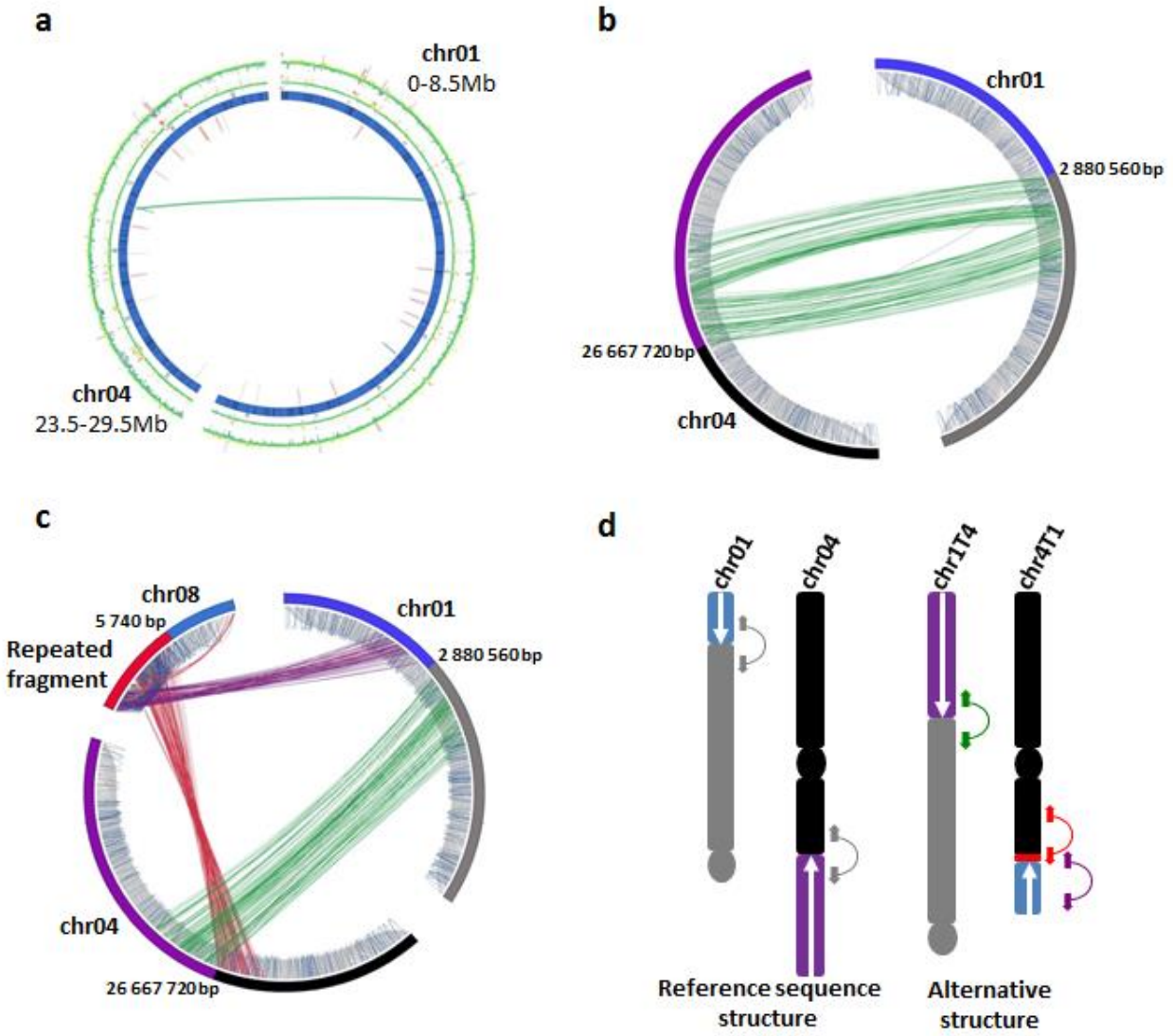

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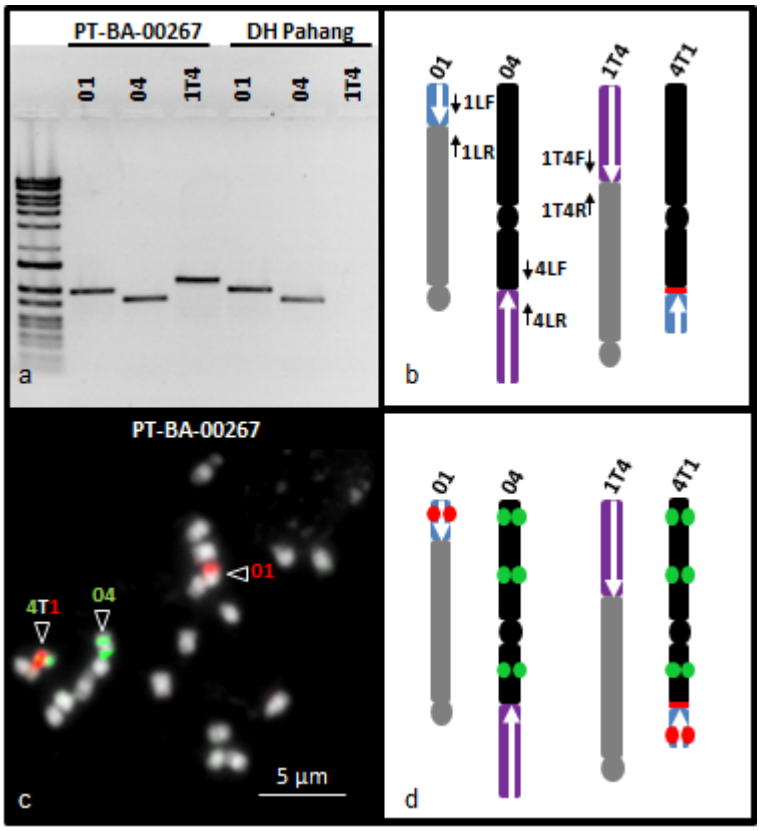

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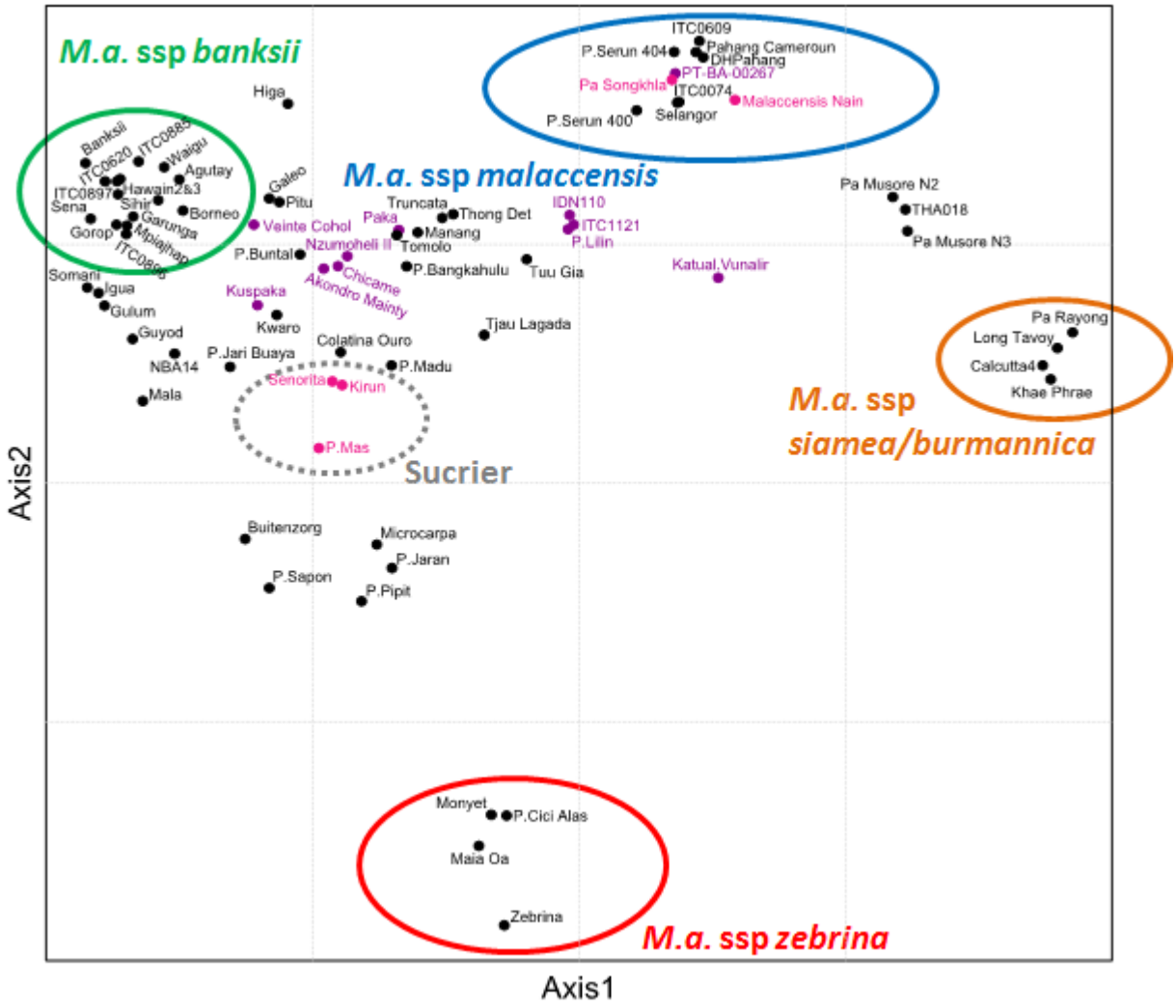

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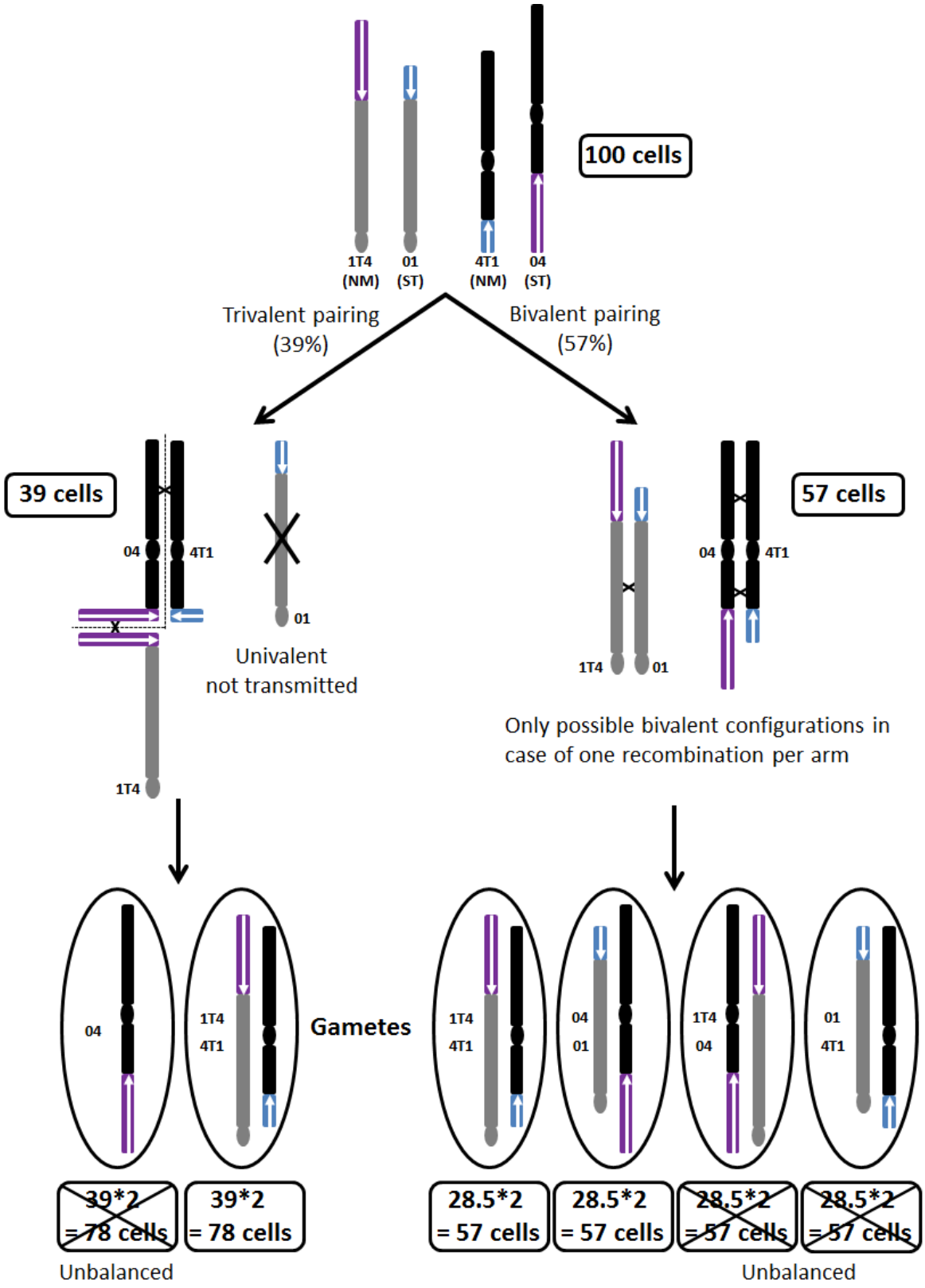
100 cells
1T4 (NM)
01 (ST)
4T1 (NM)
04 (ST)
Trivalent pairing (39%)
Bivalent pairing (57%)
39 cells
57 cells
04
4T1
01
Univalent not transmitted
1T4
Only possible bivalent configurations in case of one recombination per arm
Gametes
1T4 4T1
04 01
01 4T1
39*2 = 78 cells
28.5*2 = 57 cells
Unbalanced

717