

## First report of saffron-associated mastrevirus 1 from saffron in Iran

**SeyyedeH Atefeh Hosseini\***, Department of Plant Protection, Faculty of Agriculture, University of Birjand, Iran; and **Charlotte Julian, Serge Galzi, Denis Filloux and Philippe Roumagnac\***, PHIM Plant Health Institute, Univ. Montpellier, CIRAD, INRAE, Institut Agro, IRD, Montpellier, France.

[ahosseini@birjand.ac.ir](mailto:ahosseini@birjand.ac.ir)

[philippe.roumagnac@cirad.fr](mailto:philippe.roumagnac@cirad.fr)

In spring 2022, 40 leaf samples of saffron plants harboring a wide variety of symptoms, including curling, yellowing, mosaic, dwarfing and leaf malformation were collected from three Khorasan provinces in Iran. These samples were processed using the virion-associated nucleic acid-based metagenomics approach (Moubset et al., 2022). Noteworthy, 147 contigs with a size >500 bp distributed among 35 samples with multiple symptomatic patterns (curling, yellowing, mosaic, dwarfing and leaf malformation) shared >90% nucleotide identity with saffron-associated mastrevirus 1 (SaM1) (Martínez-Fajardo et al., 2024). This virus has recently been detected from transcriptomic datasets from saffron (*Crocus sativus* L.) collected in India. SaM1 was proposed to belong to a new species of the *Mastrevirus* genus (*Geminiviridae* family) and was tentatively named *Saffron-associated mastrevirus 1* (Martínez-Fajardo et al., 2024). In addition, contigs assigned to saffron potyviruses, including saffron latent virus, turnip mosaic virus and bean yellow mosaic virus were assembled from the 40 saffron samples pinpointing that no conclusion could be made on the causal virus of observed symptoms. Total DNAs of the 40 saffron samples were further extracted using the DNeasy Plant Mini Kit (Qiagen) and were tested for the presence of SaM1 using a primer pair amplifying a 327 bp long fragment located in the coat protein of SaM1 (CSAV\_2F 5'-

TTTAAGTCAGGGTCTGGAGATG-3' and CSAV\_3R 5'-

GGCATACTGTAACCTCGTCTTC). Amplification conditions were: an initial denaturation at 95°C for 10 min, 30 cycles at 95°C for 30 sec, 55°C for 30 sec, 72°C for 45 sec, and a final extension step at 72°C for 10 min. This PCR test confirmed that 28/40 samples tested positive for SaM1. Total DNAs from one sample (#69-32) testing positive for SaM1 was further amplified using Phi29 DNA polymerase (TempliPhi, GE Healthcare) by rolling circle amplification (RCA) as previously described (Shepherd et al., 2008). RCA products were used as a template for PCR amplification of the complete genome of SaM1 using abutting primers (CSAV\_PST1F 5'-CTGCAGTTGCGGTAAGTCTATGTTGGCTG-3' and CSAV\_PST1R 5'-CTGCAGAGACAACGATTCCCAAATTACTTTGTTCC-3').

Amplification conditions were: an initial denaturation at 95°C for 10 min, 30 cycles at 95°C for 30 sec, 55°C for 1 min, 72°C for 2 min and 30 sec, and a final extension step at 72°C for 10 min. Amplification products of approximately 2.7 Kbp were gel purified, ligated to pGEM-T Easy (Promega) and sequenced by standard Sanger sequencing using a primer walking approach (Azenta, Germany). The arrangement of open reading frames within the 2,724 nt circular DNA of the CSAV\_A9PstI clone (accession number PQ392009) is similar to those reported for mastreviruses, including the coat protein (CP), the movement protein (MP), the replication-associated protein (Rep) and the RepA protein. In addition, CSAV\_A9PstI sequence shares 95.3% genome-wide identity with SaM1 consensus sequence assembled from transcriptomics data (Martínez-Fajardo et al., 2024). Overall, these results indicate that SaM1 is widely present in saffron in different regions of Northeast Iran. In addition, while all tested plant were infected by several viruses, the etiology of SaM1 remains to be determined precisely. To our knowledge, this is the first report of saffron-associated mastrevirus 1 from saffron in Iran.

Acknowledgements

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#### References

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Photos of saffron plants collected in Iran showing symptoms of leaf curling. It is important to note that these plants are infected by both a geminivirus and a potyvirus, which means that it is not possible to conclude whether one or both viruses are responsible for the symptoms.

308x669mm (118 x 118 DPI)